



UNIVERSITY OF LEEDS

This is a repository copy of *Seed predators can increase nectar volumes in an alpine daisy: but do the insects benefit?*.

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/108502/>

Version: Accepted Version

---

**Article:**

Mu, J, Chen, Y, Yang, Y et al. (3 more authors) (2016) Seed predators can increase nectar volumes in an alpine daisy: but do the insects benefit? *Plant Ecology*, 217 (10). pp. 1195-1205. ISSN 1385-0237

<https://doi.org/10.1007/s11258-016-0646-1>

---

© Springer Science+Business Media Dordrecht 2016. This is an author produced version of a paper published in *Plant Ecology*. Uploaded in accordance with the publisher's self-archiving policy. The final publication is available at Springer via <https://doi.org/10.1007/s11258-016-0646-1>.

**Reuse**

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

1 *Running head: Pre-dispersal seed predators alter plant nectar production*

2

3 **Seed predators can increase nectar volumes in an alpine daisy: but**  
4 **do the insects benefit?**

5 Junpeng Mu<sup>1\*</sup>, Yan Chen<sup>1</sup>, Yulian Yang<sup>1</sup>, Ronghua Fu<sup>2</sup>, Hui Wang<sup>1</sup>, Stephen G.

6 Compton<sup>3,4</sup>

7

8 *<sup>1</sup>Ecological Security and Protection Key Laboratory of Sichuan Province, Mianyang*

9 *Normal University, Mianyang 621000, China*

10 *<sup>2</sup>Key Laboratory of Southwest China Wildlife Resources Conservation (Ministry of*

11 *Education), China West Normal University, Nanchong 637002, China*

12 *<sup>3</sup>School of Biology, University of Leeds, Leeds LS2 9JT, United Kingdom*

13 *<sup>4</sup>Department of Zoology and Entomology, Rhodes University, Grahamstown 6140,*

14 *South Africa*

15

16 \* Email: [gbmujp@163.com](mailto:gbmujp@163.com)

17

18

19 **Abstract**

20 Pollinators and pre-dispersal seed predators can interact via the plants they share. We  
21 examined how pre-dispersal seed predators modify nectar quality and quantity and  
22 thereby influence pollinator behavior. Working in a Tibetan alpine meadow, we  
23 hypothesized that increasing levels of pre-dispersal seed predation by larvae of  
24 tephritid flies would reduce nectar quantity and quality in *Saussurea nigrescens*  
25 (Asteraceae), and that this would make the flowers less attractive to honeybees, the  
26 most frequent floral visitors. Our field experiments showed that floret nectar volumes  
27 responded differently to high and low densities of fly larvae, with significant  
28 increases when there was one larva present, but decreases when two or more larvae  
29 were present in a capitulum. Experimental manipulations of fly larvae yielded the  
30 same result. The increases in nectar volume generated by a single larva are likely to  
31 be beneficial for these insects in locations where pollinators preferentially visit those  
32 *S. nigrescens* that produce more nectar. At our study sites, honeybees were the main  
33 pollinators and visitation rates were unaffected by the changes in nectar volumes, but  
34 they are introduced to the area, and native pollinators may be more selective.

35

36 **Key words** *Apis mellifera* • Asteraceae • Insect-plant

37 interactions • nectar • *Saussurea*, *Tephritis* • Tibetan Plateau

38

39

40 **Introduction**

41 Pollinators and flowering plants have contributed significantly to our understanding of  
42 the evolution of mutualisms involving both generalist and specialist pollination  
43 systems (Kjellberg et al. 2001; Lunau 2004; Mitchell et al. 2009; Bronstein 2015).  
44 Co-evolution is most readily apparent in relation to the accessibility of rewards, with  
45 pollinators exerting selective pressures on floral traits such as spur length (Nilsson  
46 1988) and plants selecting for pollinator tongue length (Whittall and Hodges 2007;  
47 Miller-Struttman et al. 2015). The quantity of accessible rewards thereby influences  
48 which pollinators are attracted, and of which species, and can also influence how long  
49 individual pollinators remain on each flower (Kalinganire 2001).

50

51 Plants and pollinators rarely if ever interact in isolation from other organisms, many  
52 of which are likely to have antagonistic rather than mutualistic relationships with the  
53 plants (Galen 1999). They include antagonists that can directly or indirectly influence  
54 various traits through consumption of plant parts, such as decrease flower size (Barber  
55 et al. 2012), modify nectar odour (Press and Phoenix 2005), reduce the quality of  
56 floral displays (McCall and Irwin 2006), and reduce nectar production (McDade and  
57 Kinsman 1980), all of which may influence pollinator behavior (Rodríguez-  
58 Rodríguez et al. 2015).

59

60 Seed predatory insects often have a strong influence on plant reproductive success and  
61 can generate significant selection pressures on their host plants (Kolb and Eriksson

62 2007). Their impact on their hosts varies in relation to plant reproductive traits such as  
63 flowering phenology, flower number and seed size and these attributes are also  
64 significant for the insects' own reproductive success, thereby facilitating co-  
65 adaptation and co-evolution (Thompson 2005). Pre-dispersal seed predators require  
66 seeds to feed their larvae, but often oviposit early in floral development, before ovules  
67 have been pollinated. Their reproductive success is therefore influenced by the quality  
68 and quantity of subsequent pollination events (Strauss and Irwin 2004). The adults of  
69 some seed predators such as fig wasps and yucca moths ensure that seeds are  
70 available for their larvae by routinely pollinating the flowers themselves (Pellmyr and  
71 Huth 1994), but for the majority of species this is achieved by the selection of  
72 oviposition sites in flowers or inflorescences that have a high probability of being  
73 pollinated. Many pre-dispersal seed predators have a limited range of host plants, and  
74 often just a single host species (Collin and Shykoff 2010). Within each plant species,  
75 the likelihood and quality of pollination can depend on a wide range of environmental  
76 and biological variables, but phenotypic characteristics of individual plants are also  
77 significant. For insect-pollinated species, the size and quality of floral displays and the  
78 quality of rewards they offer influence the extent of seed set (Thomson 1988;  
79 Vaughton and Ramsey 1998). Consequently, the flowers that are most rewarding for  
80 oviposition by seed predators are likely to be those that are also most favourable to  
81 the plant's pollinators (Cariveau et al. 2004).

82

83 Seeds are nitrogen-rich and often contain high concentrations of energetically-

84 expensive defensive compounds (Janzen et al. 1977; Birch et al. 1986). Damage  
85 generated by seed predators during oviposition or early larval feeding, together with  
86 adaptive responses by the plants to the presence of the insects, can result in flowers  
87 containing seed predators receiving less investment, with resources being switched to  
88 undamaged flowers or retained elsewhere (Kudoh and Whigham 1998; Westerbergh  
89 and Westerbergh 2001; Cariveau et al. 2004). Consequently, flowers containing seed  
90 predators may be less attractive to pollinators and set fewer seeds (Cariveau et al.  
91 2004).

92

93 Floral nectar is the most widely-provided reward for insect visitors to flowers (Scaven  
94 and Rafferty 2013) and plants with more and higher quality nectar can attract more  
95 pollinators (Mitchell 2004; Larsson and Franzen 2007; Wallis de Vries et al. 2012).

