



Deposited via The University of Sheffield.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/105835/>

Version: Accepted Version

---

**Article:**

Fogh, I., Lin, K., Tiloca, C. et al. (2016) Association of a Locus in the CAMTA1 Gene With Survival in Patients With Sporadic Amyotrophic Lateral Sclerosis. *JAMA Neurology*, 73 (7). pp. 812-820. ISSN: 2168-6149

<https://doi.org/10.1001/jamaneurol.2016.1114>

---

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

## **A locus in the *CAMTA1* gene is associated with survival in patients with sporadic amyotrophic lateral sclerosis**

Isabella Fogh<sup>1</sup>, Kuang Lin<sup>1</sup>, Cinzia Tiloca<sup>2</sup>, James Rooney<sup>3</sup>, Cinzia Gellera<sup>4</sup>, Frank P. Diekstra<sup>5</sup>, Antonia Ratti<sup>2,6</sup>, Aleksey Shatunov<sup>1</sup>, Michael A. van Es<sup>5</sup>, Petroula Proitsi<sup>1</sup>, Ashley Jones<sup>1</sup>, William Sproviero<sup>1</sup>, Adriano Chiò<sup>7,8</sup>, Russell Lewis McLaughlin<sup>9</sup>, Gianni Sorarù<sup>10</sup>, Lucia Corrado<sup>11</sup>, Daniel Sthal<sup>12</sup>, Roberto Del Bo<sup>13</sup>, Cristina Cereda<sup>14</sup>, Barbara Castellotti<sup>4</sup>, the SLAGEN Consortium and Collaborators, Jonathan D. Glass<sup>15</sup>, Steven Newhouse<sup>16,17</sup>, Richard Dobson<sup>16,18</sup>, Bradley N. Smith<sup>1</sup>, Vincent Meininger<sup>19</sup>, Judith Melki<sup>20</sup>, Karen E. Morrison<sup>21</sup>, Pamela J. Shaw<sup>22</sup>, Nigel P. Leigh<sup>23</sup>, Peter M. Andersen<sup>24,25</sup>, Giacomo P. Comi<sup>13</sup>, Nicola Ticozzi<sup>2</sup>, Letizia Mazzini<sup>11,26</sup>, Sandra D'Alfonso<sup>11</sup>, Bryan J. Traynor<sup>27</sup>, Philip Van Damme<sup>28,29</sup>, Wim Robberecht<sup>28</sup>, Robert H. Brown<sup>30</sup>, John E. Landers<sup>30</sup>, Orla Hardiman<sup>9</sup>, Cathryn M. Lewis<sup>31,32</sup>, Leonard H. van den Berg<sup>5</sup>, Christopher E. Shaw<sup>1</sup>, Jan H. Veldink<sup>5</sup>, Vincenzo Silani<sup>2,6</sup>, Ammar Al-Chalabi<sup>1</sup> and John Powell<sup>1</sup>

1. Department of Basic and Clinical Neuroscience, Maurice Wohl Clinical Neuroscience Institute, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom
2. Department of Neurology and Laboratory of Neuroscience, IRCCS Istituto Auxologico Italiano, Milano, Italy
3. Academic Unit of Neurology, Trinity College Dublin, Trinity Biomedical Sciences Institute, Dublin, Republic of Ireland
4. Unit of Genetics of Neurodegenerative and Metabolic Diseases, Fondazione IRCCS Istituto Neurologico 'Carlo Besta', Milano, Italy
5. Department of Neurology and Neurosurgery, Brain Center Rudolf Magnus, University Medical Center Utrecht, The Netherlands
6. Department of Pathophysiology and Transplantation, "Dino Ferrari" Center, Università degli Studi di Milano, 20122 Milano, Italy
7. "Rita Levi Montalcini" Department of Neuroscience, ALS Centre, University of Torino, Turin, Italy
8. Azienda Ospedaliera Città della Salute e della Scienza, Torino, Italy
9. Population Genetics Laboratory, Smurfit Institute of Genetics, Trinity College Dublin, Dublin, Republic of Ireland
10. Department of Neurosciences, University of Padova, Padua, Italy
11. Department of Health Sciences, Interdisciplinary Research Center of Autoimmune Diseases, 'A. Avogadro' University, 28100 Novara, Italy
12. Department of Biostatistics, Institute of Psychiatry, Psychology and Neuroscience (IoPPN), King's College London, London, United Kingdom
13. Neurologic Unit, IRCCS Foundation Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy
14. Laboratory of Experimental Neurobiology, IRCCS 'C. Mondino' National Institute of Neurology Foundation, 27100 Pavia, Italy
15. Department of Neurology, Emory University, Atlanta, GA 30322, USA
16. King's College London, Institute of Psychiatry, Psychology & Neuroscience and NIHR Biomedical Research Centre for Mental Health, London, United Kingdom
17. King's College London, Institute of Psychiatry, Psychology & Neuroscience, Department of Biostatistics, London, United Kingdom
18. NIHR Biomedical Research Unit Dementia, London, United Kingdom
19. Département des Maladies du Système Nerveux, APHP, Réseau SLA Île de France, Hôpital Pitié-Salpêtrière, Paris, France
20. INSERM UMR-788 and University of Paris 11, Bicetre Hospital, Paris, Cedex 94275, France

