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1 **Opinion for Trends in Ecology and Evolution**

2 **Title:** Re-introducing environmental change drivers in biodiversity-ecosystem functioning
3 research

4 **Authors:** Frederik De Laender^a, Jason R. Rohr^b, Roman Ashauer^c, Donald J. Baird^d, Uta
5 Berger^e, Nico Eisenhauer^{f,g}, Volker Grimm^{g,h}, Udo Hommenⁱ, Lorraine Maltby^j, Carlos J.
6 Melià^k, Francesco Pomati^l, Ivo Roessink^m, Viktoriia Radchuk^{g,n}, Paul J. Van den Brink^{m,o}

7 ^aResearch Unit in Environmental and Evolutionary Biology, University of Namur, Belgium.

8 Frederik.delaender@unamur.be

9 ^bDepartment of Integrative Biology, University of South Florida, USA. jasonrohr@gmail.com

10 ^cEnvironment Department, University of York, UK. roman.ashauer@york.ac.uk

11 ^dEnvironment Canada, Canadian Rivers Institute, University of New Brunswick, Canada.

12 [djbaire@unb.ca](mailto:djbaird@unb.ca)

13 ^eInstitute of Forest Growth and Computer Sciences, Technische Universitaet Dresden (TU
14 Dresden), Germany. uta.berger@tu-dresden.de

15 ^fInstitute of Biology, Leipzig University, Germany.

16 ^gGerman Centre for Integrative Biodiversity Research (iDiv) Halle-Jena-Leipzig, Germany.

17 nico.eisenhauer@idiv.de

18 ^hDepartment of Ecological Modelling, Helmholtz Centre for Environmental Research – UFZ,
19 Germany. volker.grimm@ufz.de

20 ⁱFraunhofer Institute for Molecular Biology and Applied Ecology (Fraunhofer IME), Germany.

21 udo.hommen@ime.fraunhofer.de

22 ^jDepartment of Animal and Plant Sciences, The University of Sheffield, UK.

23 l.maltby@sheffield.ac.uk

24 ^kCenter for Ecology, Evolution and Biogeochemistry, Swiss Federal Institute of Aquatic
25 Science and Technology, Switzerland. Carlos.Melian@eawag.ch

26 ^lEawag, Swiss Federal Institute of Aquatic Science and Technology, Department of Aquatic
27 Ecology, 8600 Dübendorf, Switzerland. Francesco.Pomati@eawag.ch

28 ^mAlterra, Wageningen University and Research centre, the Netherlands. ivo.roessink@wur.nl

29 ⁿDepartment of Evolutionary Ecology, Leibniz Institute for Zoo and Wildlife Research (IZW),
30 Germany. radchuk.victoria@gmail.com

31 ^oDepartment of Aquatic Ecology and Water Quality Management, Wageningen University, the
32 Netherlands. paul.vandenbrink@wur.nl

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34 **Corresponding author:** Frederik De Laender, Rue de Bruxelles 61 - 5000 Namur, Belgium;
35 Frederik.delaender@unamur.be, T: +32 478 42 97 61; F: +32 81 724 362

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37

38 **Abstract**

39 For the past 20 years, research on biodiversity and ecosystem functioning (B-EF) has only
40 implicitly considered the underlying role of environmental change. We illustrate that explicitly
41 re-introducing environmental change drivers in B-EF research is needed to predict the
42 functioning of ecosystems facing changes in biodiversity. Next, we show how this re-
43 introduction also allows for better control over community composition in B-EF experiments
44 and thus helps to obtain mechanistic insights about how multiple aspects of biodiversity relate
45 to function, and how biodiversity and function relate in food-webs. We also highlight
46 challenges for the proposed re-introduction, and propose analyses and experiments to better
47 understand how random biodiversity changes, as studied by classic approaches in B-EF
48 research, contribute to the shifts in function that follow environmental change.

