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1 **Predicting the risk of non-target damage to a close relative**
2 **of a target weed using sequential no-choice tests, paired-**
3 **choice tests and olfactory discrimination experiments**

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9 **Abstract**

10 We investigated host plant utilisation by the candidate biocontrol agent *Paradibolia*
11 *coerulea* (Coleoptera: Chrysomelidae) on the target plant *Spathodea campanulata*
12 Beauv. (Bignoniaceae) and a closely related non-target plant, *Kigelia africana* (Lam.)
13 Benth. (Bignoniaceae). Paired-choice and sequential no-choice experiments were
14 performed and coupled with olfactory discrimination experiments to test the insects'
15 responses to volatiles from both plant species as well as to cues from conspecific
16 beetles. Although *K. africana* was utilised by *P. coerulea*, *S. campanulata* was
17 preferred for both adult feeding and oviposition. Interestingly, whereas females were
18 attracted to olfactory cues emitted by *S. campanulata*, males demonstrated no such
19 olfactory discrimination. Females were also attracted to cues deposited by males, and
20 males were deterred by cues from other males, but neither sexes responded to female
21 olfactory cues. Very few eggs were recorded on *K. africana* and none of the larvae
22 that hatched on *K. africana* survived the first instar. Both *S. campanulata* and *K.*
23 *africana* are suitable for adult feeding, but persistent utilisation of *K. africana* in the
24 field is unlikely because larval development is only possible on *S. campanulata* and
25 because the adult females are strongly attracted to volatiles emitted by the target plant.

26 Nevertheless, if *P. coerulea* is released as a biocontrol agent, spill-over adult feeding
27 could potentially occur on *K. africana* growing sympatrically with *S. campanulata*.
28 Because *P. coerulea* cannot complete its development on *K. africana*, non-target
29 damage will only occur where the target plant is present, with an intensity dependent
30 on densities of adult beetles locally.

31

32 **Keywords:** *Paradibolia coerulea*; *Spathodea campanulata*; *Kigelia africana*; weed
33 biological control; host specificity testing; host plant selection

34

35 1. INTRODUCTION

36 *Spathodea campanulata* Beauv. (Bignoniaceae) (African tulip tree) is a problematic
37 alien invasive species in many tropical and subtropical parts of the world (Larrue et
38 al., 2014). It is native to Central and West Africa (Bidgood, 1994), but widespread
39 cultivation has led to naturalization in areas outside of the native range (Francis, 1990).
40 It is considered one of the most damaging invasive alien species in the world due to
41 the negative impact of infestations on native biodiversity and agricultural productivity
42 (Lowe et al., 2000; Labrada & Medina, 2009; Larrue et al., 2014). *Spathodea*
43 *campanulata* is considered a suitable target for biological control in the Pacific region
44 (Paynter, 2010) and biological control options are being investigated (Paterson et al.,
45 In Press). One of the potential candidate biological control agents is a flea-beetle,
46 *Paradibolia coerulea* Bryant (Chrysomelidae: Coleoptera), which feeds externally on
47 the leaves of the plant as an adult and mines between the epidermal layers of leaves
48 as a larva (Paterson et al., In Press).

49 *Spathodea campanulata* belongs to the family Bignoniaceae, which contains many
50 species of ornamental and cultural importance (Olmstead et al., 2009). It is part of the

51 paleotropical clade of Bignoniaceae which has an African and Asian distribution
52 (Olmstead et al., 2009). The closest relatives to the target weed are considered to be
53 other African Bignoniaceae species including the African sausage tree, *Kigelia*
54 *africana* (Lam.) Benth. (Olmstead et al., 2009). Although *K. africana* is not indigenous
55 to the Pacific region, where *S. campanulata* is most problematic, it is present as a
56 horticultural plant in some parts of the region. *K. africana* is also indigenous to South
57 Africa, where *S. campanulata* has been declared an invasive alien plant (Department
58 of Environmental Affairs, 2014).

59 *Spathodea campanulata* is a large tree of up to about 30m in height, has compound
60 leaves with four to eight pairs of opposite leaflets and a terminal leaflet. Each leaflet is
61 approximately 10cm long and 5cm in width. The flowers are large, bright red and bell-
62 shaped; and the fruits are cigar-shaped woody pods which contain winged seeds
63 (Hedberg et al., 2006). *Kigelia africana* is also a large tree, reaching 25m in height with
64 compound leaves containing six to ten leaflets of similar dimensions to those of *S.*
65 *campanulata* (Coates-Palgrave, 2002). The flowers of *K. africana* are also similar to
66 those of *S. campanulata*, but are a deeper red and are found on long flexible stems
67 that hang down from the mature woody branches. *Kigelia africana* has distinctive large
68 sausage-shaped woody fruits of up to 1m in length and weighing up to 10kg (Coates-
69 Palgrave, 2002).

