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1 *Running head: Pre-dispersal seed predators alter plant nectar production*

2

3 **Seed predators can increase nectar volumes in an alpine daisy: but**
4 **do the insects benefit?**

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17

18

19 **Abstract**

20 Pollinators and pre-dispersal seed predators can interact via the plants they share. We
21 examined how pre-dispersal seed predators modify nectar quality and quantity and
22 thereby influence pollinator behavior. Working in a Tibetan alpine meadow, we
23 hypothesized that increasing levels of pre-dispersal seed predation by larvae of
24 tephritid flies would reduce nectar quantity and quality in *Saussurea nigrescens*
25 (Asteraceae), and that this would make the flowers less attractive to honeybees, the
26 most frequent floral visitors. Our field experiments showed that floret nectar volumes
27 responded differently to high and low densities of fly larvae, with significant
28 increases when there was one larva present, but decreases when two or more larvae
29 were present in a capitulum. Experimental manipulations of fly larvae yielded the
30 same result. The increases in nectar volume generated by a single larva are likely to
31 be beneficial for these insects in locations where pollinators preferentially visit those
32 *S. nigrescens* that produce more nectar. At our study sites, honeybees were the main
33 pollinators and visitation rates were unaffected by the changes in nectar volumes, but
34 they are introduced to the area, and native pollinators may be more selective.

35

36 **Key words** *Apis mellifera* • Asteraceae • Insect-plant

37 interactions • nectar • *Saussurea*, *Tephritis* • Tibetan Plateau

38

39

40 **Introduction**

41 Pollinators and flowering plants have contributed significantly to our understanding of
42 the evolution of mutualisms involving both generalist and specialist pollination
43 systems (Kjellberg et al. 2001; Lunau 2004; Mitchell et al. 2009; Bronstein 2015).
44 Co-evolution is most readily apparent in relation to the accessibility of rewards, with
45 pollinators exerting selective pressures on floral traits such as spur length (Nilsson
46 1988) and plants selecting for pollinator tongue length (Whittall and Hodges 2007;
47 Miller-Struttman et al. 2015). The quantity of accessible rewards thereby influences
48 which pollinators are attracted, and of which species, and can also influence how long
49 individual pollinators remain on each flower (Kalinganire 2001).

50

51 Plants and pollinators rarely if ever interact in isolation from other organisms, many
52 of which are likely to have antagonistic rather than mutualistic relationships with the
53 plants (Galen 1999). They include antagonists that can directly or indirectly influence
54 various traits through consumption of plant parts, such as decrease flower size (Barber
55 et al. 2012), modify nectar odour (Press and Phoenix 2005), reduce the quality of
56 floral displays (McCall and Irwin 2006), and reduce nectar production (McDade and
57 Kinsman 1980), all of which may influence pollinator behavior (Rodríguez-
58 Rodríguez et al. 2015).

59

60 Seed predatory insects often have a strong influence on plant reproductive success and
61 can generate significant selection pressures on their host plants (Kolb and Eriksson

62 2007). Their impact on their hosts varies in relation to plant reproductive traits such as
63 flowering phenology, flower number and seed size and these attributes are also
64 significant for the insects' own reproductive success, thereby facilitating co-
65 adaptation and co-evolution (Thompson 2005). Pre-dispersal seed predators require
66 seeds to feed their larvae, but often oviposit early in floral development, before ovules
67 have been pollinated. Their reproductive success is therefore influenced by the quality
68 and quantity of subsequent pollination events (Strauss and Irwin 2004). The adults of
69 some seed predators such as fig wasps and yucca moths ensure that seeds are
70 available for their larvae by routinely pollinating the flowers themselves (Pellmyr and
71 Huth 1994), but for the majority of species this is achieved by the selection of
72 oviposition sites in flowers or inflorescences that have a high probability of being
73 pollinated. Many pre-dispersal seed predators have a limited range of host plants, and
74 often just a single host species (Collin and Shykoff 2010). Within each plant species,
75 the likelihood and quality of pollination can depend on a wide range of environmental
76 and biological variables, but phenotypic characteristics of individual plants are also
77 significant. For insect-pollinated species, the size and quality of floral displays and the
78 quality of rewards they offer influence the extent of seed set (Thomson 1988;
79 Vaughton and Ramsey 1998). Consequently, the flowers that are most rewarding for
80 oviposition by seed predators are likely to be those that are also most favourable to
81 the plant's pollinators (Cariveau et al. 2004).

82

83 Seeds are nitrogen-rich and often contain high concentrations of energetically-

84 expensive defensive compounds (Janzen et al. 1977; Birch et al. 1986). Damage
85 generated by seed predators during oviposition or early larval feeding, together with
86 adaptive responses by the plants to the presence of the insects, can result in flowers
87 containing seed predators receiving less investment, with resources being switched to
88 undamaged flowers or retained elsewhere (Kudoh and Whigham 1998; Westerbergh
89 and Westerbergh 2001; Cariveau et al. 2004). Consequently, flowers containing seed
90 predators may be less attractive to pollinators and set fewer seeds (Cariveau et al.
91 2004).

92

93 Floral nectar is the most widely-provided reward for insect visitors to flowers (Scaven
94 and Rafferty 2013) and plants with more and higher quality nectar can attract more
95 pollinators (Mitchell 2004; Larsson and Franzen 2007; Wallis de Vries et al. 2012).