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68 **1. Predicting effects on ecosystem functions from changes in biodiversity: a brief history**

69 Various types of environmental change, such as climate change and the deterioration of
70 environmental quality, can profoundly alter multiple facets of biodiversity [1-3]. The past 25
71 years have seen a rise in different empirical approaches to examine how such changes affect
72 ecosystem functions and services [4, 5]. Many focus on altering biodiversity while observing
73 any corresponding changes in function [6]. These approaches can be first classified based on
74 the nature of the manipulation (Table 1 and ‘Glossary’), whether species are added or removed
75 randomly or non-randomly. Random manipulations assume a random extinction or
76 colonization order, while non-random species additions or removals are done based on the
77 (presumed) response of species to environmental change [7], or based on the effects of species
78 on function (e.g. species that function best are removed first) [8]. A second distinction can be
79 based on whether manipulations of biodiversity are direct or indirect. Direct biodiversity
80 manipulations are performed by manually removing or adding species [9], whereas with
81 indirect manipulations, a relevant environmental change is introduced to alter biodiversity [10,
82 11].

83

84 Indirect and non-random manipulations of biodiversity make intuitive sense because they are
85 rooted in a recognition that environmental change drivers are often the cause of biodiversity
86 alterations [3] and that these alterations are non-random [8, 12]. As a consequence, early
87 research on biodiversity and ecosystem functioning (‘B-EF research’ [6]) often adopted
88 indirect and non-random biodiversity manipulations [10, 11, 13]. However, such approaches
89 were increasingly subject to controversy and disagreement. In his seminal paper, Huston [14]
90 criticized indirect and non-random biodiversity manipulations for difficulties in separating
91 ‘true’ biodiversity effects from the effects of ‘hidden treatments’. Huston argued that by
92 indirectly altering biodiversity using an environmental variable, researchers precluded
93 partitioning the effects of biodiversity on ecosystem functioning from the many other effects
94 environmental change can have on function (‘Glossary’). Non-random manipulations were also
95 shown to suffer from inherent bias, because results were highly dependent on the chosen order
96 of species removal or addition. Collectively, the critiques by Huston and others [14-16] pushed
97 the field towards direct and randomized biodiversity manipulations (Table 1, [6, 9]). The
98 advantage of this methodological shift was that the causal relationship between biodiversity
99 and ecosystem functioning could be more rigorously tested, and the different effects that
100 biodiversity alterations could have on ecosystem functioning were more readily quantified
101 [17]. However, despite this improvement in methodology, it still remains unclear how either

102 random or direct manipulations of biodiversity can be used to meet two of ecology's main
103 challenges: (1) to support quantitative prediction of the ecological effects of anthropogenic
104 activities [6]; and (2) to unravel the mechanisms linking community structure and composition
105 to ecosystem function [18, 19]. An emerging trend in contemporary B-EF research is to re-
106 introduce non-random and indirect manipulations of biodiversity using environmental change
107 drivers, such as various climate variables, chemical stressors, and nutrient enrichment [20-24].
108 In the present contribution we submit that this approach (1) is a prerequisite to predicting the
109 functioning of ecosystems facing changes in biodiversity that are caused by environmental
110 change (section 2); and (2) facilitates unravelling mechanistic insight into the connections
111 between community structure and composition to ecosystem function (section 3).

112

113 **2. The re-introduction of environmental change drivers is needed to predict ecosystem** 114 **functioning following changes in biodiversity**

115 In many ecosystems, environmental change causes biodiversity declines or increases [25-28].
116 Experiments that directly and randomly manipulate biodiversity are unlikely to predict function
117 in these ecosystems (Fig. 1, shaded area). This is because biodiversity changes that are non-
118 random with respect to function ('Glossary') will affect ecosystem function more or less than
119 do random biodiversity changes [8, 29]. In addition, environmental change can also alter the
120 effect species have on ecosystem functions by altering (1) per-capita functioning [30, 31], and
121 (2) population density [32, 33]. Depending on the type of environmental change, these
122 alterations can be mostly positive (e.g. nutrient enrichment, [34]), mostly negative (e.g.
123 drought, [35], pollution, [36]), or negative for some species and positive for others (e.g.
124 warming, [37-39]).