70 Close relatives of target weeds are the most likely species to be fed on as
71 alternative host plants by candidate control agents and it is now standard practice to
72 select test plants on the basis of phylogenetic separation from the target weed, starting
73 with the closest relative and incorporating more distantly-related taxa until the host-
74 range is circumscribed (Briese, 2005). Host range has almost exclusively been
75 determined by employing test designs that incorporate (i) no-choice, (ii) choice and

76 (iii) open-field (multi-choice) trials (Schaffner, 2001; Moffat et al., 2013). The no-choice
77 test applies a single biological control agent with a single test plant species under
78 standard conditions (Withers & Mansfield, 2005). Over an extended experimental
79 duration, no-choice tests may overestimate the fundamental host range of a candidate
80 agent, because over time there is increased acceptance of hosts due to starvation and
81 experience (Withers, Barton-Browne & Stanley, 1999). Choice tests, in which the
82 potential agent is exposed to two or more test species that include the target plant,
83 provide an indication of preference by the insect (Buckingham, Okrah & Christian-
84 Meier, 1991; Edwards, 1999). In order to interpret the results of choice tests it is
85 important that the mechanism of host plant selection be considered. For example, an
86 insect may be stimulated to oviposit on a test plant that it would never utilise under
87 field conditions if plant volatile cues from the target weed have accumulated around
88 the non-target test plant. Multi-choice tests under open field conditions are therefore
89 a more reliable method of determining the realised host range of potential agents, but
90 these tests are not always possible, because agents must usually be kept under
91 quarantine conditions. Under field conditions, host plant selection will be determined
92 by a number of cues that may not be present, or may be distorted, under the artificial
93 conditions simulated in quarantine, so an understanding of how a potential agent uses
94 cues to select a host plant should improve interpretation of the results of host
95 specificity testing (Marohasy, 1998).

96 The sequential no-choice test is an infrequently-utilised test design which may
97 reduce the probability of obtaining false (positive or negative) results compared with
98 standard host specificity tests (Withers, Barton-Browne & Stanley, 2000). This test
99 design effectively combines several no-choice tests into a single experiment, whereby
100 the candidate biological control agent is offered a series of test plant species,

101 alternating between the target species and an additional test species (Fig. 1). The
102 value of the sequential no-choice test is that volatiles emitted by the target plant are
103 less likely to confound the utilization of other test plants, that behavioural aspects of
104 host-selection (such as learning and experience) are incorporated into the test design
105 by exposing the herbivore to test plants on multiple occasions, and because time-
106 dependent influences on host-acceptance and utilization are accounted for (Withers,
107 Barton-Browne & Stanley, 2000).

108 Host plant selection is a vital element in the survival of most herbivorous insects,
109 but species with less mobile immature stages, such as endophagous beetle larvae,
110 are particularly reliant on appropriate decisions being made by ovipositing females
111 that select plants offering suitable requirements for larval development (Bernardo,
112 1996; Singer, 1986; Casagrande & Dacey, 2007). The agent screening process
113 utilised in biological control programs could be significantly improved if the behavioural
114 and physiological mechanisms which mediate host selection and acceptance are
115 better understood (Marohasy, 1998). A range of sensory cues are utilized by insects
116 to locate their hosts, including: visual (Fischer et al., 2004), olfactory (Visser, 1986)
117 and tactile/contact cues (Müller & Hilker, 2001). The efficiency and reliability of each
118 cue can be variable, but Heisswolf et al. (2007) proposed that olfactory cues are more
119 reliable than both visual and many contact cues for chrysomelid beetles (Bernays &
120 Chapman, 1994; Horiuchi et al., 2003).

121 Conspecific pheromones, or other aggregation cues, may also be involved in
122 host selection by certain chrysomelid beetles (Wood, 1982). Conspecific cues are not
123 usually explicitly considered in host specificity testing procedures, but in some
124 circumstances they could lead to false positive or false negative results. For example,
125 cues from one sex may influence host choice or feeding behaviour of the other sex,

126 leading to different results when both sexes are present or if one sex is tested alone.
127 A better understanding of the role of conspecific cues in host plant selection and
128 feeding behaviour could therefore help improve the reliability of host specificity testing
129 procedures.

130 In this study, data from sequential no-choice tests and paired choice tests were
131 combined with data from olfactory discrimination experiments in order to predict the
132 risk of non-target impacts to *K. africana* if *P. coerulea* were released as a biological
133 control agent in areas where both species are present.

134

135 **2. METHODS AND MATERIALS**

136

137 2.1 Insect and Plant Cultures

138 Potted *S. campanulata* and *K. africana* plants were obtained from local nurseries and
139 housed in a greenhouse before being taken into quarantine for experiments.
140 *Paradibolia coerulea* individuals were collected from 10 natural populations in March
141 2014 in Ghana, West Africa. The 10 wild populations were combined into a single
142 culture population after importation into quarantine in South Africa. The *P. coerulea*
143 culture was maintained under a 16:8 hour light/dark regime and temperatures were
144 maintained at 25 ± 2 °C (S.E.). The culture was provided with potted *S. campa nulata*
145 when required. Larvae were transferred to 2 L tubs layered with potting soil, and
146 provided with fresh plant material daily. The soil was kept moist at all times to provide
147 a suitable pupation substrate.