96 Nectar is composed mainly of sugars derived from photosynthesis and forms part of a
97 plant's overall carbohydrate content. Carbohydrates are stored in both reproductive
98 and vegetative organs (Pacini and Nepi 2007) and linkage has been recorded between
99 carbohydrate storage and nectar production, with more nectar produced at times of
100 day when overall carbohydrate content is higher (Mu et al. 2015). Herbivory often
101 reduces the quantity of reserves stored within plants (Machado et al. 2013), which
102 suggests it may also reduce nectar production, and indirectly influence pollinator
103 behavior to the further detriment of the plant.

104

105 Here, we describe the interplay between a pre-dispersal seed predator, the quantity and

106 quality of nectar in flowers of its host plant, the carbohydrates stored throughout the
107 plant and the willingness of bees to visit its flowers. We hypothesized that

108 (1) Pre-dispersal seed predators reduce the quantity and quality of nectar
109 in the flowers where they were feeding,

110 (2) Increasing numbers of seed predators have a progressively greater
111 impact on nectar production,

112 (3) Seed predators reduce the proportion of the plant's carbohydrates
113 stored in the flowers, relative to other parts of the plant, and

114 (4) Any changes in nectar rewards as a result of the seed predators
115 reduce flower visitation by the pollinators of the plant.

116

117 **Methods**

118 Natural History

119 *Saussurea nigrescens* (Asteraceae section Compositae, ECCAS 1999) is a
120 widespread high-altitude perennial distributed 2000-4300m on the Tibetan Plateau. Its
121 growing season is short, with plants reviving in mid-May and senescing by mid-
122 September. Mature plants vary in height about 15-45cm. Flowering occurs from July
123 to August and seeds mature and disperse in late-September. Each plant produces 2-5
124 dull-purple capitula, each of which contains 20-55 florets. The florets have an annular
125 bowl-shaped nectary between the ovary and anthers (bowl diameter is 1.5-2.0mm).
126 The flowers are monoclinal (with stamens and pistils in the same flower) but
127 outcrossing is favoured by protandry (stamens mature before the ovaries), see Mu et

128 al. (2014, 2015).

129 *Saussurea nigrescens* is the most economically important nectar-producing plant in
130 parts of the Tibetan plateau and contributes approximately 80% of the annual honey
131 harvest in Hongyuan County, China (Mu et al. 2014). The plant is allogamous and
132 requires cross-pollination, which is usually by honeybees (Mu et al. 2014).

133

134 The only recorded pre-dispersal seed predators of *S. nigrescens* are larvae of several
135 tephritid flies belonging to the genera *Campiglossa*, *Tephritis* and *Urophora* (Xi et al.
136 2016). Tephritid females oviposit into the flower buds of the host plant in the middle
137 of July. Their larvae develop within the capitula, where they consume developing
138 seeds and damage the receptacles (Xi et al. 2016). Between one and three larvae share
139 a capitulum. No obvious gall formation takes place. The tephritid larvae do not feed
140 on floral nectar, nor do they directly damage the nectaries. They pupate inside the
141 capitula, where they remain until the adults emerge the following spring.

142

143 Study sites

144 The study was conducted from 2014 to 2015 at the Hongyuan Alpine Meadow
145 Ecosystem Research Station of the Chinese Academy of Sciences, located in
146 Hongyuan County, Sichuan Province, China (32°48'-32°52'N, 102°01'-102°33'E).
147 This area of the eastern Qinghai-Tibetan Plateau is at an altitude of about 3500m.
148 With short and cool spring, summer and autumn seasons and a long cold winter. Liu et
149 al. (2012) and Mu et al. (2015) provided details of the local climate, soil conditions

150 and vegetation. The meadows are sometimes used for cattle grazing during the winter,
151 but are otherwise undisturbed. *Tephritis femoralis* Chen is the common tephritid seed
152 predator of *S. nigrescens* locally.

153

154 In late July 2014 we selected three sites (sites 1-3) located about 5 km apart that had
155 similar soil moisture and physical and chemical properties (Mu et al. 2014). Their
156 plant communities were also similar. *Saussurea nigrescens* was the most abundant
157 species at the sites, contributing 33-46% of total plant cover during its flowering
158 period. *Elymus nutans* Griseb, *Anemone rivularis* Buch.-Ham. ex DC. and *Potentilla*
159 *anserina* L. were also common. The three sites were at roughly similar distances from
160 the nearest apiaries and similar numbers of bees and adult tephritid flies were
161 recorded visiting the flowers of *S. nigrescens* (Mu et al. 2014). *Saussurea nigrescens*
162 was the major source of nectar for bees at all three study sites (Mu et al. 2014). Native
163 pollinators of *S. nigrescens* include an Asian honeybee (*Apis cerana*) and the
164 bumblebees *Bombus filchnerae*, *B. humilis*, and *B. supremus* (Macior et al. 2001). The
165 introduced *Apis mellifera* has been tended by beekeepers in the area since 1981 (Sun
166 et al. 2013).