96 Nectar is composed mainly of sugars derived from photosynthesis and forms part of a  
97 plant's overall carbohydrate content. Carbohydrates are stored in both reproductive  
98 and vegetative organs (Pacini and Nepi 2007) and linkage has been recorded between  
99 carbohydrate storage and nectar production, with more nectar produced at times of  
100 day when overall carbohydrate content is higher (Mu et al. 2015). Herbivory often  
101 reduces the quantity of reserves stored within plants (Machado et al. 2013), which  
102 suggests it may also reduce nectar production, and indirectly influence pollinator  
103 behavior to the further detriment of the plant.

104

105 Here, we describe the interplay between a pre-dispersal seed predator, the quantity and

106 quality of nectar in flowers of its host plant, the carbohydrates stored throughout the  
107 plant and the willingness of bees to visit its flowers. We hypothesized that

108 (1) Pre-dispersal seed predators reduce the quantity and quality of nectar  
109 in the flowers where they were feeding,

110 (2) Increasing numbers of seed predators have a progressively greater  
111 impact on nectar production,

112 (3) Seed predators reduce the proportion of the plant's carbohydrates  
113 stored in the flowers, relative to other parts of the plant, and

114 (4) Any changes in nectar rewards as a result of the seed predators  
115 reduce flower visitation by the pollinators of the plant.

116

## 117 **Methods**

### 118 Natural History

119 *Saussurea nigrescens* (Asteraceae section Compositae, ECCAS 1999) is a  
120 widespread high-altitude perennial distributed 2000-4300m on the Tibetan Plateau. Its  
121 growing season is short, with plants reviving in mid-May and senescing by mid-  
122 September. Mature plants vary in height about 15-45cm. Flowering occurs from July  
123 to August and seeds mature and disperse in late-September. Each plant produces 2-5  
124 dull-purple capitula, each of which contains 20-55 florets. The florets have an annular  
125 bowl-shaped nectary between the ovary and anthers (bowl diameter is 1.5-2.0mm).  
126 The flowers are monoclinal (with stamens and pistils in the same flower) but  
127 outcrossing is favoured by protandry (stamens mature before the ovaries), see Mu et

128 al. (2014, 2015).

129 *Saussurea nigrescens* is the most economically important nectar-producing plant in  
130 parts of the Tibetan plateau and contributes approximately 80% of the annual honey  
131 harvest in Hongyuan County, China (Mu et al. 2014). The plant is allogamous and  
132 requires cross-pollination, which is usually by honeybees (Mu et al. 2014).

133

134 The only recorded pre-dispersal seed predators of *S. nigrescens* are larvae of several  
135 tephritid flies belonging to the genera *Campiglossa*, *Tephritis* and *Urophora* (Xi et al.  
136 2016). Tephritid females oviposit into the flower buds of the host plant in the middle  
137 of July. Their larvae develop within the capitula, where they consume developing  
138 seeds and damage the receptacles (Xi et al. 2016). Between one and three larvae share  
139 a capitulum. No obvious gall formation takes place. The tephritid larvae do not feed  
140 on floral nectar, nor do they directly damage the nectaries. They pupate inside the  
141 capitula, where they remain until the adults emerge the following spring.

142

143 Study sites

144 The study was conducted from 2014 to 2015 at the Hongyuan Alpine Meadow  
145 Ecosystem Research Station of the Chinese Academy of Sciences, located in  
146 Hongyuan County, Sichuan Province, China (32°48'-32°52'N, 102°01'-102°33'E).  
147 This area of the eastern Qinghai-Tibetan Plateau is at an altitude of about 3500m.  
148 With short and cool spring, summer and autumn seasons and a long cold winter. Liu et  
149 al. (2012) and Mu et al. (2015) provided details of the local climate, soil conditions



150 and vegetation. The meadows are sometimes used for cattle grazing during the winter,  
151 but are otherwise undisturbed. *Tephritis femoralis* Chen is the common tephritid seed  
152 predator of *S. nigrescens* locally.

153

154 In late July 2014 we selected three sites (sites 1-3) located about 5 km apart that had  
155 similar soil moisture and physical and chemical properties (Mu et al. 2014). Their  
156 plant communities were also similar. *Saussurea nigrescens* was the most abundant  
157 species at the sites, contributing 33-46% of total plant cover during its flowering  
158 period. *Elymus nutans* Griseb, *Anemone rivularis* Buch.-Ham. ex DC. and *Potentilla*  
159 *anserina* L. were also common. The three sites were at roughly similar distances from  
160 the nearest apiaries and similar numbers of bees and adult tephritid flies were  
161 recorded visiting the flowers of *S. nigrescens* (Mu et al. 2014). *Saussurea nigrescens*  
162 was the major source of nectar for bees at all three study sites (Mu et al. 2014). Native  
163 pollinators of *S. nigrescens* include an Asian honeybee (*Apis cerana*) and the  
164 bumblebees *Bombus filchnerae*, *B. humilis*, and *B. supremus* (Macior et al. 2001). The  
165 introduced *Apis mellifera* has been tended by beekeepers in the area since 1981 (Sun  
166 et al. 2013).

167

168 Field records of tephritid numbers and nectar variables

169 In July 2014 we tagged 50 healthy *S. nigrescens* with undamaged leaves at each  
170 of the three sites. After the tephritids had an opportunity to oviposit, but before the  
171 flowers had opened, we enclosed all the capitula on each plant within fine mesh

172 netting to exclude further insect visitors (Real and Rathcke 1991). This plant produces  
173 its maximum nectar volumes when the anthers are white and emerging from the  
174 capitula (Mu et al. 2014). When they reached this stage, we selected five florets at  
175 random from each of the capitula and on sunny days at 10:00-15:00Hrs we measured  
176 their nectar volumes and concentrations. Nectar volumes were measured using 1 µl or  
177 5 µl micropipettes (Hirschmann Laborgeräte, Germany). Nectar concentrations were  
178 measured concurrently with a hand-held refractometer (Eclipse, Stanley Company,  
179 England Bellingham, UK) following the methods of Johnson et al. (2006). The  
180 numbers of tephritid larvae inside each capitulum were then counted using a binocular  
181 microscope. A total of 472 capitula were selected to monitor nectar volume and  
182 concentration.

183

#### 184 Plant responses to experimental manipulations

185 In July 2015 we selected 120 vigorous and undamaged *S. nigrescens* growing at  
186 site 2 and enclosed groups of six plants within cylindrical steel netting enclosures  
187 (100cm diameter × 100cm high, mesh size 0.1 mm × 0.1 mm). When flower buds  
188 were at the stage suitable for tephritid oviposition, all but one randomly chosen  
189 capitulum on each plant were removed. One, two or three *Tephritis femoralis*  
190 (Tephritidae) larvae (about 2mm in length) were then inserted into the remaining  
191 capitula. The control group was physically manipulated in the same way as the other  
192 groups, but no larvae were inserted. The larvae were obtained as described by Xi et al.  
193 (2016). The netting remained in place until the capitula reached peak nectar

194 production, when floret nectar volumes and concentrations were recorded as before. A  
195 total of 120 capitula were selected to monitor nectar volume and concentration.