148

149

150 2.2 Sequential No-Choice Tests

151 Recently-eclosed naïve *P. coerulea* adults (eclosed within 24 hours of the
152 experiment and not exposed to any food plant) were inoculated onto potted *S.*
153 *campanulata* (test plant A) or *K. africana* (test plant B) plants, in an ABA and BAB
154 sequence (Fig. 1). The plants were approximately 1m in height and were housed
155 individually in standard 1.2m x 0.6m x 0.6m insect cages. Each cage was inoculated
156 with a pair of adult beetles (1 female, 1 male). [Beetles are readily sexed as female *P.*](#)
157 [coerulea possess a red, vertical stripe on the dorsal surface of the abdomen, which is](#)
158 [absent in male beetles.](#) After 14 days, the plants species in each cage was alternated
159 and the original *P. coerulea* pair was offered the alternate plant in the test sequence.
160 After an additional 14 days, the beetle pair were offered again the test plant species
161 provided during the first 14 days. A new, undamaged plant was used each time the
162 plant was changed. Controls employed an AAA test sequence. The number of feeding
163 scars, eggs, larvae, larval tunnels and leaves damaged on the plants were recorded
164 for each of the plants in the sequence. The experiment was replicated five times.

165

166 2.3 Paired Choice Tests

167 Two recently-eclosed naïve *P. coerulea* adults (1 male, 1 female) were placed in
168 insect cages (1.2m x 0.6m x 0.6m) with similar sized potted *S. campanulata* and *K.*
169 *africana* plants. After 28 days the number of feeding scars, eggs, larvae, larval tunnels
170 and leaves damaged were recorded. If the adults died during the 28 days then that
171 replicate was discarded. The experiment was replicated six times.

172

173

174

175 2.4 Host Plant Olfactory Cues

176 The response of *P. coerulea* to olfactory stimuli from the two test plant species was
177 determined using a stem-arena design ([Müller Müller & Hilker, 2000](#)). [Stems](#)
178 [\(approximately 60mm in length x 20mm in diameter\) were erected by enclosing](#)
179 [detached plant material within a piece of filter-paper that was shaped into a cylinder,](#)
180 [to ensure that *P. coerulea* had no contact with, or visual of, plant material \(e.g.,](#)
181 [Heisswolf et al., 2007\).](#) A petri dish [\(90mm x 12mm\)](#) was half-filled with soil ~~so that~~
182 ~~leaves wrapped in filter paper and stems were~~ ~~could be~~ pressed into the soil, allowing
183 them to stand upright. The petri dish was placed in a 2 L tub, and covered with a fine
184 mesh cloth to prevent any beetles from escaping the arena ([see Heisswolf et al., 2007](#)
185 [for a graphical depiction of the stem-arena design](#)). A single leaf [\(approximately 60mm](#)
186 [x 40mm\)](#) of each test plant was placed within ~~rolled filter paper~~ [a stem](#), and placed in
187 the stem-arena. Controls were erected by placing a toothpick inside ~~a the rolled filter~~
188 ~~paper~~ ~~stem~~. There were four stems used per arena, with two stems used per treatment,
189 and the position of each treatment was randomly allocated to account for spatial bias.
190 The treatments that were applied include: (i) *Spathodea campanulata*/Control, (ii)
191 *Spathodea campanulata*/*Kigelia africana* and (iii) *Kigelia africana*/Control

192 The experiment was performed by introducing a single, naïve *P. coerulea* adult into
193 the middle of the arena, allowing for a 5 minute acclimatisation period. Each beetle
194 was sexed before being placed in the arena. The beetle was then observed for 30
195 minutes, recording the amount of time spent on each stem. Each treatment was
196 replicated ≥ 9 times, using a beetle only once to control for the effects of experience
197 and learning. Only beetles that made a choice within the allotted time were included
198 in the analysis.

199

200 2.5 Conspecific Olfactory Cues

201 An aggregation test was employed to determine whether conspecific contact cues
202 were involved in beetle location behaviour (Tansey et al., 2005). A single adult *P.*
203 *coerulea*, between 12 and 15 days old, was sexed and then placed inside a glass vial
204 (60 emm x 1.5 mem) lined with filter paper. The beetle was left for 24 hours before
205 being removed from the vial to allow for adequate deposition of any potential contact
206 cues. Another *P. coerulea* adult was then introduced into the treated vial, which was
207 connected to a second glass vial lined with untreated filter paper, and the two were
208 sealed together with masking tape. The newly-introduced beetle was allowed to
209 acclimate inside the apparatus for 30 minutes. The position of the beetle in the vials
210 containing treated (“1”) or untreated (“0”) filter paper was scored as a binary response
211 at 15 minute intervals over a two-hour observation period, whereby each two-hour
212 observational period constituted a single replicate.

213 Thirty replicates were performed for each treatment, which included: (1) female-
214 male (female response to male contact cues), (2) male-female, (3) female-female and
215 (4) male-male combinations.

216

217 2.6 Statistical Analyses

218 All data were subjected to normality testing by visually observing box plots and
219 Shapiro-Wilks tests, and homogeneity of variances were analysed by performing
220 Bartlett’s Test. Data that failed to meet these assumptions were subjected to
221 appropriate transformations, or analysed using non-parametric statistical methods.