167

168 Field records of tephritid numbers and nectar variables

169 In July 2014 we tagged 50 healthy *S. nigrescens* with undamaged leaves at each
170 of the three sites. After the tephritids had an opportunity to oviposit, but before the
171 flowers had opened, we enclosed all the capitula on each plant within fine mesh

172 netting to exclude further insect visitors (Real and Rathcke 1991). This plant produces
173 its maximum nectar volumes when the anthers are white and emerging from the
174 capitula (Mu et al. 2014). When they reached this stage, we selected five florets at
175 random from each of the capitula and on sunny days at 10:00-15:00Hrs we measured
176 their nectar volumes and concentrations. Nectar volumes were measured using 1 µl or
177 5 µl micropipettes (Hirschmann Laborgeräte, Germany). Nectar concentrations were
178 measured concurrently with a hand-held refractometer (Eclipse, Stanley Company,
179 England Bellingham, UK) following the methods of Johnson et al. (2006). The
180 numbers of tephritid larvae inside each capitulum were then counted using a binocular
181 microscope. A total of 472 capitula were selected to monitor nectar volume and
182 concentration.

183

184 Plant responses to experimental manipulations

185 In July 2015 we selected 120 vigorous and undamaged *S. nigrescens* growing at
186 site 2 and enclosed groups of six plants within cylindrical steel netting enclosures
187 (100cm diameter × 100cm high, mesh size 0.1 mm × 0.1 mm). When flower buds
188 were at the stage suitable for tephritid oviposition, all but one randomly chosen
189 capitulum on each plant were removed. One, two or three *Tephritis femoralis*
190 (Tephritidae) larvae (about 2mm in length) were then inserted into the remaining
191 capitula. The control group was physically manipulated in the same way as the other
192 groups, but no larvae were inserted. The larvae were obtained as described by Xi et al.
193 (2016). The netting remained in place until the capitula reached peak nectar

194 production, when floret nectar volumes and concentrations were recorded as before. A
195 total of 120 capitula were selected to monitor nectar volume and concentration.

196

197 After nectar production had been monitored, each plant was removed and the lengths
198 and fresh weights of capitula, leaves, stems and roots were weighed in the field using
199 a 0.001g capacity balance. The plant components were then stored immediately in a
200 portable icebox cooled with dry ice and their soluble sugar and starch contents were
201 measured later following the procedures of Hansen et al. (1975) and Yoshida et al.
202 (1976). The relative amounts of carbohydrates (soluble sugars and starch, mg/g) in the
203 capitula, leaves, stems and roots were expressed as a percentage of the total
204 carbohydrates in each plant. Carbohydrate contents were calculated as in Rivera-Solís
205 et al. (2012). A total of 120 capitula were selected to monitor the carbohydrate
206 contents.

207

208 Honeybee responses to experimental manipulations

209 Twelve 2 × 2m × 1m high enclosures covered in the steel netting described above
210 were erected at site 2 to enclose groups of *S. nigrescens* with recently-developed
211 flower buds. Once they reached the stage suitable for tephritid oviposition 30
212 vigorous, undamaged plants in each plot had all but one of their capitula removed and
213 zero, one, two or three tephritid larvae were inserted as before. Any additional plants
214 in the enclosures had all their capitula removed, leaving a consistent 30 capitula
215 within each 4 m² plot. To monitor honeybee visitation, we first removed the netting

216 screens. Six operatives then simultaneously recorded honeybee visits to individual
217 capitula in pairs of adjacent plots every hour (each plot was observed for 30 minutes
218 per hour) at 9:00 -17:00 on three sunny days in July 2015. Observers were located
219 about 3m from each plot, which allowed for clear observation of pollinator behavior.
220 Honeybee visitation rates to each capitulum per hour were calculated following the
221 protocol of Arroyo et al. (1985). We then harvested the plants and measured fresh
222 weights as before. The plant parts were then dried to constant mass and re-weighed to
223 the nearest 0.001g.

224

225 Statistical analyses

226 All statistical analyses were performed in R (R Development Core Team 2013,
227 URL <http://www.R-project.org>). Plant traits were first tested for normality using the
228 Shapiro-Wilk test and for homogeneity of variance using Levene's test. Floret nectar
229 volume was log₁₀-transformed and the proportion of flowering plants per subplot were
230 arcsine-transformed to achieve normality. Correlation analyses were used to
231 determine the relationship between floret nectar volume and capitula carbohydrate
232 contents. Pollinator visitation rates were assessed using one-way ANOVA followed by
233 Tukey's test.

234

235 We used general linear-mixed models (GLMMs) to assess the effects of pre-dispersal
236 seed predator numbers and capitula mass on variation in floret nectar volume and
237 concentration, capitulum carbohydrates and total carbohydrates storage as percentages

238 of dry and wet weights. To account for interaction effects between pre-dispersal seed
239 predator and capitulum size, we used a model with fly numbers and capitulum mass
240 as fixed factors, and site and above-ground vegetative mass as random factors. For the
241 mixed models, we used the *lme* function in the *lme4* package in R (Bates et al. 2011).

242

243

Results

244 *Saussurea nigrescens* colonized naturally by tephritid larvae produced different
245 volumes of nectar depending on how many larvae were present, but there was no
246 simple relationship between numbers of these seed predators and nectar volume (Fig.
247 1, Table 1, Appendix A). Capitula containing a single tephritid larva produced more
248 nectar than controls, whereas capitula containing two or three larvae produced
249 significantly less nectar. The presence of one larva increase nectar volumes by an
250 average of 15.1%, but two or three larvae led to decreases of 76.9% and 83.1%
251 respectively (Fig. 1). In contrast to nectar volume, the tephritids had no effect on
252 nectar concentration (Appendix A). Very similar results were obtained when tephritid
253 numbers were manipulated experimentally (Fig. 1, Table 1). With one larva added,
254 nectar increased by 17.0% relative to controls, and when two or three larvae were
255 added nectar volumes fell by 77.8% and 79.4% respectively. Nectar concentrations
256 were again unchanged (Appendix A).