21. School of Clinical and Experimental Medicine, College of Medicine and Dentistry, University of Birmingham, and Neurosciences Division, University Hospitals Birmingham NHS Foundation Trust, Birmingham B15 2TT, United Kingdom
22. Academic Neurology Unit, Department of Neuroscience, Faculty of Medicine, Dentistry and Health, University of Sheffield, Sheffield S10 2HQ, United Kingdom
23. Brighton and Sussex Medical School, Trafford Centre for Biomedical Research, University of Sussex, Falmer East Sussex BN1 9RY, United Kingdom
24. Institute of Clinical Molecular Biology, Kiel University, Kiel, Germany
25. Department of Pharmacology and Clinical Neuroscience, Umeå University, Umeå, Sweden.
26. ALS Center Department of Neurology, "Maggiore della Carità" University Hospital, 28100 Novara, Italy
27. Neuromuscular Diseases Research Group, Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, MD 20892, USA
28. Department of Neurology, University Hospital Leuven, Leuven Belgium
29. Divisions of Endocrinology and Genetics and Centre for Basic Translational Obesity Research, Boston Children's Hospital, Boston, Massachusetts, United States of America
30. Department of Neurology, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America
31. IoPPN Genomics & Biomarker Core, Translational Genetics Group, MRC Social, Genetic and Developmental Psychiatry Centre, King's College London, London, United Kingdom
32. Department of Medical and Molecular Genetics, King's College London, London, United Kingdom

**Corresponding author:** Isabella Fogh, Maurice Wohl Clinical Neuroscience Institute, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, United Kingdom. E-mail: [isabella.fogh@kcl.ac.uk](mailto:isabella.fogh@kcl.ac.uk)

### **Keywords**

Amotrophic lateral sclerosis, Genome wide association studies, survival, Cox proportional hazards regression analysis.

## Summary (300 words)

### Background

Amyotrophic lateral sclerosis (ALS) has a poor prognosis with a median survival of 3 years. However, a significant proportion of patients survive more than 10 years from symptom onset. Identification of gene variants influencing survival is crucial, as ALS is a rare disease and prevention is not feasible, leaving treatment that modifies survival the only realistic strategy. We performed a genome-wide association study (GWAS) to identify genetic modifiers of ALS survival.

### Methods

We analysed survival in a large international GWAS data collection of 4255 patients (73.4% deceased) with genotype data extended to 7174392 variants by imputation analysis. We employed Cox proportional hazards regression under an additive model with adjustment for age of onset, sex and the first four principal components of ancestry, followed by meta-analysis. In a subset of 3439 patients Cox proportional hazards regression model was additionally adjusted by site of onset. Survival distribution for the most associated variants was assessed by Kaplan-Meier analysis.

### Findings

We identified two novel loci significantly associated with ALS survival at 10q23 (rs139550538,  $p = 1.87 \times 10^{-9}$ ) and in the *CAMTA1* gene at 1p36 (rs2412208,  $p = 3.53 \times 10^{-8}$ ). At locus 10q23 the adjusted hazard ratio for patients with the rs139550538 AA or AT genotype was 1.61 (95%CI: 1.38-1.89), corresponding to a 7 month reduction in survival compared to TT carriers. For rs2412208-*CAMTA1* the adjusted hazard ratio for patients with the GG or GT genotype was 1.17 (95%CI 1.11-1.24), corresponding to a 4 month reduction in survival compared to TT carriers.