222 Herbivore performance between test-plant species for paired-choice experiments
223 was analysed by paired t-tests (Horton, 1995).

224 Sequential no-choice experiments were analysed by two-way analysis of variance
225 (ANOVA) with regards to plant-sequence effect on the number of feeding scars
226 observed. Paired t-tests were performed on contrasting treatments to determine
227 whether prior experience on a test-plant influenced host-use. Reproductive output of
228 *P. coerulea* was not subjected to statistical analyses as only *P. coerulea* pairs on *S.*
229 *campanulata* produced larvae.

230 The preference of *P. coerulea* to stems ([i.e., filter paper cylinders](#) containing leaf
231 material) of either test-plant or controls were analysed by binomial tests, which were
232 further partitioned into male and female responses, to determine whether gender
233 influenced olfactory host-discrimination. The latency period ([i.e.,](#) time taken before a
234 stem was chosen), (Heisswolf et al., 2007), was analysed by one-way ANOVA, and
235 again was partitioned into gender-wise categories.

236 The responses obtained during the conspecific cues experiment were recorded as
237 the number of times out of the possible eight sampling events that the beetles were
238 observed in the vial containing the treated filter paper (denoted N_T). Kruskal-Wallis
239 one-way ANOVA was used to determine any treatment effects. Post-hoc analysis was
240 performed by implementing a multiple comparisons test after Kruskal-Wallis to
241 elucidate which gender-combination treatments yielded significant responses. One-
242 sample t-tests were performed to demonstrate whether the N_T responses differed from
243 the hypothesized mean of four responses for each filter paper treatment (Tansey et
244 al., 2005).

245
246
247
248

249 3. RESULTS

250

251 3.1 Sequential no-choice tests

252 *Paradibolia coerulea* demonstrated a clear preference for *S. campanulata* over *K.*
253 *africana* in sequential no-choice experiments (Fig. 2). The test plant species offered
254 to the beetle significantly influenced the number of feeding scars measured ($F_{1,20} =$
255 348.56 , $P = <0.0001$) (Fig. 2a), while a significant interaction between the sequence
256 of plants offered and the test species was also observed ($F_{1,20} = 9.01$, $P = 0.0085$).

257 *P. coerulea* Beetle larvae were only observed on *S. campanulata*, with the majority
258 of larvae found on the control plant sequence AAA (Fig. 2b). Larvae were not found
259 for BAB sequence *S. campanulata* plants, however larvae were produced on the
260 second presentation of *S. campanulata* during ABA sequence replicates, indicating a
261 host-plant and temporal effect on the beetle's reproductive output (Fig. 2b).

262 3.2 Paired-choice tests

263

264 *Paradibolia coerulea* adults demonstrated a clear preference for *S. campanulata*
265 during choice-test trials when offered both plant species together (Table 1). The
266 number of feeding scars measured was approximately four times greater on *S.*
267 *campanulata* than on *K. africana* ($t_{(5)} = 4.48$, $P = 0.0065$) while the number of *K.*
268 *africana* leaves damaged by adult feeding was less than half than the number of leaves
269 damaged on *S. campanulata* plants ($t_{(5)} = 4.73$, $P = 0.0072$). The number of eggs laid
270 on *S. campanulata* was six times more than for *K. africana* ($t_{(5)} = 6.74$, $P = 0.0011$),
271 while the number of larvae observed on *K. africana* was approximately 20 times fewer
272 than larvae on *S. campanulata* ($t_{(5)} = 5.71$, $P = 0.0023$). The number of feeding tunnels
273 was significantly greater on *S. campanulata* than *K. africana* ($t_{(5)} = 6.94$, $P = 0.0001$),

274 with 28.17 ± 4.36 (mean \pm S.E.) feeding tunnels on *S. campanulata* and only $2.00 \pm$
275 1.12 tunnels on *K. africana*. Only two larvae were observed on *K. africana* and neither
276 survived through to the end of the first instar (100 ± 0 % mortality), while larval mortality
277 on *S. campanulata* was significantly lower at 15 ± 6 % ($t_{(5)} = 17.12$, $P = 0.0034$).

278

279 3.3 Host Plant Olfactory Cues

280

281 The beetles demonstrated a clear preference for arena-stems containing *S.*
282 *campanulata* leaf material rather than empty controls (Fig. 3A, $n = 10$, $P = 0.0107$).
283 There was an appreciable attraction to *K. africana* leaf material rather than to the
284 controls although this result was not significant (Fig. 3B, $n = 13$, $P = 0.1334$).

285 *Paradibolia coerulea* demonstrated a preference for *S. campanulata* leaf material
286 over the *K. africana* leaves (Fig. 3C, $n = 29$, $P = 0.0121$). Female beetles demonstrated
287 a clear preference for *S. campanulata* (Fig. 3D, $n = 20$, $P = 0.0059$), however there
288 was no apparent olfactory discrimination between *S. campanulata* or *K. africana* for
289 male beetles (Fig. 3E, $n = 9$, $P = 0.5000$), indicating a gender-based response to
290 olfactory cues from the host plants.