125

126 Trait-based frameworks are available to predict how non-random effects of environmental
127 change on per-capita functioning, population densities, and biodiversity translate to changes in
128 ecosystem function [8, 29]. A simple extension of this framework with species interactions
129 (Box 1) and using richness as a biodiversity indicator illustrates two important points. First,
130 environmental change can cause a variety of B-EF relationships (Fig. 1). The shape of this
131 relationship critically depends on (1) whether the responses elicited by the environmental
132 change driver are positive or negative, and (2) the type of non-randomness exerted by the
133 environmental change driver [27, 40] (Box 1). Second, changes in function are expected before
134 any change in species richness is observed (Fig. 1A and D; levels 0-0.1), and – more generally
135 – the variability of ecosystem function within one level of species richness is substantial (Box

136 1, Box 3, ‘Outstanding questions’). The ensemble of B-EF relationships constructed through
137 direct and random biodiversity manipulation (Fig. 1, shaded area) does not capture the variation
138 in B-EF shapes arising from indirect and non-random biodiversity manipulation, and can both
139 over- (e.g. Fig. 1B) and underestimate variation of function within one biodiversity level (e.g.
140 Fig. 1C).

141

142 **3. The re-introduction of environmental change drivers can augment mechanistic insight**

143 Many descriptors of biodiversity (richness, community composition and structure), but also
144 total density (community size) and per-capita functioning, can affect ecosystem functions [32,
145 41, 42]. A main research theme in ecology is to understand their relative importance to function
146 [6, 43]. Using environmental change drivers to indirectly manipulate biodiversity, total density,
147 and per-capita functioning facilitates such studies. This is because different environmental
148 change levels trigger effects on different subsets of these variables (Fig. 1). For example, in
149 Fig. 1A, environmental change levels between 0.25 and 0.7 will all lead to the same species
150 richness, but alter total density and per-capita functioning. In Fig. 1B, effects on richness are
151 always more important than effects on total density or per-capita functioning. In Fig. 1A and
152 D, low levels of change only affect per-capita functioning and total density. The fact that
153 different levels of environmental change cause different effects offers greater control over the
154 different mechanisms underlying change of function than do direct manipulations of
155 biodiversity. Controlling per-capita functioning is by definition impossible through direct
156 manipulations of biodiversity because it is no descriptor of biodiversity, and always requires
157 some change of environmental conditions. Even community composition, structure, and
158 richness will often be uncontrollable through direct biodiversity manipulations. For example,
159 in the model presented in Fig. 1, persistence of species 0 or dominance by any other species
160 than species 9 is only possible in the continuous presence of an appropriate environmental
161 change driver, i.e. through indirect manipulations. Without this presence, community structure
162 will always converge to the one shown in Box 1 and richness will be 9, even when all 10 species
163 are added to the initial community. Many examples illustrate community compositions and
164 structures that only emerge in the presence of specific environmental change drivers and do
165 not occur in their absence (Table 1).

166 The relationship between biodiversity and functioning in multi-trophic communities (food-
167 webs) has been an important research theme in ecology since the 1990s [6, 44-46]. For
168 example, the biodiversity of one food-web compartment can drive functions performed by

169 other parts of the food-web [47], or both can be unrelated [48]. Using environmental change
170 drivers to indirectly and non-randomly manipulate biodiversity facilitates such studies. This is
171 because environmental change drivers often target specific food-web compartments so that it
172 becomes possible to experimentally alter biodiversity and related functions of specific food-
173 web compartments and measure corresponding changes in other compartments (Table 1). In
174 addition to the well-known cases of resource addition or manipulation of climate variables,
175 chemical stressors comprise an exceptionally useful group of experimental agents that can be
176 used for both non-random manipulations as well as for manipulations that are random with
177 respect to the effects species have on function. This is illustrated by the many studies that have
178 exposed relatively complex food-webs composed of field organisms (typically primary
179 producers and invertebrate grazers and predators) to concentration series of chemical stressors
180 during several weeks to months (Fig. 2). For example, many pyrethroid insecticides will target
181 arthropod consumers and predators [49, 50], while photosystem-inhibiting herbicides will
182 target specific algal taxa [51, 52]. In contrast, certain biocides such as triphenyltin [53] and
183 narcotic chemicals [54] are examples of chemical substances that exert effects that are random
184 with respect to function. Directly manipulating food-webs to persistently exclude certain
185 trophic levels or functional groups (e.g. small-bodied benthic grazers, specific bacterial
186 communities or algal taxa) will be nearly impossible. Indirect non-random manipulations might
187 therefore be the only solution.