257 The starch and soluble sugar contents of *S. nigrescens* capitula showed the same
258 response to tephritid larvae as nectar volume, with elevated carbohydrates compared
259 with controls in capitula containing one larva and significantly lower concentrations

260 of carbohydrates when two or three larvae were present (Fig.2). Across treatments
261 there was a significant positive correlation between capitulum carbohydrate
262 concentrations and average nectar volume, with indications that the same pattern was
263 also present within capitula that contained one larva (Fig. 3). The changes in
264 capitulum carbohydrate concentrations in response to the tephritids reflected changes
265 in the relative distributions of carbohydrates within the plants (Fig. 4). A single larva
266 was associated with an increase in the percentage of the plant's total carbohydrates
267 stored within the capitula, relative to controls, but two or three larvae were associated
268 with declines in the capitula carbohydrates (Fig. 4; $t = -2.44$, $P < 0.05$). Corresponding
269 changes were present in the proportion of carbohydrates stored in the stems, with
270 declines when one larva was present, and increases when there were more larvae (Fig.
271 4, $t = 1.99$, $P < 0.05$). There were no treatment effects on the proportion of the plants'
272 carbohydrates in the leaves and roots (Fig. 4, $t = -0.56$, $P = 0.58$ and $t = 0.86$, $P = 0.39$,
273 respectively).

274

275 Despite the changes in nectar volumes associated with the tephritid larvae, they had
276 no significant impact on the visitation rates of honeybees to the capitula ($F = 0.903$,
277 $P = 0.388$, Appendix B).

278

279

Discussion

280

281

We hypothesized that pre-dispersal seed predators would reduce floret nectar
production and nectar concentrations in proportion to their densities, and that this

282 would lead to a decline in pollinator visitation rates. These hypotheses were not
283 supported by the data from both field records and experimental manipulations. Floret
284 nectar volumes did decline in capitula that contained two or three tephritid larvae, but
285 volumes increased significantly in capitula with one larva. Furthermore, nectar
286 concentrations were consistently independent of seed predator numbers, as were
287 visitation rates by honeybees. Although carbohydrate storage in the capitula was
288 influenced by the presence of the tephritid larvae, it was a non-linear response that
289 mirrored nectar production, with capitula containing a single larva having a higher
290 proportion of the plant's carbohydrates than controls with no seed predators.

291

292 Rates of nectar production and secretion are related to the quantities of soluble
293 sugars and starch available and thereby to carbohydrate reserves and rates of
294 photosynthesis (Búrquez and Corbet 1991; Pacini et al. 2003). Physical conditions
295 such as elevated night-time temperatures, and damage to leaves and flowers can all
296 reduce nectar production (Wäckers et al. 2001; Mu et al. 2015). In our study system
297 the positive relationship between local carbohydrate storage and floret nectar volume
298 was particularly clear, because they increased and decreased together in response to
299 varying densities of tephritid larvae. Relative to un-occupied capitula, carbohydrate
300 storage increased by 19.4% and floret nectar volumes increase by 17.0% in capitula
301 with one larva, but were reduced by 30.3% and 32.6% respectively when more larvae
302 were present. Carbohydrate storage elsewhere on the plants reflected these changes.

303

304 The non-linear carbohydrate storage and nectar volume responses by the plants
305 to the presence of insects feeding in their capitula can be interpreted in terms of
306 adaptive responses by the plants in the face of varying levels of damage, but also as
307 reflecting manipulations by the insects. The increase in capitula carbohydrate storage
308 and nectar volumes when a single larva is present could be a plant compensation
309 effect (Rivera-Solís et al. 2012). Plants attacked by herbivores often allocate more
310 resources to their reproductive organs (reviewed by Trumble et al. 1993), but
311 increased allocation to reproduction, including elevation of nectar quality, is most
312 evident when vegetative structures are damaged (Inouye 1982; Lanza 1988; Smith et
313 al. 1990).

314

315 An alternative explanation for the increase in nectar volumes produced by
316 capitula occupied by a single tephritid larva is that the insects are actively inducing
317 this effect, and that increased nectar volumes favour their own reproductive success,
318 rather than that of their host plants. The tephritid larvae feed on developing seeds, and
319 depend on capitula being visited by pollinators after their eggs have been laid. Any
320 increase in the likelihood of those capitula being adequately pollinated, such as might
321 be the case by increasing nectar volumes, would therefore be to the advantage of the
322 insect. Many of the species of tephritids with larvae that develop in the capitula of
323 Asteraceae are gall formers (Varley 1947; Straw 1989). This includes congeners of the
324 *Tephritis* species from *S. nigrescens* (Goeden et al. 1988). Gall forming insects
325 routinely generate nutrient sinks that draw in resources from elsewhere on their host

326 plants (Price et al. 1987). Although *T. femoralis* does not generate clearly-defined
327 galls in the capitula of *S. nigrescens*, its larvae may be capable of initiating some of
328 the plant physiological effects associated with galling, including stimulation of local
329 carbohydrate storage. This may have direct beneficial effects in terms of food quality
330 for developing larvae, and indirect benefits via larger nectar volumes leading to
331 increased numbers of seed in the capitula it occupies.