291 The latency period did not differ for *S. campanulata* or *K. africana* ($F_{1,28} = 0.39$, $P =$
292 0.5390). However, latency was significantly greater for male than for female *P.*
293 *coerulea* ($F_{1,28} = 5.7320$, $P = 0.0239$).

294

295 3.4 Conspecific Cues

296 There was a significant difference in the number of times *P. coerulea* was observed
297 in the vial containing treated filter paper with respect to the gender combination
298 treatment ($H_{3,120} = 25.18$, $P = <0.0001$). Significant deviation from the expected $N_T =$

299 4, was observed for the female response to male and male response to other male
300 treatments (Table 2). Females were appreciably attracted to male cues ($P = <0.0001$),
301 whereas male-treated filter paper deterred other male beetles ($P = 0.0107$). There was
302 no apparent response to female cues by either other females ($P = 0.4203$), or by males
303 ($P = 0.1694$).

304

305 **4. DISCUSSION**

306

307 Adult *P. coerulea* will feed on the leaves of both *S. campanulata* and *K. africana*,
308 but larval development is not possible on *K. africana*. Acceptability for oviposition was
309 12 times greater on *S. campanulata* than for *K. africana* in paired choice tests, and in
310 sequential no-choice tests larvae were only ever found on *S. campanulata*. Adult
311 feeding on *S. campanulata* was approximately four times greater than on *K. africana*,
312 indicating a clear preference for the target plant. Non-target feeding on *K. africana* was
313 recorded under both choice and no-choice conditions, suggesting that a spill-over
314 effect might occur in the field if populations of *P. coerulea* built-up on *S. campanulata*
315 growing in close proximity to *K. africana*, although the relative performance was so
316 poor on *K. africana* that significant levels of attack are unlikely (see Paynter et al.,
317 2015).

318 *Paradibolia coerulea* utilises olfactory cues to help select its typical host plant. This
319 has also been shown to be the case for many other Chrysomelidae (McIndoo, 1926;
320 Metcalf & Metcalf, 1992; Muller & Hiker, 2000; Hori, Ohuchi & Matasuda, 2006;
321 Fernandez & Hilker, 2007). There was a preference for cues emitted by *S.*
322 *campanulata* over both control and *K. africana* containing-stems, but this was limited
323 to female individuals. Male beetles demonstrated no olfactory discrimination between
324 host plants and had a significantly greater latency period before selecting either host.

325 During choice experiments there were several instances where eggs or larvae were
326 evident on *K. africana*, yet in sequential no-choice assays there were no eggs on *K.*
327 *africana* plants, which could indicate that oviposition-stimulating volatiles from *S.*
328 *campanulata* accumulated in the paired-choice cages and this resulted in some
329 oviposition on the atypical host. Our findings indicate that if female *P. coerulea* are
330 unable to locate *S. campanulata*, there is little risk of oviposition on *K. africana*.

331 Another factor that may contribute to host plant selection by insects is that of
332 conspecific herbivore chemical emissions (Tansey et al., 2005; Fernandez & Hilker,
333 2007). There is substantial evidence among chrysomelid beetles for roles in attracting
334 conspecifics by both female-emitted sex pheromones (Cuthbert & Reid, 1964; Zhang
335 & McEvoy, 1994) and male-emitted aggregation pheromones (Peng, Bartelt & Weiss,
336 1999; Dickens et al., 2002). Our results demonstrate a strong attraction to male-
337 emitted cues by conspecific female *P. coerulea* individuals, while males were deterred
338 by the presence of other male cues. Whether the individuals used in the study had
339 mated was unknown, and it is possible that the response to conspecifics could change
340 after mating. Tansey et al. (2005) obtained a similar result with female *Aphthona*
341 *nigriscutis* Faudras (Chrysomelidae) orientating to male-emitted cues. Wood (1982)
342 proposed that this male-attract-female synergism for *Dendroctonus frontalis*
343 Zimmermann (Coleoptera: Chrysomelidae) allowed for host plant selection by the
344 male, which would then attract female conspecifics. Our results indicate that male *P.*
345 *coerulea* are unable to discriminate between the two plants but that the females are
346 attracted to the preferred host, *S. campanulata*. Females are required to locate their
347 host as they select plants for oviposition and the larvae can survive on *S. campanulata*,
348 but not *K. africana*. Both sexes can feed on either species as adults so there may be
349 no selection pressure for males to be attracted to *S. campanulata* specific volatiles.

350 Females are also apparently required to seek out males for mating as females showed
351 an attraction to male beetles but no attraction of males to females was recorded. This
352 suggests that another cue, such as a visual or tactile cue, attracts males to *S.*
353 *campanulata* plants under natural conditions. Alternatively, males may only be
354 attracted to virgin females and most of the 12-15 day old beetles used in the trials may
355 have been mated.