188

189 **4. Back to the future: methods to connect indirect manipulations of biodiversity with** 190 **classic B-EF research**

191 Several methods are available that facilitate connecting indirect manipulations of biodiversity
192 to the classic approach of direct, random manipulations (Box 3, ‘Outstanding questions’) [22].
193 The literature is replete with studies exposing communities to environmental gradients
194 (bottom-right entry in Table 1). When a sufficient number of change levels has been tested
195 across a sufficiently broad gradient of change, the contributions of biodiversity-mediated
196 effects can be separated from the other effects of environmental change on ecosystem function
197 (‘Glossary’ contains examples of such effects) using available analytical techniques. One
198 possible way to do so is by applying multivariate statistical techniques, such as structural
199 equation modelling [58, 59] (Box 2). However, sophisticated structural equation models [20,
200 23] can also be used to partition the effects on function that are not mediated by biodiversity
201 into their constituents (‘Glossary’). In addition, methods based on versions of the Price
202 equation that do not require monoculture data but only need species contributions to function

203 before and after environmental change can be used to separate the effects of species loss and
204 gain that is random and non-random with respect to function from all other effects
205 environmental change can have on function [41].

206

207 Post-hoc analyses are a useful first step to quantify biodiversity-mediated effects on function.
208 However, we recommend combining direct and indirect biodiversity manipulations as separate
209 treatments in a single experiment, which is rarely done (Table 1). In a first design, we
210 recommend using a well-known environmental change driver to non-randomly manipulate a
211 community, while setting up another treatment where the same community is manipulated
212 directly, but non-randomly in a manner that aims to match the community resulting from the
213 application of the environmental change driver. For example, in Fig. 1B, applying a level of
214 change of 0.1 would constitute an indirect biodiversity manipulation that excludes species 1.
215 Higher levels would exclude species 2, 3, and so on. Thus, the direct biodiversity manipulation
216 treatments should represent the same gradient of community compositions, by consecutively
217 excluding species 1, 2, 3, and so on. Next, the B-EF relationship resulting from the indirect
218 manipulation (e.g. Fig. 1B, ‘resulting B-EF’ panel) could be compared to the one resulting
219 from the direct species removal. If both were not significantly different, this would suggest that
220 the chosen type of environmental change mainly acts upon ecosystem functioning through
221 effects on biodiversity. If B-EF relationships do differ, follow-up studies could examine in
222 more detail the potential mechanisms explaining this difference, for example by inspecting the
223 magnitude of effects on per-capita functioning [24]. However, we recognize that this design
224 can be challenging because, as mentioned in section 3, certain community compositions are
225 impossible to reconstruct without the use of environmental variables. This problem could be
226 addressed by statistically testing if per-capita functioning (functional contribution of a species,
227 e.g. its total biovolume divided by its population density) differs between the direct and indirect
228 biodiversity treatment. If the inferred values of per-capita functioning do not differ between
229 both biodiversity treatments, this suggests that the selected type of environmental change
230 impacts ecosystem functioning through other mechanisms than effects on per-capita
231 functioning.

232

233 A second design consists of a factorial experiment where the presence or absence of a direct
234 biodiversity manipulation that aims to match the community composition resulting from the
235 indirect biodiversity manipulation (as discussed in the first design) is crossed with the presence
236 and absence of an environmental change driver [60]. If all the effects of the driver on ecosystem

237 functioning are mediated by biodiversity changes, then the combination of direct biodiversity
238 manipulation and the environmental change treatment should display the same level of
239 ecosystem functioning as both the direct manipulation alone and the environmental change
240 treatment alone. If this were not the case, then it would suggest non-biodiversity-mediated
241 effects on ecosystem functioning.