332

333 Quite different carbohydrate storage and nectar volume responses were recorded
334 in capitula where more than one tephritid larva was present. The declines seen in
335 capitula containing multiple larvae are likely to reflect the damage they cause to the
336 receptacles, through which nutrients and metabolites are transported (Teuber et al.
337 1983; Rivera-Solís et al. 2012). Damage to the receptacles was more frequent and
338 much more extensive when two or more larvae were sharing a capitulum, with more
339 than 90% of the receptacles damaged in capitula with two or three larvae, compared
340 with only around 10% of the receptacles when only a single larva was present (J Mu
341 unpublished data). The increased feeding on the receptacles in capitula with several
342 larvae may be the result of antagonistic behavior among the fly larvae and a shortage
343 of seeds remaining to be eaten (Averill and Prokopy 1987).

344

345 Seed predators are expected to preferentially oviposit on flowers that are more
346 attractive to pollinators, because these flowers are most likely to set seed (Strauss and
347 Irwin 2004). Competition is often intense among seed predators sharing what are

348 usually finite and spatially-limited resources. Only 15% of the *S. nigrescens* capitula
349 at our study site that contained tephritid larvae had more than one larva present (X XI
350 unpublished data), suggesting that females typically lay a single egg on each plant.
351 This oviposition pattern not only reduces the likelihood of competition, but also
352 means that most of the larvae are developing in capitula that are producing more
353 nectar than adjacent capitula that lack the insects.

354

355 Contrary to expectations that the insects would gain from developing in capitula
356 with higher nectar volumes, capitula with one larva present did not attract more
357 honeybees than unoccupied capitula. This result was contrary to much of the
358 literature, which suggests that floral nectar volumes have a positive relationship with
359 pollinator visitation (Schemske and Bradshaw 1999; Pyke 1982, 2016). Different bee
360 species may respond differently to aspects of nectar rewards, such as nectar volume
361 and concentration. For example, honeybees have been shown to prefer flowers that
362 have high sugar concentrations (Scheiner et al. 1999, 2001; Vaudo et al. 2015),
363 whereas bumblebees are sensitive to nectar volume (Harder and Real 1987). In this
364 study, we found no changes in nectar concentration (sugar content) resulting from the
365 pre-dispersal seed predators, and this may have led to the similar honeybee visitation
366 rates. We did not examine visitation rates by native bees such as bumblebees and they
367 may be more responsive to nectar volumes than honeybees. Our future investigations
368 will examine whether pre-dispersal seed predators have different effects on the
369 behavior of honeybees and native bees.