356 The different response of the sexes to conspecifics, as well as to the host plant,
357 should be considered when further host specificity testing is conducted. If choice tests
358 are required, it is important that the sex of the beetles that are used in the tests are
359 known. If male beetles are used, then they are likely to feed indiscriminately on all
360 close relatives of *S. campanulata* because they are not attracted by the volatiles of the
361 primary host. If females are used in the same tests, a much higher proportion of
362 feeding on *S. campanulata* would be expected. The presence of males may also
363 influence which plants female beetles feed on, because females are attracted to
364 males. A higher proportion of males in a choice experiment could therefore result in
365 greater levels of feeding on non-target plants.

366 The results of the sequential no-choice tests suggest that female adult feeding on
367 *S. campanulata* is required for the production of eggs. *P. coerulea* was capable of egg
368 production if newly emerged beetles had fed on *S. campanulata* (in the ABA treatment)
369 but not when the newly emerged beetles had fed on *K. africana* (in the BAB treatment).
370 Egg production was also greatest in the AAA treatment where the beetles had the
371 longest period of exposure to *S. campanulata*. A similar effect was observed in host
372 specificity testing of the leaf-mining fly, *Phytomyza vitalbae* Kaltenbach
373 (Agromyzidae), a biological control agent for *Clematis vitalba* L. (Ranunculaceae),
374 which produced fertile eggs on closely related non-target species, but only when adults

375 had previously fed on the target plant (Schwarzländer, Hinz & Wittenberg, 1996). Adult
376 *P. vitalbae* that had not fed on *C. vitalba* were infertile and spill-over onto non-target
377 species in the field was therefore limited (Paynter et al., 2008). Non-target damage in
378 the field was usually restricted to an area within 4km of populations of the target plant
379 and was never found further than 30km from a *C. vitalba* population (Paynter et al.,
380 2008). A similar spill-over effect on *K. africana* would be expected if *P. coerulea* were
381 released in a region where both *S. campanulata* and *K. africana* occur.

382 This study demonstrates how investigations into the effect of olfactory cues can
383 be useful in elucidating the host range and behaviours of potential biological control
384 agents. In this case, *P. coerulea* prefers *S. campanulata* and cannot complete its
385 development on *K. africana*, but some spill-over onto *K. africana* may occur if the
386 insect were released in areas where the two plants grow together. Any damage to *K.*
387 *africana* would be a temporary spill-over at times when abundance of *P. coerulea* on
388 *S. campanulata* was high. Further host specificity testing, using other test plants in the
389 family Bignoniaceae and from closely related families, is required before a full
390 assessment of the host range of *P. coerulea* or decisions about the safety of the beetle
391 as a biological control agent can be made, but the design and interpretation of these
392 host specificity tests should take the olfactory responses of *P. coerulea* to both the
393 host plant and conspecifics into consideration.

394

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407 **REFERENCES**

408

409 Bernardo, J. (1996). Maternal effects in animal ecology. *American Zoology*, 36, 83-
410 105.

411

412 Bernays, E.A., & Chapman, R.F. (1994). *Host-Plant Selection by Phytophagous*
413 *Insects*. Chapman and Hall, New York.

414

415 Bidgood, S. (1994). Intraspecific variation in *Spathodea campanulata*
416 (Bignoniaceae). In: Seyani, J.H., Chikuni, A.C. (Eds). *Proceedings of the XIIIth*
417 *Plenary Meeting AETFAT (Association for the Taxonomic study of the flora of*
418 *Tropical Africa)*, Malawi. (pp. 327-331).

419

420 Briese, D.T. (2005). Translating host-specificity test results into the real world: The
421 need to harmonize the yin and yang of current testing procedures. *Biological Control*.
422 35, 208–214.

423

424 Buckingham, G.R., Okrah, E.A., & Christian-Meier, M. (1991). Laboratory biology and
425 host range of *Hydrellia balciunasi* (Diptera: Ephydridae). *Entomophaga*. 36, 575-586.

426 Casagrande, R.A., & J.E. Dacey. (2007). Monarch Oviposition on Swallow-
427 worts (*Vincetoxicum* spp.). *Environmental Entomology*. 36, 631-636.

428

429 Coates-Palgrave, M. (2002) *Keith Coates-Palgrave Trees of Southern Africa*. 3rd
430 Edition. Struik Nature, Century City. Pg 1013.

431 Cuthbert, F.P., & Reid, J.R. (1964). Studies of a sex attractant of the banded cucumber
432 beetle. *Journal of Economic Entomology*. 57, 247-250.

433

434 Department of Environmental Affairs. (2014). Notice No. R. 598. National
435 Environmental Management: Biodiversity Act (10/2004): Alien and Invasive Species
436 Regulations, 2014, Government Gazette 37885, Pretoria, 1 August 2014.

437 (Regulation Gazette No. 10244).

438

439 Dickens, J.C., Oliver, J.E., Hollister, B., Davis, J.C., & Klun, J.A. (2002). Breaking a
440 paradigm: Male-produced aggregation pheromone of the Colorado potato beetle.
441 *Journal of Experimental Biology*. 205, 1925-1933.

442

443 Edwards, P.B. (1999). The use of choice tests in host-specificity testing of herbivorous
444 insects (pp. 35-43). In: Withers, T.M., Barton-Browne, L. & J. Stanley (Eds.). *Host*
445 *Specificity Testing in Australasia: Towards Improved Assays for Biological Control*.
446 Cooperative Research Centre for Tropical Pest Management, Brisbane, Australia.