242

243 **5. Challenges of re-introducing environmental change drivers in B-EF research**

244 Although we advocate re-introducing environmental change drivers to B-EF research, there are
245 are at least two challenges that need to be addressed for this to be successful. First, in the
246 approach we advocate, we implicitly assume that environmental change does not affect per-
247 capita species interactions (the α in Box 1). The effects of species interactions on a focal
248 species can only be altered through changes in the density of species with which it interacts.
249 This assumption has been shown to prevail in some systems [61], but not in others [62, 63].
250 Arguably the best-known example of environmental effects on per-capita interactions is the
251 ‘stress gradient hypothesis’, where there is a shift from competitive (i.e. negative) to facilitative
252 (i.e. positive) interactions as the level of stress increases [62, 63]. Such effects can lead to a
253 variety of effects of stress on community composition and ecosystem function, depending on
254 the type of stress factor and species traits [64]. Suttle et al. [65] found that sustained increased
255 precipitation eventually caused negative interactions among plant species that were not
256 apparent before the treatment. In alfalfa communities, Barton and Ives [66] found that reduced
257 precipitation changed interactions between spotted aphids and their ladybeetle predators
258 through dietary shifts of the latter. These examples make clear that species interactions
259 prevailing in the pre-change system cannot always be used to predict the chain of secondary
260 and higher-order effects occurring after the change. In such cases, knowledge about shifts in
261 per-capita species interactions is needed to predict how the selected environmental change
262 driver affects biodiversity and ecosystem function (Box 3, ‘Outstanding questions’).

263

264 Second, we have discussed environmental change drivers eliciting either positive or negative
265 responses that change monotonically as the level of environmental change increases, and stay
266 constant through time. However, many environmental change drivers can elicit positive
267 responses in some species but negative responses in others (e.g. temperature [37]), and many
268 responses are non-monotonic, with the sign of the response depending on the level of
269 environmental change (e.g. [67]). In addition, depending on the life history of the considered
270 species, populations can genetically adapt [68], which can alter their response to environmental

271 change through time. While these features do not threaten the general principle of our thesis,
272 they do indicate that community composition can be harder to predict, and therefore more
273 difficult to control, for certain combinations of environmental change drivers and ecosystem
274 types.

275

276 **6. Opportunities for ecosystem assessment and management**

277 Novel tools for biological monitoring will substantially increase the amount of biodiversity
278 data [69, 70]. However, linking monitored biodiversity trends to ecosystem functions remains
279 a major difficulty for ecosystem assessment, as has been discussed in the framework of several
280 environmental regulations worldwide [71, 72]. Re-introducing environmental change drivers
281 in B-EF research could help ecosystem managers by realistically translating observed
282 biodiversity trends to trends of ecosystem function for a suite of well-studied environmental
283 change drivers. Studies compiling and comparing different types of environmental change [21,
284 73] will be instrumental to ask if knowledge about one type of environmental change can be
285 transposed to other types of environmental change (Box 3, ‘Outstanding questions’). Following
286 ecosystem assessments, predicted changes of ecosystem functions could be used to inform
287 management as well, for example by triggering mitigating measures if needed. In addition,
288 ecosystem managers could propose critical levels of biodiversity change that, when exceeded,
289 lead to unacceptable loss of ecosystem functioning. The connection of B-EF research to applied
290 science has often been debated [74]. We conclude that re-introducing the use of environmental
291 change drivers to B-EF research can reinforce this connection.

292

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305 **Tables**

306 Table 1: Biodiversity manipulation can occur directly or indirectly and randomly or non-
 307 randomly ('Glossary'). Classic research on biodiversity and ecosystem function ('Classic B-
 308 EF') has focused on direct and random biodiversity manipulations. We provide four examples
 309 of environmental change drivers that cause negative (A and B) or positive (C and D) non-
 310 random direct effects in communities. Note that effects in case D are only initial effects, as
 311 explained in section 5. Direct non-random manipulations are rare and are proposed in section
 312 4 as a way forward to elucidate the role of biodiversity-mediated effects of environmental
 313 change on function. Indirect and random manipulations are rare as well but some chemical
 314 stressors can achieve such manipulations. Indirect biodiversity manipulations are by definition
 315 non-random with respect to species responses to environmental change.