370

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378

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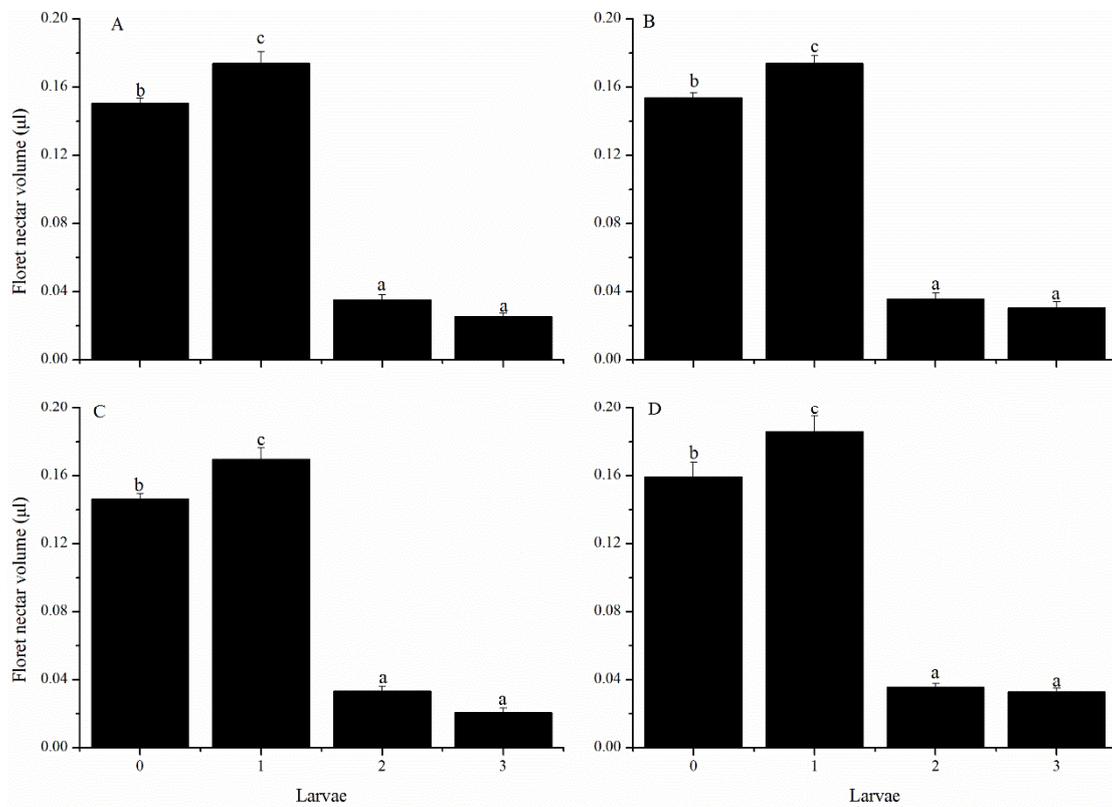
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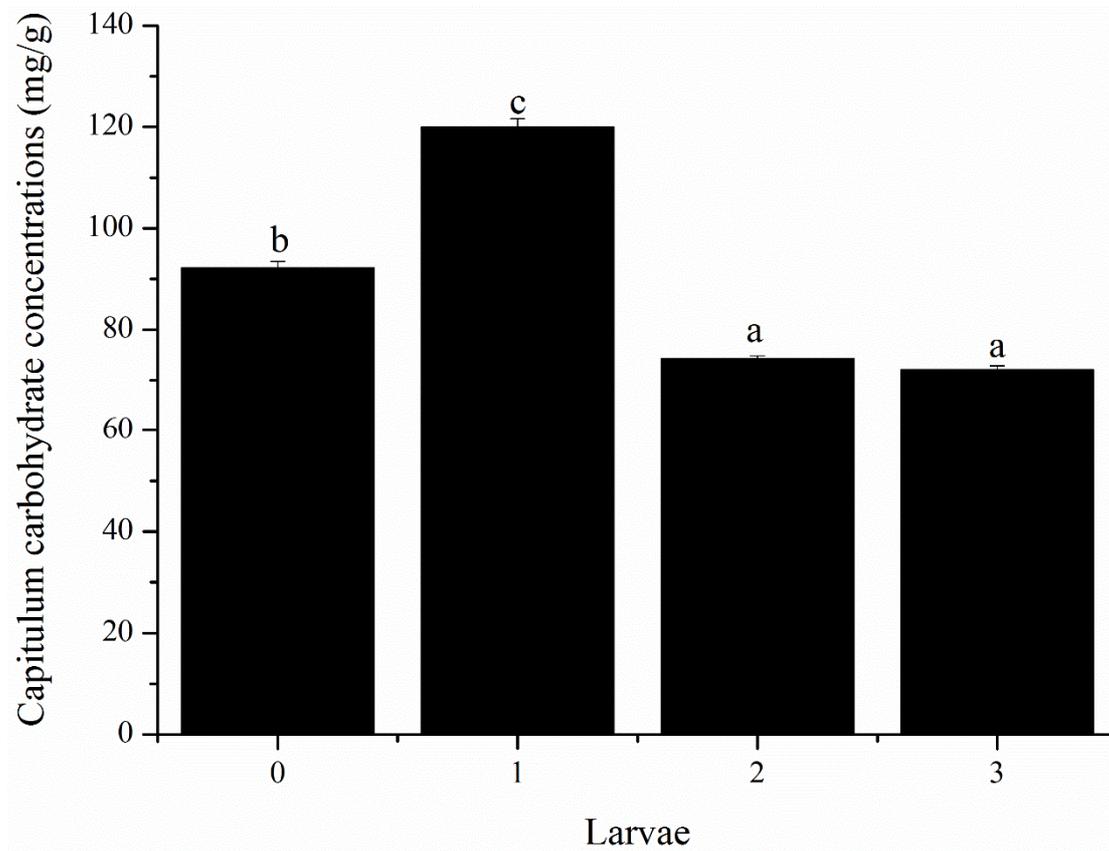
1 Figure 1. Variation in *Saussurea nigrescens* nectar volumes in relation to the numbers
 2 of tephritid larvae in their capitula (Means \pm SEs per floret). Figures 1A-1C plants
 3 growing at sites 1-3 with natural variation in larval numbers, Figure 1D
 4 experimentally introduced tephritid larvae (site 2 only). Different letters above
 5 columns indicate differences within sites at $P < 0.05$ (one-way analysis of variance).

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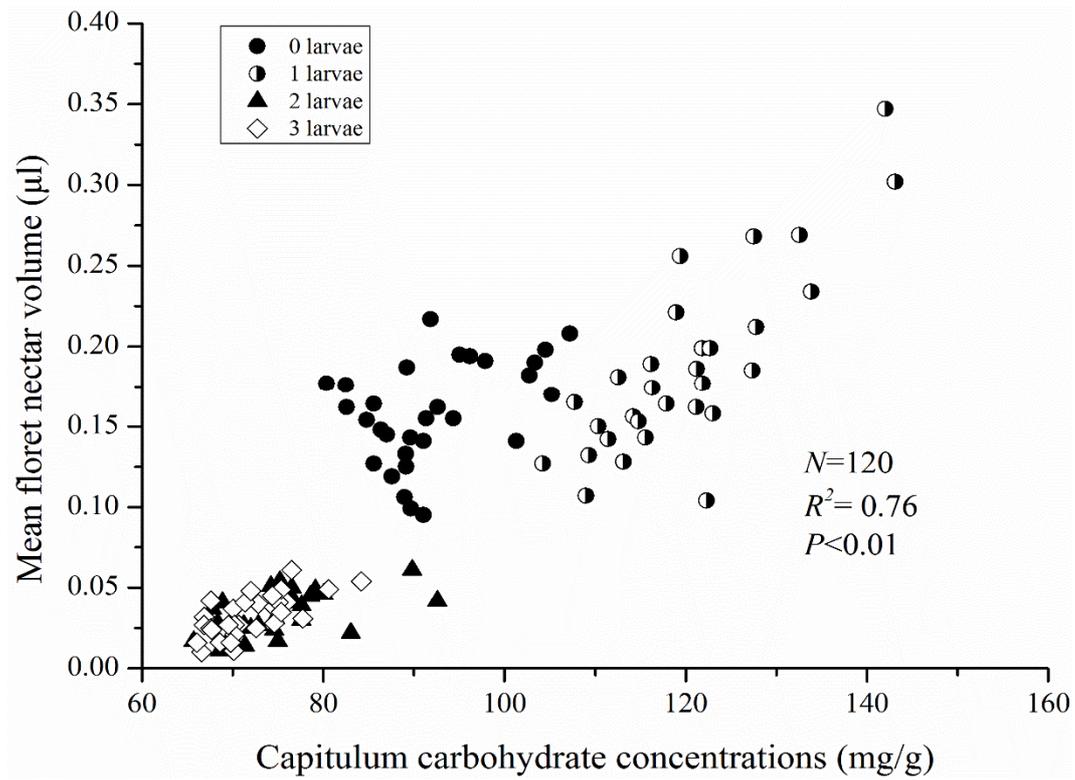
10 Figure 2. Capitulum carbohydrate concentrations of *S. nigrescens* that had contained
11 varying numbers of tephritid larvae (Means \pm 1 SE). Different letters above columns
12 indicate differences at $P < 0.05$ (one-way analysis of variance).