447

448 Fernandez, P., & Hilker, M. (2007). Host plant location by Chrysomelidae. *Basic and*
449 *Applied Ecology*. 8, 97-116.

450

451 Fischer, S., Samietz, J., Wäckers, F. L., & Dorn, S. (2004). Perception of chromatic
452 cues during host location by the pupal-parasitoid *Pimpla turionellae* (L.)
453 (Hymenoptera: Ichneumonidae). *Environmental Entomology*. 33, 81–87.

454

455 Francis, J.K. (1990). *Spathodea campanulata* Beauv. African Tulip Tree.
456 Bignoniaceae. Bignonia family. USDA Forest Service, Southern forest Experiment
457 Station, Institute of Tropical Forestry. 5 pg. (SO-ITF-SM-32).

458

459 Hedberg, I., Kelbessa, E., Edwards, S., Demissew, S., & Persson, E. (2006). Flora of
460 Ethiopia and Eritrea Volume 5. (pp. 327-328). Uppsala University, Sweden.

461

462 Heisswolf, A., Gabler, D., Obermaier, E., & Müller, C. (2007). Olfactory versus contact
463 cues in host plant recognition of a monophagous chrysomelid beetle. *Journal of Insect*
464 *Behaviour*. 20, 247–266.

465

466 Horiuchi, J., Arimura, G., Ozawa, R., Shimoda, T., Takabayashi, J., & Nishioka, T.
467 (2003). A comparison of the responses of *Tetranychus urticae* (Acari: Tetranychidae)
468 and *Phytoseiulus persimilis* (Acari: Phytoseiidae) to volatiles emitted from Lima Bean
469 leaves with different levels of damage made by *T. Urticae* or *Spodoptera exigua*
470 (Lepidoptera: Noctuidae). *Applied Entomology and Zoology*. 38, 109-116.

471

472 Hori, M., Ohuchi, K., & Matasuda, K. (2006). Role of host plant volatile in the host-
473 finding behavior of the strawberry leaf beetle, *Galerucella vittaticollis* Baly (Coleoptera:
474 Chrysomelidae). *Applied Entomology and Zoology*. 41, 357-363.

475

476 Horton, D.R. (1995). Statistical considerations in the design and analysis of paired-
477 choice assays. *Environmental Entomology*. 24, 179-192.

478

479 Labrada, R., & Diaz Medina, A. (2009). The invasiveness of the African Tulip Tree,
480 *Spathodea campanulata* Beauv. *Biodiversity*. 10, 79-82.
481

482 Larrue, S., Daehler, C., Vautier, F., & Bufford, J.L. (2014). Forest invasion by the
483 African Tulip Tree (*Spathodea campanulata*) in the Hawaiian Islands: Are seedlings
484 shade tolerant? *Pacific Science*. 68, 1-27.
485

486 Lowe, S., Browne, M., Boudjelas, S., & De Poorter, M. (2000). 100 of the World's
487 Worst Invasive Alien Species: a selection from the Global Species Database. Invasive
488 Species Specialist Group (ISSG) a specialist group of the Species Survival
489 Commission (SSC) of the World Conservation Union (IUCN). *Aliens*. 12, 1-12.
490

491 Marohasy, J. (1998). The design and interpretation of host-specificity tests for weed
492 biological control with particular reference to insect behaviour. *Biocontrol News and*
493 *Information* 19, 13-20.
494

495 McIndoo, N.E. (1926). An insect olfactometer. *Journal of Economic Entomology* 19,
496 545-571.
497

498 Metcalf, R.L., & Metcalf, E.R. (1992). *Plant kairomones in insect ecology and control*.
499 Chapman & Hall, New York.
500

501 Moffat, C., Lalonde, R.G., Ensing, D., De-Clerke Floate, D., Grosskopf-Achat, G., &
502 Pither, J. (2013). Frequency-dependent host species use by a candidate biological
503 control insect within its native European range. *Biological Control*, 67, 498-508.

504

505 Müller, C., & Hilker, M. (2000). The effect of a green leaf volatile on host plant finding
506 by larvae of a herbivorous insect. *Naturwissenschaften*. 87, 216–219.

507

508 Müller, C., & Hilker, M. (2001). Host finding and oviposition behaviour in a chrysomelid
509 specialist – the importance of host plant surface waxes. *Journal of Chemical Ecology*.
510 27, 985–994.

511

512 Olmstead, R. G., Zjhra, M. L., Lohmann, L. G., Grose, S. O., & Eckert, A. J. (2009). A
513 Molecular Phylogeny and Classification of Bignoniaceae. *American Journal of Botany*.
514 96, 1731–1743.

515

516 Paterson, I.D., Paynter, Q., Nesar, S., Akpabey, F.J., Orapa, W., Compton, S. West
517 African arthropods hold promise as biological control agents for a Pacific Island
518 invader. *African Entomology*. IN PRESS

519

520 Paynter, Q. 2010. Prioritisation of targets for biological control of weeds in the Pacific
521 region. LandCare Research Contract Report LC0910/190.