196

197 After nectar production had been monitored, each plant was removed and the lengths  
198 and fresh weights of capitula, leaves, stems and roots were weighed in the field using  
199 a 0.001g capacity balance. The plant components were then stored immediately in a  
200 portable icebox cooled with dry ice and their soluble sugar and starch contents were  
201 measured later following the procedures of Hansen et al. (1975) and Yoshida et al.  
202 (1976). The relative amounts of carbohydrates (soluble sugars and starch, mg/g) in the  
203 capitula, leaves, stems and roots were expressed as a percentage of the total  
204 carbohydrates in each plant. Carbohydrate contents were calculated as in Rivera-Solís  
205 et al. (2012). A total of 120 capitula were selected to monitor the carbohydrate  
206 contents.

207

208 Honeybee responses to experimental manipulations

209 Twelve 2 × 2m × 1m high enclosures covered in the steel netting described above  
210 were erected at site 2 to enclose groups of *S. nigrescens* with recently-developed  
211 flower buds. Once they reached the stage suitable for tephritid oviposition 30  
212 vigorous, undamaged plants in each plot had all but one of their capitula removed and  
213 zero, one, two or three tephritid larvae were inserted as before. Any additional plants  
214 in the enclosures had all their capitula removed, leaving a consistent 30 capitula  
215 within each 4 m<sup>2</sup> plot. To monitor honeybee visitation, we first removed the netting

216 screens. Six operatives then simultaneously recorded honeybee visits to individual  
217 capitula in pairs of adjacent plots every hour (each plot was observed for 30 minutes  
218 per hour) at 9:00 -17:00 on three sunny days in July 2015. Observers were located  
219 about 3m from each plot, which allowed for clear observation of pollinator behavior.  
220 Honeybee visitation rates to each capitulum per hour were calculated following the  
221 protocol of Arroyo et al. (1985). We then harvested the plants and measured fresh  
222 weights as before. The plant parts were then dried to constant mass and re-weighed to  
223 the nearest 0.001g.

224

#### 225 Statistical analyses

226 All statistical analyses were performed in R (R Development Core Team 2013,  
227 URL <http://www.R-project.org> ). Plant traits were first tested for normality using the  
228 Shapiro-Wilk test and for homogeneity of variance using Levene's test. Floret nectar  
229 volume was log<sub>10</sub>-transformed and the proportion of flowering plants per subplot were  
230 arcsine-transformed to achieve normality. Correlation analyses were used to  
231 determine the relationship between floret nectar volume and capitula carbohydrate  
232 contents. Pollinator visitation rates were assessed using one-way ANOVA followed by  
233 Tukey's test.

234

235 We used general linear-mixed models (GLMMs) to assess the effects of pre-dispersal  
236 seed predator numbers and capitula mass on variation in floret nectar volume and  
237 concentration, capitulum carbohydrates and total carbohydrates storage as percentages

238 of dry and wet weights. To account for interaction effects between pre-dispersal seed  
239 predator and capitulum size, we used a model with fly numbers and capitulum mass  
240 as fixed factors, and site and above-ground vegetative mass as random factors. For the  
241 mixed models, we used the *lme* function in the *lme4* package in R (Bates et al. 2011).

242

243

## Results

244 *Saussurea nigrescens* colonized naturally by tephritid larvae produced different  
245 volumes of nectar depending on how many larvae were present, but there was no  
246 simple relationship between numbers of these seed predators and nectar volume (Fig.  
247 1, Table 1, Appendix A). Capitula containing a single tephritid larva produced more  
248 nectar than controls, whereas capitula containing two or three larvae produced  
249 significantly less nectar. The presence of one larva increase nectar volumes by an  
250 average of 15.1%, but two or three larvae led to decreases of 76.9% and 83.1%  
251 respectively (Fig. 1). In contrast to nectar volume, the tephritids had no effect on  
252 nectar concentration (Appendix A). Very similar results were obtained when tephritid  
253 numbers were manipulated experimentally (Fig. 1, Table 1). With one larva added,  
254 nectar increased by 17.0% relative to controls, and when two or three larvae were  
255 added nectar volumes fell by 77.8% and 79.4% respectively. Nectar concentrations  
256 were again unchanged (Appendix A).

257 The starch and soluble sugar contents of *S. nigrescens* capitula showed the same  
258 response to tephritid larvae as nectar volume, with elevated carbohydrates compared  
259 with controls in capitula containing one larva and significantly lower concentrations

260 of carbohydrates when two or three larvae were present (Fig.2). Across treatments  
261 there was a significant positive correlation between capitulum carbohydrate  
262 concentrations and average nectar volume, with indications that the same pattern was  
263 also present within capitula that contained one larva (Fig. 3). The changes in  
264 capitulum carbohydrate concentrations in response to the tephritids reflected changes  
265 in the relative distributions of carbohydrates within the plants (Fig. 4). A single larva  
266 was associated with an increase in the percentage of the plant's total carbohydrates  
267 stored within the capitula, relative to controls, but two or three larvae were associated  
268 with declines in the capitula carbohydrates (Fig. 4;  $t = -2.44$ ,  $P < 0.05$ ). Corresponding  
269 changes were present in the proportion of carbohydrates stored in the stems, with  
270 declines when one larva was present, and increases when there were more larvae (Fig.  
271 4,  $t = 1.99$ ,  $P < 0.05$ ). There were no treatment effects on the proportion of the plants'  
272 carbohydrates in the leaves and roots (Fig. 4,  $t = -0.56$ ,  $P = 0.58$  and  $t = 0.86$ ,  $P = 0.39$ ,  
273 respectively).

274

275 Despite the changes in nectar volumes associated with the tephritid larvae, they had  
276 no significant impact on the visitation rates of honeybees to the capitula ( $F = 0.903$ ,  
277  $P = 0.388$ , Appendix B).

278

## 279 **Discussion**

280 We hypothesized that pre-dispersal seed predators would reduce floret nectar  
281 production and nectar concentrations in proportion to their densities, and that this

282 would lead to a decline in pollinator visitation rates. These hypotheses were not  
283 supported by the data from both field records and experimental manipulations. Floret  
284 nectar volumes did decline in capitula that contained two or three tephritid larvae, but  
285 volumes increased significantly in capitula with one larva. Furthermore, nectar  
286 concentrations were consistently independent of seed predator numbers, as were  
287 visitation rates by honeybees. Although carbohydrate storage in the capitula was  
288 influenced by the presence of the tephritid larvae, it was a non-linear response that  
289 mirrored nectar production, with capitula containing a single larva having a higher  
290 proportion of the plant's carbohydrates than controls with no seed predators.

291

292 Rates of nectar production and secretion are related to the quantities of soluble  
293 sugars and starch available and thereby to carbohydrate reserves and rates of  
294 photosynthesis (Búrquez and Corbet 1991; Pacini et al. 2003). Physical conditions  
295 such as elevated night-time temperatures, and damage to leaves and flowers can all  
296 reduce nectar production (Wäckers et al. 2001; Mu et al. 2015). In our study system  
297 the positive relationship between local carbohydrate storage and floret nectar volume  
298 was particularly clear, because they increased and decreased together in response to  
299 varying densities of tephritid larvae. Relative to un-occupied capitula, carbohydrate  
300 storage increased by 19.4% and floret nectar volumes increase by 17.0% in capitula  
301 with one larva, but were reduced by 30.3% and 32.6% respectively when more larvae  
302 were present. Carbohydrate storage elsewhere on the plants reflected these changes.