13
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15 Figure 3. The relationship between *Saussurea nigrescens* capitulum carbohydrate
16 concentrations and mean floret nectar volume in capitula where different numbers of
17 tephritid larvae had been introduced. N = 30 florets for each larval density.

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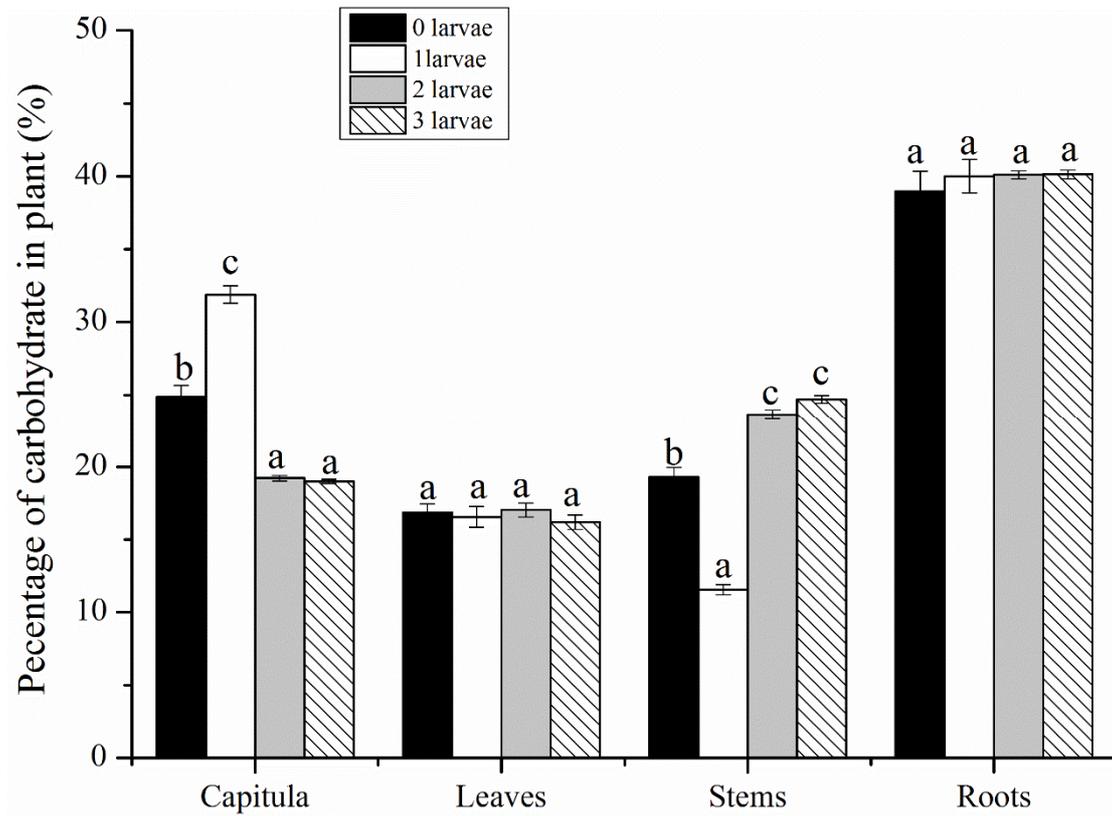
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23 Figure 4. Percentage of total carbohydrates (soluble sugars and starch, mg/g) in
 24 capitula, leaves, stems and roots of *S. nigrescens* (means \pm 1 SE). Different letters
 25 above columns indicate differences between parts of the plants at $P < 0.05$ (one-way
 26 analysis of variance).



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Table 1. Summary of linear mixed effect models comparing the effects of pre-dispersal seed predators and capitulum size on floret nectar volume, nectar concentration, and capitulum carbohydrate concentrations. AIC = Akaike Information Criterion and BIC = Bayesian Information Criterion. The numbers after Floret nectar volume and Nectar concentration are AIC and BIC, respectively.