316

	Direct	Indirect
Random	Common: Classic B-EF [6, 9]	Rare: Narcotic chemicals decrease survival of all species with lipid bilayers, and therefore manipulate biodiversity randomly with respect to function [54].
Non-random	Rare [7]: Proposed designs (section 4)	Common: A: Drought in streams <u>decreases</u> survival of large-bodied consumers and predators and reduces growth of encrusting green algae [35]. B: Pyrethroid insecticides in ditches <u>decrease</u> survival of arthropod predators [49, 50]. C: Nitrogen enrichment in grasslands <u>increases</u> growth of nitrogen demanding grasses [34]. D: Increased precipitation in grasslands <u>increases</u> growth of nitrogen-fixing forbs [65].

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323 **Box 1: Non-random and indirect vs. random and direct biodiversity manipulations.** We
324 simulate richness and ecosystem functioning in a community of 10 species responding to a
325 level l of an environmental change driver and contributing to an ecosystem function F :

$$326 \frac{dN_i}{dt} \cdot \frac{1}{N_i} = [\mu_i(l) + \sum_{j=1}^{10} \alpha_{i,j} \cdot N_j] \quad (\text{based on [44]})$$

$$327 F = \sum_{i=1}^{10} f_i(l) \cdot N_i^*$$

328 The $\alpha_{i,j}$ are per-capita effects of species j on species i ($\alpha_{i,j} = \alpha_{j,i} = -0.2$; intraspecific effects $\alpha_{i,i}$
329 are set to -1). N_i is the density of species i (asterisks denote equilibrium densities); $\mu_i(l)$ and
330 $f_i(l)$ are growth rates and per-capita contributions to F ('per-capita functioning') as a
331 function of l :

$$332 \mu_i(l) = \mu_{i,max} \cdot (1 + r_i \cdot l)$$

$$333 f_i(l) = f_{i,max} \cdot (1 + r_i/2 \cdot l)$$

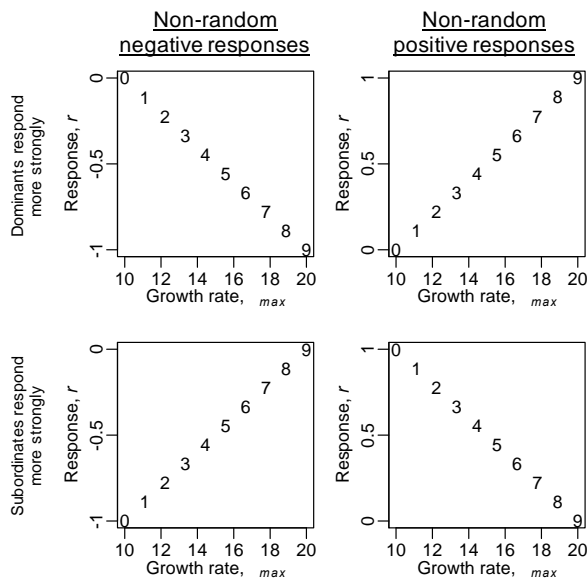
334 where r_i represents the response of species i to environmental change and the division by two
335 ensures per-capita functioning responds more strongly than density [75]. All species have
336 $f_{i,max} = 10$, respond differently to environmental change (Fig. I), have different growth rates
337 (Fig. I) and therefore different competitive strengths (Fig. II).

338 We manipulated richness indirectly and non-randomly by exposing the community to levels l
339 between 0 (no change) and 1 (100% increase or decrease of μ of the most responsive
340 species), and measured the corresponding F (Fig. 1, colored symbols). When dominants
341 respond most negatively (Fig. 1A), function decreases but richness is higher with than
342 without environmental change because of competitive release of species 0. Thus,
343 environmental change promotes co-existence and richness only decreases at high levels of
344 change. The resulting B-EF relationship is therefore non-monotonic. When environmental
345 change mostly elicits negative responses of subordinates (Fig. 1B), richness decreases already
346 at low levels of change because subordinates (species 1) combine a low density, which makes
347 them inherently prone to competitive exclusion, with a large negative response. In this case, a
348 monotonic positive B-EF relationship emerges. When environmental change elicits positive
349 responses, negative (Fig. 1C) or positive B-EF relationships (Fig. 1D) emerge from exactly
350 the same mechanisms as in Fig. 1A and 1B.