303

304       The non-linear carbohydrate storage and nectar volume responses by the plants  
305 to the presence of insects feeding in their capitula can be interpreted in terms of  
306 adaptive responses by the plants in the face of varying levels of damage, but also as  
307 reflecting manipulations by the insects. The increase in capitula carbohydrate storage  
308 and nectar volumes when a single larva is present could be a plant compensation  
309 effect (Rivera-Solís et al. 2012). Plants attacked by herbivores often allocate more  
310 resources to their reproductive organs (reviewed by Trumble et al. 1993), but  
311 increased allocation to reproduction, including elevation of nectar quality, is most  
312 evident when vegetative structures are damaged (Inouye 1982; Lanza 1988; Smith et  
313 al. 1990).

314

315       An alternative explanation for the increase in nectar volumes produced by  
316 capitula occupied by a single tephritid larva is that the insects are actively inducing  
317 this effect, and that increased nectar volumes favour their own reproductive success,  
318 rather than that of their host plants. The tephritid larvae feed on developing seeds, and  
319 depend on capitula being visited by pollinators after their eggs have been laid. Any  
320 increase in the likelihood of those capitula being adequately pollinated, such as might  
321 be the case by increasing nectar volumes, would therefore be to the advantage of the  
322 insect. Many of the species of tephritids with larvae that develop in the capitula of  
323 Asteraceae are gall formers (Varley 1947; Straw 1989). This includes congeners of the  
324 *Tephritis* species from *S. nigrescens* (Goeden et al. 1988). Gall forming insects  
325 routinely generate nutrient sinks that draw in resources from elsewhere on their host



326 plants (Price et al. 1987). Although *T. femoralis* does not generate clearly-defined  
327 galls in the capitula of *S. nigrescens*, its larvae may be capable of initiating some of  
328 the plant physiological effects associated with galling, including stimulation of local  
329 carbohydrate storage. This may have direct beneficial effects in terms of food quality  
330 for developing larvae, and indirect benefits via larger nectar volumes leading to  
331 increased numbers of seed in the capitula it occupies.

332

333       Quite different carbohydrate storage and nectar volume responses were recorded  
334 in capitula where more than one tephritid larva was present. The declines seen in  
335 capitula containing multiple larvae are likely to reflect the damage they cause to the  
336 receptacles, through which nutrients and metabolites are transported (Teuber et al.  
337 1983; Rivera-Solís et al. 2012). Damage to the receptacles was more frequent and  
338 much more extensive when two or more larvae were sharing a capitulum, with more  
339 than 90% of the receptacles damaged in capitula with two or three larvae, compared  
340 with only around 10% of the receptacles when only a single larva was present (J Mu  
341 unpublished data). The increased feeding on the receptacles in capitula with several  
342 larvae may be the result of antagonistic behavior among the fly larvae and a shortage  
343 of seeds remaining to be eaten (Averill and Prokopy 1987).

344

345       Seed predators are expected to preferentially oviposit on flowers that are more  
346 attractive to pollinators, because these flowers are most likely to set seed (Strauss and  
347 Irwin 2004). Competition is often intense among seed predators sharing what are

348 usually finite and spatially-limited resources. Only 15% of the *S. nigrescens* capitula  
349 at our study site that contained tephritid larvae had more than one larva present (X XI  
350 unpublished data), suggesting that females typically lay a single egg on each plant.  
351 This oviposition pattern not only reduces the likelihood of competition, but also  
352 means that most of the larvae are developing in capitula that are producing more  
353 nectar than adjacent capitula that lack the insects.

354

355         Contrary to expectations that the insects would gain from developing in capitula  
356 with higher nectar volumes, capitula with one larva present did not attract more  
357 honeybees than unoccupied capitula. This result was contrary to much of the  
358 literature, which suggests that floral nectar volumes have a positive relationship with  
359 pollinator visitation (Schemske and Bradshaw 1999; Pyke 1982, 2016). Different bee  
360 species may respond differently to aspects of nectar rewards, such as nectar volume  
361 and concentration. For example, honeybees have been shown to prefer flowers that  
362 have high sugar concentrations (Scheiner et al. 1999, 2001; Vaudo et al. 2015),  
363 whereas bumblebees are sensitive to nectar volume (Harder and Real 1987). In this  
364 study, we found no changes in nectar concentration (sugar content) resulting from the  
365 pre-dispersal seed predators, and this may have led to the similar honeybee visitation  
366 rates. We did not examine visitation rates by native bees such as bumblebees and they  
367 may be more responsive to nectar volumes than honeybees. Our future investigations  
368 will examine whether pre-dispersal seed predators have different effects on the  
369 behavior of honeybees and native bees.

370

371 **Acknowledgements**

372 We thank Xinqiang Xi, Xinwei Wu, Jie Xiong, Hongli Chen, Yongping Li, Xu Luo,  
373 Xue Wei, Rui Cao, Hanxiang Chen and Kai He for their field and laboratory  
374 assistance. This study was funded by National Science Foundation of China  
375 (31270513, 31270387), Innovative Research Team by Sichuan Provincial Department  
376 of Education (13TD0015) and Project of Introduced Excellent Talents by Mianyang  
377 Normal University (QD2012A07).

378

379 **References**

380 Arroyo MTK, Armesto JJ, Primack RB (1985) Community studies in pollination  
381 ecology in the high temperate Andes of central Chile. II. Effect of temperature on  
382 visitation rates and pollination possibilities. *Plant Syst. Evol.* 149: 187-203.

383 Averill AL, Prokopy RJ (1987) Intraspecific competition in the tephritid fruit fly  
384 *Rhagoletis pomonella*. *Ecology* 68: 878-886.

385 Barber NA, Adler LS, Theis N, Hazzard RV, Kiers ET (2012) Herbivory reduces plant  
386 interactions with above- and belowground antagonists and mutualists. *Ecology* 93:  
387 1560-1570.

388 Bates D, Maechler M, Bolker B (2011) Lme 4: Linear mixed-effects models using S4  
389 classes. R package version 0.999375-38

390 Birch ANE, Fellows LE, Evans SV, Doherty K (1986). Para-aminophenylalanine in  
391 *Vigna*: possible taxonomic and ecological significance as a seed defence against

392 bruchids. *Phytochemistry* 25: 2745-2749.

393 Bronstein JL. (Ed.) (2015) *Mutualism*. Oxford University Press, Oxford 297 pp.

394 Búrquez A, Corbet SA (1991) Do flowers reabsorb nectar? *Funct Ecol* 5: 369-379.

395 Cariveau, D. et al. 2004. Direct and indirect effects of pollinators and seed predators to  
396 selection on plant and floral traits. *Oikos* 104: 15-26.

397 Collin CL and Shykoff JA (2010) Flowering phenology and female fitness: impact of  
398 a pre-dispersal seed predator on a sexually polymorphic species. *Plant Ecol* 206:  
399 1-13.

400 ECCAS (Editor Committee of the Chinese Academy of Sciences for Flora of China)  
401 (1999) *Flora of China*. Beijing: Science Press (in Chinese) 78: 38.

402 Galen C (1999) Why do flowers vary? The functional ecology of variation in flower  
403 size and form within natural plant population. *Bioscience* 49: 631-640.

404 Goeden RD (1988) Gall formation by the capitulum-infesting fruit fly, *Tephritis*  
405 *stigmatica* (Diptera: Tephritidae). *P Entomol Soc Wash* 90: 37-43.

406 Hansen J and Møller IB (1975) Percolation of starch and soluble carbohydrates from  
407 plant tissue for quantitative determination with anthrone. *Anal Biochem* 68: 87-  
408 94.