522

523 Paynter, Q., Fowler, S.V., Gourlay, A.H., Peterson, P.G., Smith, L.A., & Winks, C.J.
524 (2015). Relative performance on test and target plants in laboratory tests predicts the
525 risk of non-target attack in the field for arthropod weed biocontrol agents. *Biological*
526 *Control*. 80, 133-142.

527

528 Paynter, Q., Martin, N., Berry, J., Hona, S., Peterson, P., Gourlay, A.H., Wilson-Davey,
529 J., Smith, L., Winks, C., Fowler, S.V., 2008. Non-target impacts of *Phytomyza vitalbae*
530 a biological control agent of the European weed *Clematis vitalba* in New Zealand.
531 *Biological Control*. 44, 248-258.

532

533 Peng, C., Bartelt, R.J., & Weiss, M.J. (1999). Male crucifer flea beetles produce an
534 aggregation pheromone. *Physiological Entomology*. 24, 98–99.

535

536 Schaffner, U. (2001). Host Range Testing of Insects for Biological Weed Control: How
537 Can It Be Better Interpreted? *BioScience*. 51, 951-959.

538

539 Schwarzländer, M., Hinz, H.L., & Wittenberg, R., (1996). Oogenesis requirements and
540 weed biocontrol: an essential part in host-range evaluation of insect agents or just
541 wasted time? (pp. 79.85). In: Moran, V.C., Hoffmann, J.H., (Eds.), *Proceedings of the*
542 *9th International Symposium on Biological Control of Weeds*. University of Cape Town,
543 South Africa.

544

545 Singer, M.C. (1986). The definition and measurement of oviposition preference in
546 plant-feeding insects. (pp. 74-86). In: Miller, J.R. & Miller, T.A. (Eds.) *Insect-Plant*
547 *Interactions*. New York: Springer Verlag.

548

549 Tansey, J.A., McClay, A.S., Cole, D.E., & Keddie, B.A. (2005). Evidence for the
550 influence of conspecific chemical cues on *Aphthona nigriscutis* (Coleoptera:
551 Chrysomelidae) behaviour and distribution. *BioControl*. 50, 343-358.

552

553 Withers, T.M., Barton-Browne, L., & Stanley, J. (1999). Host Specificity Testing in
554 Australasia: Towards Improved Assays for Biological Control. Queensland
555 Department of Natural Resources, Brisbane.

556

557 Withers, T. M., Barton-Browne L., & Stanley J.N. (2000). How time-dependent
558 processes can affect the outcomes of assays used in host specificity testing. (pp. 27-
559 41). In: Van Driesche, R.G., T. A. Heard, A. S. McClay, & R. Reardon (Eds.)
560 Proceedings of the 10th International Symposium on Biological Control of Weeds.
561 Bozeman, Montana, USA. July 4-14, 1999. Forest Service Bulletin, FHTET-99-1,
562 Morgantown, West Virginia, USA.

563

564 Withers, T.M. & Mansfield, S. (2005). Choice or no-choice tests? Effects of
565 experimental design on the expression of host range. (pp. 620-633). In: M. Hoddle
566 (Ed.) Proceedings of the 2nd International Symposium of Biological Control of
567 Arthropods. USDA Forest Service, Morgantown, West Virginia, U.S.A.

568

569 Wood, D.L. (1982). The role of pheromones, kairomones, and allomones in the host
570 selection and colonization behaviour of bark beetles. Annual Review of Entomology.
571 27, 411–446.

572

573 Visser, J.H. (1986). Host odour perception in phytophagous insects. Annual Review
574 of Entomology. 31, 121–144.

575 Zhang, Z. & McEvoy, P.B. (1994). Attraction of *Longitarsus jacobaeae* males to cues
576 associated with conspecific females (Coleoptera: Chrysomelidae). Physiological and
577 Chemical Ecology. 23, 732–737.

578 **Legend to figures**

579 **Fig. 1.** Schematic overview of sequential no-choice test design for host-range
580 determination of *Paradibolia coerulea*.

581 **Fig. 2.** Feeding (a) and reproductive (b) responses of *P. coerulea* pairs to sequential
582 presentation of *Spathodea campanulata* (A) and *Kigelia africana* (B) in an ABA and
583 BAB sequence, with a control employed in an AAA sequence. Bars indicate a mean
584 (\pm SD), with grey bars for *S. campanulata* and white bars for *K. africana*. An asterisk
585 indicates a value of zero. Five replicates were conducted per plant sequence.

586 **Fig. 3.** Preference responses (%) of *Paradibolia coerulea* to olfactory cues emitted by
587 *Spathodea campanulata* (SP) and *Kigelia africana* (KI) leaves, in a stem-arena.
588 Controls (CT) had no plant material placed inside a stem. Treatments were: (A) SP/CT,
589 (B) KI/CT, (C) SP/KI, (D) SP/KI (female beetles) and (E) SP/KI (male beetles). Sample
590 sizes (n) and P-values (P) are provided for each treatment, with significant preferences
591 following binomial tests indicated by an asterisk.

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