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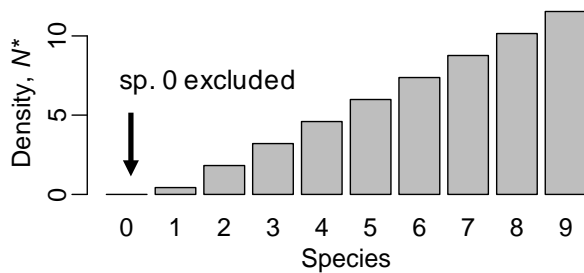
352 We also manipulated richness directly and randomly by removing all possible combinations
353 of 1 to 5 species from the community and measuring the corresponding F while setting $l=0$
354 (Fig. 1, shaded area, identical for all four scenarios).

355



356

357 Figure I: Environmental change elicits negative (left panels) or positive responses (right panels)
358 that are strongest for species with high (top row) or low (bottom row) growth rates, i.e. species
359 that are dominant and subordinate in pre-change conditions, respectively (Fig. II). Numbers
360 give species identity.



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362 Figure II: Equilibrium densities in absence of environmental change.

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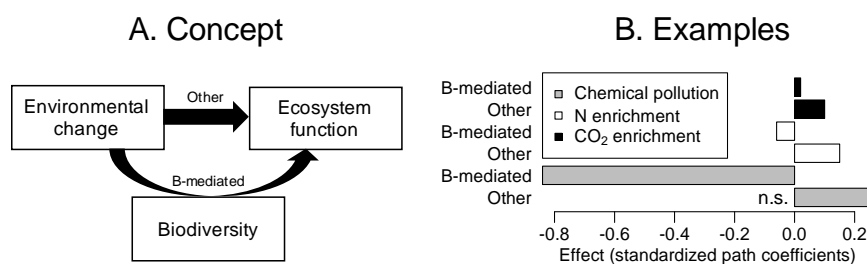
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374 **Box 2: Separating biodiversity-mediated effects on ecosystem functioning** Structural
 375 equation models (SEMs) can be used to compare biodiversity-mediated effects on ecosystem
 376 functioning with the other effects environmental change can have on function ('Glossary'). A
 377 SEM is described as "the use of two or more structural [cause-effect] equations to model
 378 multivariate relationships", which allows for an intuitive graphical representation of complex
 379 causal networks [58, 59]. Most notably, a SEM cannot only be used to isolate biodiversity-
 380 mediated effects on ecosystem functioning, but also to investigate the partial contributions of
 381 correlated explanatory variables to test alternative hypotheses [58].
 382 For illustrative purposes, we analysed data from a previously published microcosm study
 383 evaluating the effects of chemical pollution (a mixture of insecticides) on aquatic invertebrate
 384 richness and decomposition in a ditch community [76, 77] with a simple structural equation
 385 model. We also present previously published effects of nitrogen and carbon dioxide enrichment
 386 on plant richness and biomass production in grasslands [22]. These analyses show that
 387 richness-mediated effects are negative for environmental change drivers that have negative
 388 effects on richness and that this effect can only partly be compensated by other effects of
 389 environmental change. These analyses also show that the relative contribution of both types of
 390 effect differs among types of environmental change drivers. Many examples in the literature
 391 support the conclusion that cosm studies with chemical agents can be successfully analysed
 392 with SEMs, including SEMs with more extended effect pathways [20, 23]. In more replicated
 393 experimental setups [58], different biodiversity and community metrics could be tested in
 394 parallel to extract the most relevant biodiversity metric causing alterations in ecosystem
 395 functioning.



396
 397 Figure I. A: Environmental change drivers can affect functions by altering biodiversity or
 398 through other mechanisms ('Glossary') [22]. B: Structural equation models for three
 399 environmental change drivers. All effects are significant ($P < 0.05$) except when indicated
 400 (n.s.). The variance of diversity and function explained by the model (R^2) for the case of
 401 chemical pollution was 68% and 65%, respectively. Effects are standardized path coefficients

402 [59]. Details on the analysis for the other two drivers can be found in the original publication
403 [22].

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435 **Box 3: Outstanding questions**

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437 • Model simulations indicate that environmental change can affect function
438 without changing richness but how important are such effects in real ecosystems?