409 Harder LD, Real LA (1987) Why are bumble bees risk averse? *Ecology* 68: 1104-1108.

410 Janzen DH, Juster HB, Bell EA (1977) Toxicity of secondary compounds to the seed-  
411 eating larvae of the bruchid beetle *Callosobruchus macutatus*. *Phytochemistry*  
412 16: 223-227.

413 Johnson SD, Hargreaves AL, Brown M (2006) Dark, bitter-tasting nectar functions as

414 filter of flower visitors in a bird-pollinated plant. *Ecology* 87: 2706-2716.

415 Kalinganire A, Harwood CE, Slee MU, Simons A J (2001) Pollination and fruit-set of  
416 *Grevillea robusta* in western Kenya. *Austral Ecol* 26: 637-648.

417 Kjellberg F, Jousselein E, Bronstein JL, Patel AJ, Yokoyama J, Rasplus JY (2001)  
418 Pollination mode in fig wasps: The predictive power of correlated traits. *P Roy*  
419 *Soc B Bio* 268: 1113-1121.

420 Kolb A, Ehrlén J, Eriksson O (2007) Ecological and evolutionary consequences of  
421 spatial and temporal variation in pre-dispersal seed predation. *Perspect Plant Ecol*  
422 9: 79-100.

423 Kudoh H and Whigham DF (1998) The effect of petal size manipulation on  
424 pollinator/seed-predator mediated female reproductive success of *Hibiscus*  
425 *moscheutos*. *Oecologia* 117: 70 -79.

426 Lanza J (1988) Ant preferences for *Passiflora* nectar mimics that contain amino acids.  
427 *Biotropica* 20: 341-344.

428 Larsson M and Franzen M (2007) Critical resource levels of pollen for the declining  
429 bee *Andrena hattorfiana* (Hymenoptera, *Andrenidae*). *Biol Conserv* 134: 405-414.

430 Liu YZ, Mu JP, Li GY, Sun SC (2012) Global warming may reduce plant reproductive  
431 effort for temperate multi-flowered species. *New Phytol* 195: 427-436.

432 Lunau K (2004) Adaptive radiation and coevolution — pollination biology case studies.  
433 *Org Divers Evol* 4: 207-224.

434 Machado RAR, Ferrieri AP, Robert CA, Glauser G, Kallenbach M, Baldwin IT, Erb M  
435 (2013) Leaf-herbivore attack reduces carbon reserves and regrowth from the roots

436 via jasmonate and auxin signaling. *New Phytol* 200: 1234-1246.

437 Macior LW, Tang Y, Zhang JC (2001) Reproductive biology of *Pedicularis*  
438 (Scrophulariaceae) in the Sichuan Himalaya. *Plant Spec Biol* 16: 83-89.

439 McCall AC and Irwin RE (2006) Florivory: The intersection of pollination and  
440 herbivory. *Ecol Lett* 9: 1351-1365.

441 McDade LA and Kinsman S (1980) The impact of floral parasitism in two Neotropical  
442 hummingbird-pollinated plant species. *Evolution* 34: 944-958.

443 Miller-Struttman NE, Geib JC, Frankin JD, Kevan PG, Holdo RM et al (2015)  
444 Functional mismatch in a bumble bee pollination mutualism under climate change.  
445 *Science* 349: 1541-1544.

446 Mitchell RJ, Karron JD, Holmquist KJ, Bell JM (2004) The influence of *Mimulus*  
447 *ringens* floral display size on pollinator visitation patterns. *Funct Ecol* 18: 116-  
448 124.

449 Mitchell RJ, Irwin RE, Flanagan RJ, and Karron JD (2009) Ecology and evolution of  
450 plant-pollinator interactions. *Ann Bot-London* 103: 1355-1364.

451 Mu JP, Peng YH, Xi XQ, Wu XW, Griffin JN, Niklas KJ, Su SC (2014) Domesticated  
452 honeybees evolutionarily reduce flower nectar volume in a Tibetan Asteraceae.  
453 *Ecology* 95: 3161-3172.

454 Mu JP, Peng YH, Xi XQ, Wu XW, Griffin JN, Niklas KJ, Sun SC (2015) Artificial  
455 asymmetric warming dramatically reduces nectar production in a Tibetan lotus.  
456 *Ann Bot-London* 116: 899-906.

457 Nilsson LA (1988) The evolution of flowers with deep corolla tubes. *Nature* 334: 147-

458 149.

459 Pacini E and Nepi M (2007) Nectar production and presentation. In Nectaries and  
460 Nectar. Nicolson SW, Nepi M, Pacini E (eds). Dordrecht, the Netherlands: Springer,  
461 pp.167-214.

462 Pacini E, Nepi M, and Vesprini JL (2003) Nectar biodiversity: a short review. *Plant*  
463 *Syst Evol* 238: 7 -21.

464 Pellmyr O and Huth CJ (1994) Evolutionary stability of mutualism between yuccas and  
465 yucca moths. *Nature* 372: 257-260.

466 Press CA and Phoenix GK (2005) Impacts of parasitic plants on natural communities.  
467 *New Phytol* 166: 737-751.

468 Price PW, Fernandes GW, Waring GL (1987) Adaptive Nature of Insect Galls *Environ*  
469 *Entomol* 16: 15-24.

470 Pyke GH. 1982. Foraging in bumblebees: rule of departure from an inflorescence. *Can*  
471 *J Zool* 60: 417-428.

472 Pyke GH. 2016. Floral nectar: pollinator attraction or manipulation. *Trends Ecol Evol*  
473 31: 339-341.

474 Real LA and Rathcke BJ (1991) Individual variation in nectar production and its effects  
475 on fitness in *Kalmia latifolia*. *Ecology* 72: 149-155.

476 Rivera-Solís G, Abdala-Roberts L, Cervera JC, Parra-Tabla V, Ruiz-Ruiz J, Betancur-  
477 Ancona D (2012) Mechanisms and traits associated with compensation for  
478 defoliation in *Ruellia nudiflora*. *Plant Ecol* 213: 303-314.

479 Rodríguez- Rodríguez MC, Jordano P, and Valido A (2015) Hotspots of damage by

480 antagonists shape the spatial structure of plant-pollinator interactions. *Ecology* 96:  
481 2181-2191.

482 Scaven V and Rafferty NE (2013) Physiological effects of climate warming on  
483 flowering plants and insect pollinator and potential consequences for their  
484 interaction. *Curr Zool* 59: 418-426.

485 Scheiner R, Erber J, Page RE (1999) Tactile learning and the individual evaluation of  
486 the reward in honeybees (*Apis mellifera* L.). *J Comp Physiol A* 185: 1-10.

487 Scheiner R, Page RE, Erber J (2001) Responsiveness to sucrose affects tactile and  
488 olfactory learning in preforaging honeybees of two genetic strains. *Behav Brain*  
489 *Res* 120: 67-73.

490 Schemske DW, Bradshaw HD (1999) Pollinator preference and the evolution of floral  
491 traits in monkeyflowers (*Mimulus*). *P Natl Acad Sci USA* 96: 11910-11915.

492 Smith LL, Lanza J, Smith GC (1990) Amino acid concentrations in extrafloral nectar  
493 of *Impatiens sultani* increase after stimulated herbivory. *Ecology* 71: 107-115.

494 Strauss SY and Irwin RE (2004) Ecological and evolutionary consequences of  
495 multispecies plant–animal interactions. *Annu Rev Ecol Evol S* 35: 435-466.