### Interpretation

We have identified two loci that influence survival in ALS. Identification of the underlying mechanisms may suggest new therapeutic targets for ALS treatment.

### Funding

This project was funded by the European Union FP7 and JPND programmes, the UK NIHR, the Motor Neurone Disease Association.

## Introduction

Amyotrophic lateral sclerosis (ALS) is a neurodegenerative disease of motor neurons in which relentlessly progressive weakness of voluntary muscles leads to death from respiratory failure on average within 3 years of symptom onset. ALS is a heterogeneous disease with a poorly understood aetiology. Phenotypic variability in ALS is remarkable, comprising heterogeneity in disease duration, age and site of onset, and type of motor neuron affected.<sup>1</sup> Several ALS genes have been identified. Of these, a massive hexanucleotide repeat expansion in the *C9orf72* gene is the most common mutation in both familial and sporadic patients.<sup>2,3</sup> Large genome wide association studies (GWAS) have identified a number of susceptibility genes including *UNC13A*<sup>4</sup>, *C9orf72*<sup>5</sup> and *SARM1*.<sup>6</sup>

Despite the poor prognosis of ALS, about 5% of patients may survive more than 10 years.<sup>7</sup> Long survivors are more likely to have primary lateral sclerosis, but all phenotypic patterns are represented. Younger age at onset correlates with longer survival, and other prognostic factors include disease progression rate at diagnosis, site of involvement at onset, certain phenotypic patterns (flail limb variants), cognitive impairment and respiratory involvement.<sup>8-12</sup>

Previous studies have reported association of survival with single nucleotide polymorphisms (SNPs) in the *KIFAP3* and *UNC13A* genes,<sup>13,14</sup> although the *KIFAP3* finding has not been replicated.<sup>15,16</sup>

Identification of gene variants influencing survival is crucial. ALS is a rare disease and prevention is not feasible, leaving treatment that modifies survival the only realistic strategy. An approach that leads to improved understanding of the biological basis of survival in ALS could lead to the development of a rational treatment. Therefore, to identify modifier genes that might influence ALS survival, we performed a GWAS using Cox proportional hazards regression model including age and site of onset as covariates followed by meta-analysis.

## Methods

### *Samples and data*

Genotypes were obtained from published GWAS of ALS sporadic patients from Italy, USA, UK, Ireland, Sweden, Belgium and France, including the Italian Consortium for the Genetics of ALS (SLAGEN) collected after ethically approved informed consent (webappendix p 2, Table 1). Participating patients fulfilled the El Escorial revised criteria for ALS<sup>17,18</sup> without a reported family history of ALS. Individuals included were of European ancestry by self-declaration. Clinical information was collected from medical notes, including date of last consultation, and survival data from death certificates, hospital or public records. Symptom onset was defined as the date of first weakness, speech or swallowing disturbances. Survival duration was defined as the difference between date of death/tracheostomy and date of symptom onset and, for those still alive, as the difference between censor date and symptom onset. The censor date was taken as date of last follow-up. Site of onset was defined as bulbar for those with first weakness affecting speech or swallowing, or spinal for those with limb or respiratory symptoms at onset.

### *Genotyping and quality control*

Genotyping was performed on Illumina DNA microarrays as previously described.<sup>6</sup> Standard GWAS quality control measures were used to exclude samples or SNP assays of poor quality (webappendix p 2, webappendix tables 2 and 3). Imputation was performed using 1000 Genomes Project Phase I version 3 (NCBI build 37, hg19 coordinates, August 2012) as the reference panel and IMPUTE2 software.<sup>19,20</sup> Imputed genotypes were filtered for uncertainty of inferred genotype and minor allele frequency (MAF).

### **Statistical analysis**

Multivariate Cox proportional hazards regression was modelled to estimate crude hazard ratios (HRs) and build by backward elimination (Wald test), estimation of hazard ratio, and 95% confidence interval (CI). Cox proportional hazards baseline model included age at onset (as continuous variable), gender and site at onset (bulbar versus spinal) as factor variables (webappendix table 6).

The proportional hazard assumption was tested by comparing the hazard curves stratified by sex, age and site at onset.

All tests were two-tailed and significance was assessed at  $p < 0.05$  and performed in SPSS (version 22, IBM Corporation, Chicago, IL, USA).