439 How do effects on function at invariant richness vary among ecosystems?

440 • Biodiversity-ecosystem functioning research has mostly focused on the effects
441 of random species loss on functions. How do these effects compare to those occurring
442 following environmental change?

443 • How does environmental change alter per-capita species interactions and how
444 does this affect our capacity to manipulate biodiversity using environmental change
445 drivers?

446 • How can knowledge about a selection of well-studied environmental change
447 drivers be used to manage ecosystems exposed to other types of environmental
448 change?

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468 **Glossary**

469 **Environmental change driver:** An environmental variable that exhibits long-term changes,
470 often as a result of anthropogenic activities. Examples include nutrient deposition, climate
471 warming, habitat destruction, and chemical stress.

472 **Direct biodiversity manipulation:** If biodiversity is manipulated directly, communities with
473 different biodiversity levels are composed, e.g. by taking different subsets of a species pool in
474 case of taxonomic richness.

475 **Indirect biodiversity manipulation:** If biodiversity is manipulated indirectly, one applies
476 different levels of an environmental change driver to create a biodiversity gradient. Indirect
477 biodiversity manipulations are by definition non-random with respect to species responses to
478 environmental change.

479 **Random biodiversity manipulation:** If biodiversity is manipulated randomly, community
480 composition is varied within a diversity level. By doing so, one can statistically control for
481 effects of community composition on ecosystem function.

482 **Non-random biodiversity manipulation:** Non-random biodiversity manipulations are done
483 based on known or presumed extinction or colonization orders (non-random with respect to
484 species responses to environmental change), or based on the contribution of species to function
485 (non-random with respect to species effects on ecosystem functions) [78].

486 **Biodiversity-mediated effect of environmental change on ecosystem function:** Effects
487 occurring through changes in any aspect of biodiversity (community composition or structure,
488 richness).

489 **Other effects of environmental change on ecosystem function:** Effects occurring through
490 mechanisms other than biodiversity changes. Examples include changes of total density
491 (community size), changes of per-capita functioning ($f_i(l)$ in Box 1, e.g. physiological
492 responses to warming), or changes of the bioavailability of macronutrients such as carbon,
493 nitrogen, or phosphorous [79].

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502 **Figure legends**

503 Figure 1: Simulations illustrating that indirect and non-random manipulations of biodiversity
504 can result in a multitude of biodiversity-ecosystem function relationships ('Resulting B-EF',
505 colours represent a scale from $l=0$ (yellow) to $l=1$ (red), the value for $l=0$ is indicated with a
506 'x' for clarity) by affecting not only richness but also per-capita functioning (average effect
507 across all species) and total density (sum of all species). These effects depend on the level of
508 environmental change ('level-dependent effects') and the shape of the resulting B-EF critically
509 depends on whether dominants (A and C) or subordinates (B and D) respond more strongly to
510 environmental change, and on whether the elicited responses are negative (A and B) or positive
511 (C and D). The shaded area indicates the expected B-EF under direct and random biodiversity
512 manipulations.

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514 Figure 2: Chemical stressors can be used to non-randomly and indirectly manipulate
515 biodiversity, as illustrated by empirically observed effects of continuous exposure of
516 freshwater ditch food-webs to chemical stressors in published micro- and mesocosm
517 experiments. A: Predators, herbivores and detritivores are separated into arthropod (Arth) and
518 non-arthropod (Non-arth) species; primary producers are separated into macrophytes (Macro)
519 and algae; Det. represents detrital material and its associated microflora. B: Results for
520 exposure to 50 μ g/L linuron, a photosystem ('PS') inhibitor [51, 52]. C: Results for exposure
521 to 35 μ g/L chlorpyrifos, a pyrethroid insecticide [49, 50]. Significant primary responses by the
522 corresponding chemical stressor are shown in red, secondary effects mediated by species
523 interactions are shown in green. White circles indicate that there was no effect. The relative
524 sizes of the coloured and dotted circles indicate whether the effect was positive (increase in
525 abundance - coloured circle larger than dotted circle) or negative (decrease of abundance -
526 coloured circle smaller than dotted circle).

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