496 Straw NA (1989) Taxonomy, attack strategies and host relations in flowerhead  
497 Tephritidae: a review. *Ecol Entomol* 14: 455-462.

498 Sun SG, Huang SQ, Guo YH (2013) Pollinator shift to managed honeybees enhances  
499 reproductive output in a bumblebee-pollinated plant. *Plant Syst Evol* 299: 139-150.

500 Thompson JN (2005) *The Geographic Mosaic of Coevolution*. University of Chicago  
501 Press, Chicago.



502 Thomson JD (1988) Effects of variation in inflorescence size and floral rewards on the  
503 visitation rates of traplining pollinators of *Aralia hispida*. *Evol Ecol* 2: 65-76.

504 Teuber LR, Barnes DK, Rincker CM (1983) Effectiveness of selection for nectar  
505 volume, receptacle diameter, and seed yield characteristics in Alfalfa. *Crop Sci* 23:  
506 283-289.

507 Trumble JT (1993) Plant compensation for arthropod herbivory. *Annu Rev Entomol* 38:  
508 93-119.

509 Vaudo AD, Tooker JF, Grozinger CM, Patch HM (2015) Bee nutrition and floral  
510 resource restoration. *Curr Opin Insect Sci* 10: 133-141.

511 Varley GC (1947) The natural control of population balance in the Knapweed Gall-Fly  
512 (*Urophora jaceana*). *J Anim Ecol* 16: 139-187.

513 Vaughton G and Ramsey M (1998) Floral display, pollinator visitation and reproductive  
514 success in the dioecious perennial herb *Wurmbea dioica* (Liliaceae). *Oecologia*  
515 115: 93-101.

516 Wäckers FL, Zuber D, Wunderlin R, Keller F (2001) The effect of herbivory on  
517 temporal and spatial dynamics of foliar nectar production in cotton and castor. *Ann*  
518 *Bot-London* 87: 365-370.

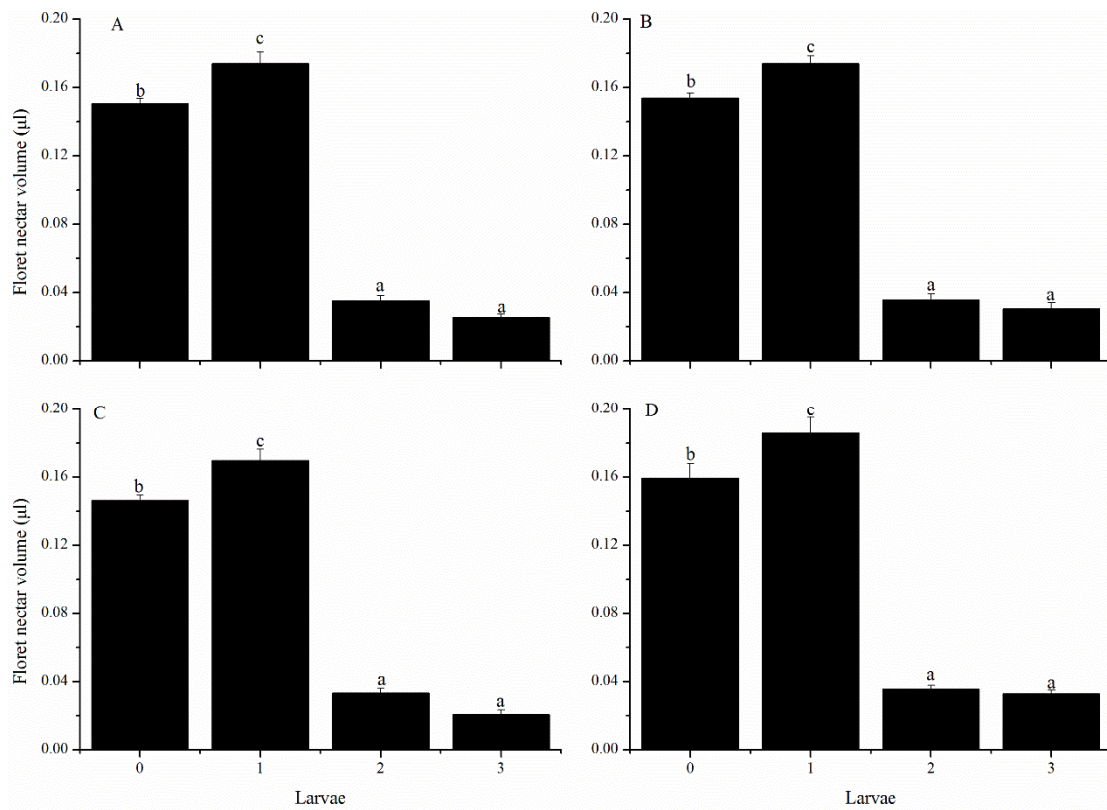
519 Wallis de Vries MF, van Swaay CAM, Plate CL (2012) Changes in nectar supply: a  
520 possible cause of widespread butterfly decline. *Curr Zool* 58: 384-391.

521 Westerbergh A and Westerbergh J (2001) Interactions between seed predators /  
522 pollinators and their host plants: a first step towards mutualism. *Oikos* 95: 324-  
523 334.

- 524 Whittall JB and Hodges SA (2007) Pollinator shifts drive increasingly long nectar spurs  
525 in columbine flowers. *Nature* 447: 706-709.
- 526 Xi XQ, Wu XW, Nylin S, and Sun SC (2016) Body size response to warming: time of  
527 the season matters in tephritid fly. *Oikos* 125: 386-394.
- 528 Yoshida S, Forno D, Cock J, Gomez KA (1976) *Laboratory manual for*  
529 *physiological studies of rice*. Los Baños, Laguna, Philippines. pp46.