Variables /AIC/ BIC	Fixed effects			
	value	SE	t-value	p-value
Field conditions				
Floret nectar volume /-1539.99/-1515.10				
Intercept	0.159	0.009	18.119	<0.001
Pre-dispersal seed predators (P)	-0.028	0.011	-2.441	0.015
Capitulum mass (C)	-0.050	0.054	0.929	0.353
P×C	-0.097	0.069	-1.403	0.161
Nectar concentration/3449.41/3474.30				
Intercept	39.977	2.000	19.984	<0.001
Pre-dispersal seed predators(P)	-1.568	2.388	-0.657	0.512
Capitula mass (C)	10.411	12.789	0.814	0.416
P×C	8.517	14.612	0.583	0.560
Experimental manipulations				
Floret nectar volume /-338.37/-321.85				
Intercept	0.159	0.018	8.983	<0.001
Pre-dispersal seed predators (P)	-0.046	0.010	-4.587	<0.001
Capitula mass (C)	0.158	0.106	1.495	0.138
P×C	-0.046	0.061	-0.759	0.449
Nectar concentration/747.25/763.77				
Intercept	41.630	1.909	21.802	<0.001
Pre-dispersal seed predators (P)	-0.055	1.083	-0.051	0.959
Capitulum mass (C)	-2.152	11.400	-0.189	0.851
P×C	-1.901	6.548	-0.290	0.772
Capitulum carbohydrate concentrations /1016.50/1033.02				
Intercept	187.545	46.583	4.026	<0.001
Pre-dispersal seed predators (P)	-8.602	3.921	-2.194	0.030
Capitulum mass (C)	16.649	30.306	0.549	0.584
P×C	-1.391	2.544	-0.545	0.587

Appendix list

Appendix A

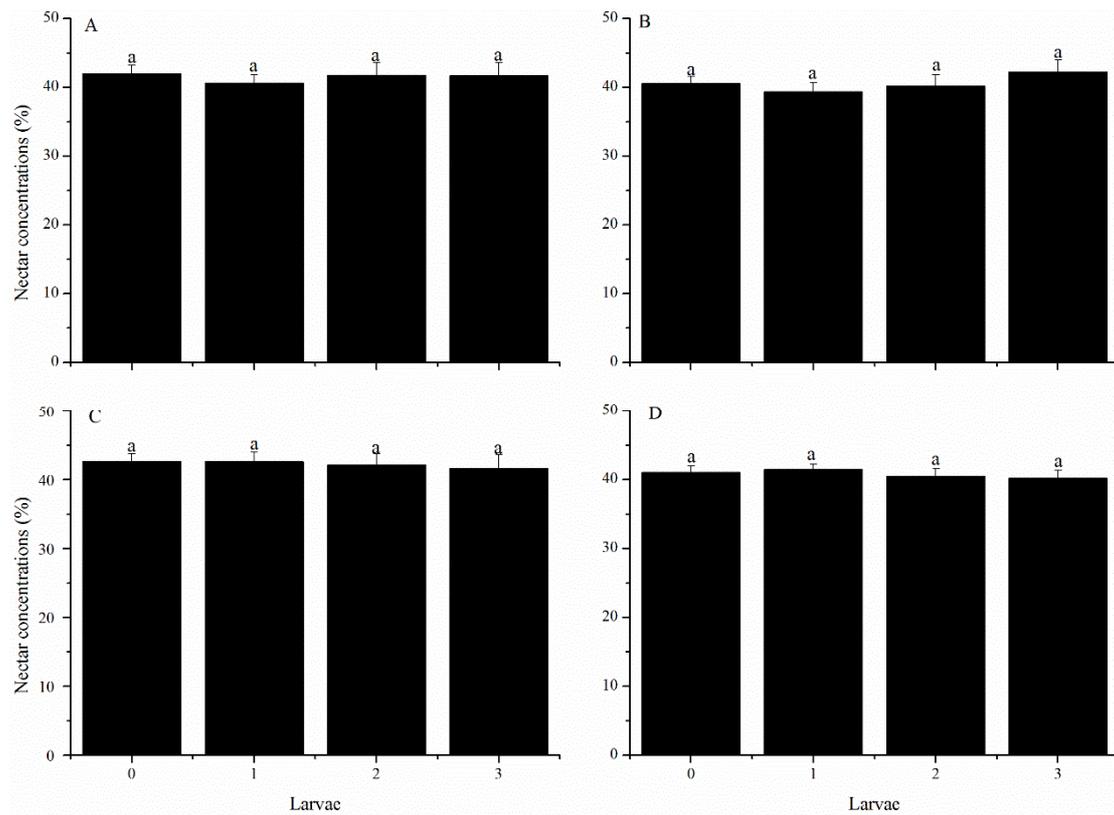
S. nigrescens nectar concentrations at sites 1 (A), 2 (B), and 3 (C) under natural condition and after experimental introductions (D).

Appendix B

Apis mellifera visitation rates per capitulum of *Saussurea nigrescens* in relation to tephritid larval numbers.

Appendix A

S. nigrescens nectar concentrations with natural variation in numbers of tephritid larvae at sites 1-3 (A-C) and in the larval-addition experiment at site 2 (D). Similar letters above columns indicate no differences within sites at $P < 0.05$ (one-way analysis of variance).



Appendix B

Apis mellifera visits to capitula of *Saussurea nigrescens* that contained different numbers of tephritid larvae (means \pm 1 SE). Capitulum visitation rates were compared by one-way ANOVAs followed by Tukey's tests. Similar letters above columns indicate no difference in visitation rates at $P < 0.05$ (one-way analysis of variance).