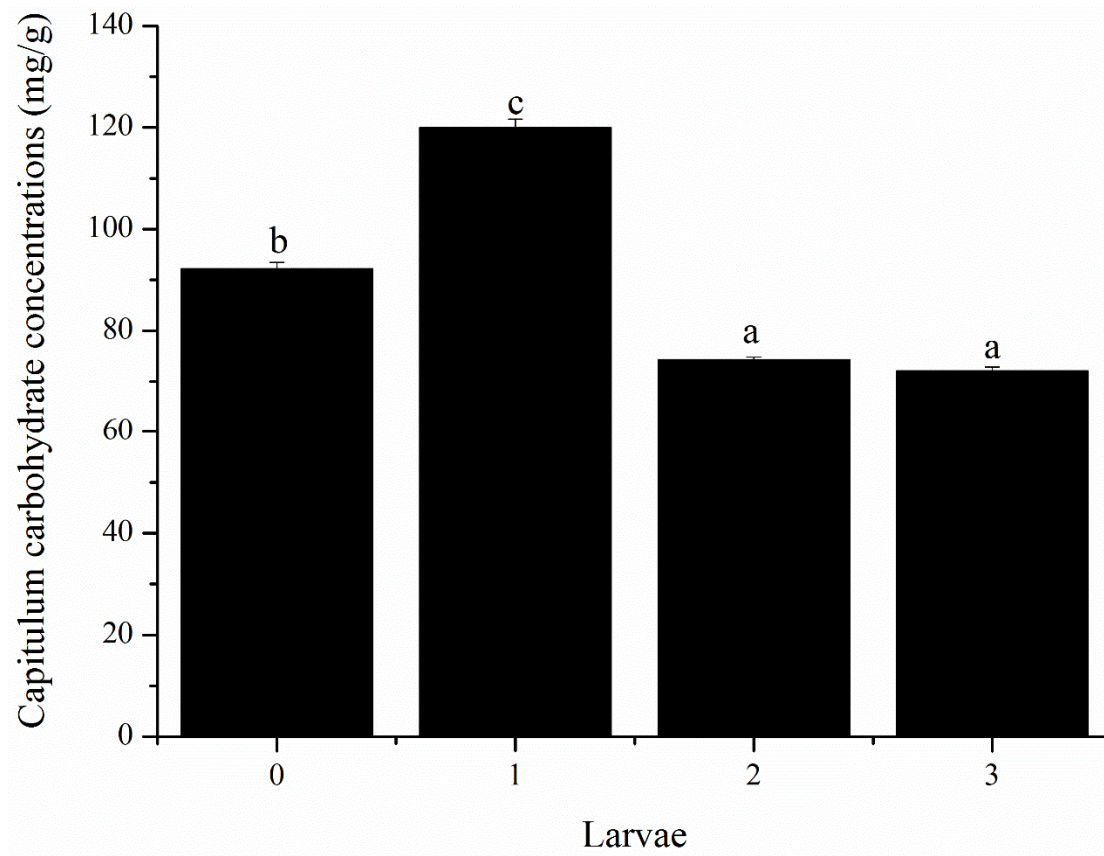
1 Figure 1. Variation in *Saussurea nigrescens* nectar volumes in relation to the numbers  
2 of tephritid larvae in their capitula (Means  $\pm$  SEs per floret). Figures 1A-1C plants  
3 growing at sites 1-3 with natural variation in larval numbers, Figure 1D  
4 experimentally introduced tephritid larvae (site 2 only). Different letters above  
5 columns indicate differences within sites at  $P < 0.05$  (one-way analysis of variance).

6  
7



8  
9

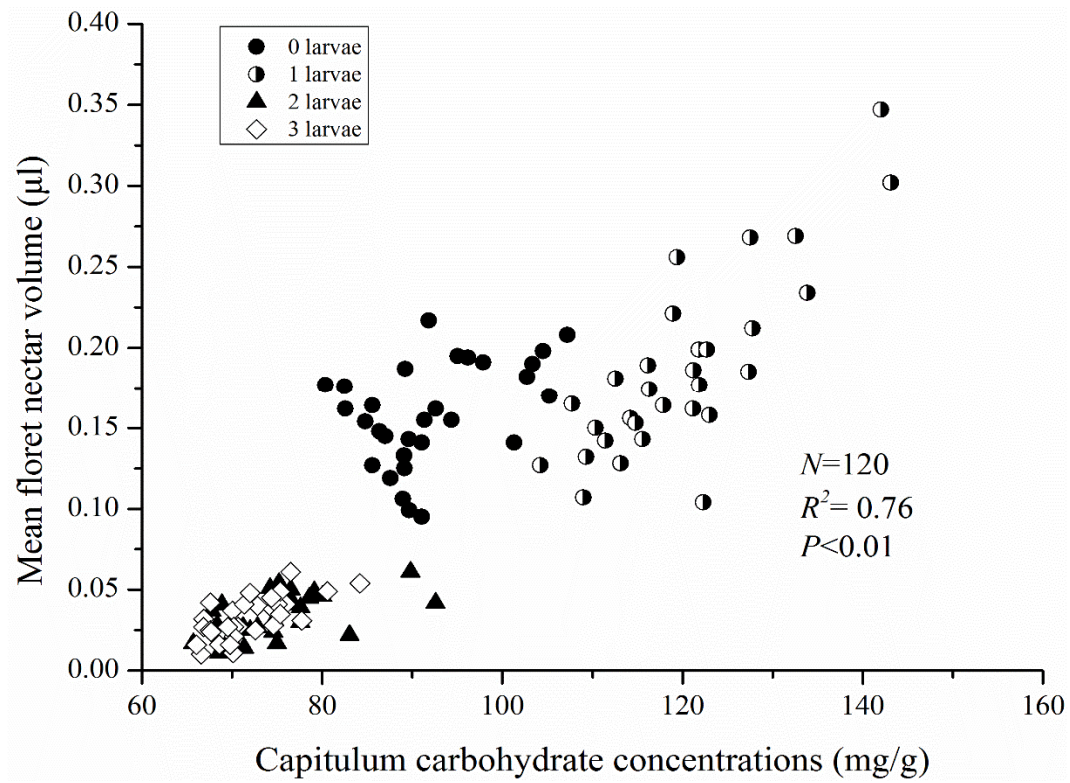
10 Figure 2. Capitulum carbohydrate concentrations of *S. nigrescens* that had contained  
11 varying numbers of tephritid larvae (Means  $\pm$  1 SE). Different letters above columns  
12 indicate differences at  $P < 0.05$  (one-way analysis of variance).



13  
14

15 Figure 3. The relationship between *Saussurea nigrescens* capitulum carbohydrate  
16 concentrations and mean floret nectar volume in capitula where different numbers of  
17 tephritid larvae had been introduced. N = 30 florets for each larval density.

18



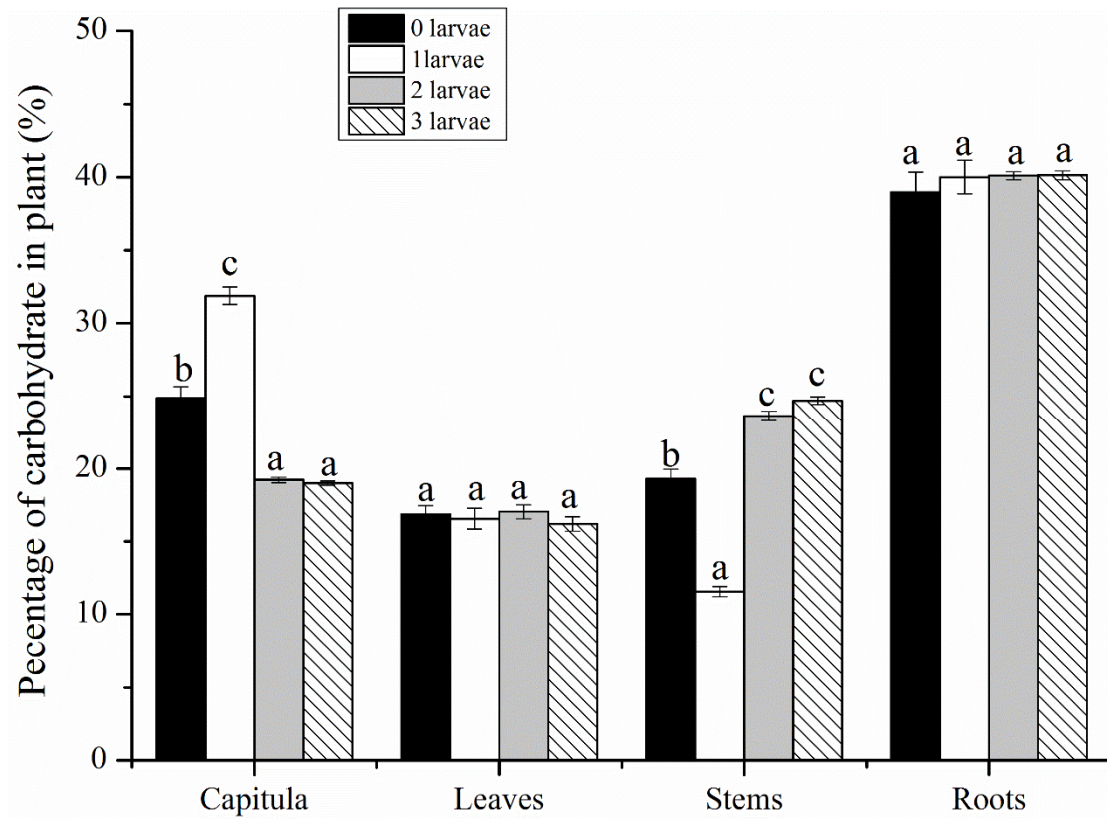
19

20

21

22

23 Figure 4. Percentage of total carbohydrates (soluble sugars and starch, mg/g) in  
 24 capitula, leaves, stems and roots of *S. nigrescens* (means  $\pm$  1 SE). Different letters  
 25 above columns indicate differences between parts of the plants at  $P < 0.05$  (one-way  
 26 analysis of variance).



27  
 28  
 29  
 30

Table 1. Summary of linear mixed effect models comparing the effects of pre-dispersal seed predators and capitulum size on floret nectar volume, nectar concentration, and capitulum carbohydrate concentrations. AIC = Akaike Information Criterion and BIC = Bayesian Information Criterion. The numbers after Floret nectar volume and Nectar concentration are AIC and BIC, respectively.

Variables /AIC/ BIC	Fixed effects			
	value	SE	t-value	p-value
<b>Field conditions</b>				
Floret nectar volume /-1539.99/-1515.10				
Intercept	0.159	0.009	18.119	<b>&lt;0.001</b>
Pre-dispersal seed predators (P)	-0.028	0.011	-2.441	<b>0.015</b>
Capitulum mass (C)	-0.050	0.054	0.929	0.353
P×C	-0.097	0.069	-1.403	0.161
Nectar concentration/3449.41/3474.30				
Intercept	39.977	2.000	19.984	<b>&lt;0.001</b>
Pre-dispersal seed predators(P)	-1.568	2.388	-0.657	0.512
Capitula mass (C)	10.411	12.789	0.814	0.416
P×C	8.517	14.612	0.583	0.560
<b>Experimental manipulations</b>				
Floret nectar volume /-338.37/-321.85				
Intercept	0.159	0.018	8.983	<b>&lt;0.001</b>
Pre-dispersal seed predators (P)	-0.046	0.010	-4.587	<b>&lt;0.001</b>
Capitula mass (C)	0.158	0.106	1.495	0.138
P×C	-0.046	0.061	-0.759	0.449
Nectar concentration/747.25/763.77				
Intercept	41.630	1.909	21.802	<b>&lt;0.001</b>
Pre-dispersal seed predators (P)	-0.055	1.083	-0.051	0.959
Capitulum mass (C)	-2.152	11.400	-0.189	0.851
P×C	-1.901	6.548	-0.290	0.772
Capitulum carbohydrate concentrations /1016.50/1033.02				
Intercept	187.545	46.583	4.026	<b>&lt;0.001</b>
Pre-dispersal seed predators (P)	-8.602	3.921	-2.194	<b>0.030</b>
Capitulum mass (C)	16.649	30.306	0.549	0.584
P×C	-1.391	2.544	-0.545	0.587

## Appendix list

### Appendix A

*S. nigrescens* nectar concentrations at sites 1 (A), 2 (B), and 3 (C) under natural condition and after experimental introductions (D).

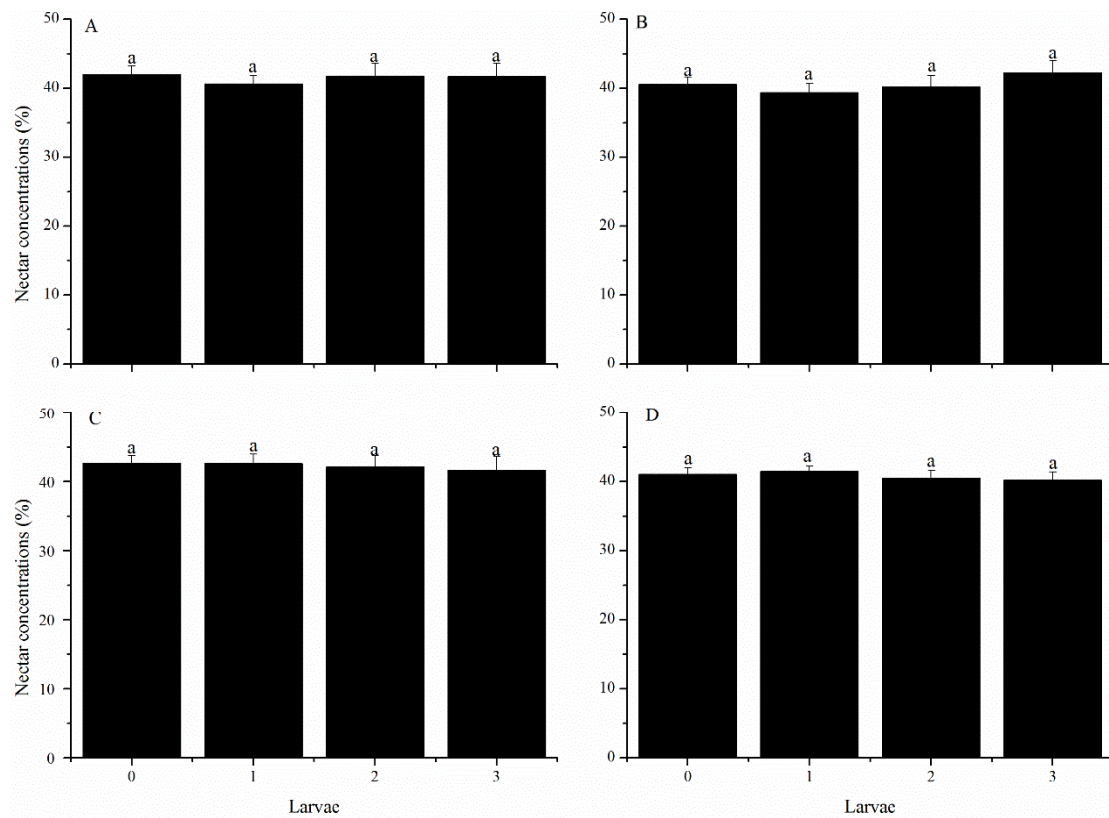
### Appendix B

*Apis mellifera* visitation rates per capitulum of *Saussurea nigrescens* in relation to tephritid larval numbers.



## Appendix A

*S. nigrescens* nectar concentrations with natural variation in numbers of tephritid larvae at sites 1-3 (A-C) and in the larval-addition experiment at site 2 (D). Similar letters above columns indicate no differences within sites at  $P < 0.05$  (one-way analysis of variance).



## Appendix B

*Apis mellifera* visits to capitula of *Saussurea nigrescens* that contained different numbers of tephritid larvae (means  $\pm$  1 SE). Capitulum visitation rates were compared by one-way ANOVAs followed by Tukey's tests. Similar letters above columns indicate no difference in visitation rates at  $P < 0.05$  (one-way analysis of variance).