The Cox proportional hazards model was applied genome-wide to filtered imputed data in each population, with the following independent variables: SNP genotype under a log-additive model, four principal components of ancestry, sex and age of onset. To maximise power in the exploratory analysis, site at onset was omitted in the final model due to the smaller numbers of patients (81%) with this information.

The model was built by backward elimination using the *pacoph* program in the ProbABEL<sup>21</sup> toolset to estimate for each SNP the hazard ratio with 95% CIs, model and covariate p-values. Statistical significance was assessed at the genome-wide level ( $p = 5 \times 10^{-8}$ ).

Summary statistics for 7174392 overlapping SNPs were combined in meta-analysis using METAL<sup>22</sup> weighted by  $\beta$ -coefficients and the inverse of the corresponding standard errors; fixed-effects model was applied to adjust data from the seven independent studies. Genomic inflation was tested by Q-Q plots and factor lambda estimate ( $\lambda_{(gc)} = 1.05$ ) (webappendix p 2, fig.1).

The most associated variants were tested for heterogeneity of allele frequencies between studies by Cochran's Q test (Q) (webappendix p 3, Table 7). SNPs achieving genome-wide significance in the meta-analysis were tested by Kaplan-Meier analysis and a log rank test. Kaplan-Meier curves for additive and dominant models were compared by  $\chi^2$  likelihood ratio tests.

### Role of the funding source

The funders had no role in study design, data collection, analysis or interpretation, nor writing of the report. All authors had full access to all the data and the corresponding authors had final responsibility for deciding to submit for publication.

### Results

The international ALS cohort analysed in the present study included 4255 patients, 2591 male and 1664 female, of whom 3125 (73.4%) had died with median survival of 33 (interquartile range (IQR): 27) months. The mean age of onset including censored individuals was 59 (standard deviation (SD) $\pm$ 12) years (webappendix Table 4).

Site of onset data were available on a subset of 3439 (81%) patients (2066 male, 1372 female), 27% had bulbar onset, with a mean age of onset of 60.3 (SD $\pm$ 11.3), compared with spinal onset mean age 56.6 (SD $\pm$ 12.3) while median survival was 26.3 (IQR: 19.2) and 33.8 (IQR: 21.2-54) months in bulbar and spinal patients, respectively. Full details are reported in the webappendix Table 5.

There were 7174392 SNPs with genotypes passing quality control. There were two loci exceeding the genome-wide significance threshold, one on chromosome 10q23 and one on chromosome 1p36 (figure 1, table 1). At locus 10q23, the top ranked SNP was rs139550538, hazard ratio 1.61 (95% CI: 1.38, 1.89),  $p = 1.87 \times 10^{-9}$ . This is a moderately rare variant (MAF 0.03), intronic within the insulin-degrading enzyme gene, *IDE* (figure 2).

At the 1p36 locus there were four SNPs exceeding genome wide significance, with the top ranked SNP being rs2412208, hazard ratio 1.17 (95% CI: 1.11, 1.24),  $p = 3.53 \times 10^{-8}$  followed by 87 SNPs in strong linkage disequilibrium with rs2412208. All these SNPs fell within a 90 Kb region encompassing intron 3-4 of the calmodulin binding transcription activator 1 gene, *CAMTA1* (figure 2 and table 1). Cox proportional hazard regression analyses conditioning upon the most associated SNPs in both loci showed no evidence of residual association.

Because rs139550538 is rare, Kaplan-Meier analysis was performed under a dominant model (226 patients (5.6%) carried at least one A allele). AA or AT genotype was associated with ALS survival (log rank  $p = 1.3 \times 10^{-7}$ ) and a median survival of 30.7 months compared with 36.7 months for the TT homozygotes (figure 3, table 2).

Kaplan-Meier analysis of SNP rs2412208 under an additive model showed that carrying a G allele (45% of patients) was significantly associated with a decreased survival (log-rank  $p = 5.6 \times 10^{-7}$ ) with median survivals of 36.0 months (GG), 36.8 months (GT) and 40.8 months (TT) (Figure 4, Table2). The hazard ratio estimates were consistent across the seven datasets

analysed (figure 5). Under a dominant model the results were similar (Figure 4) and a  $\chi^2$  likelihood ratio test comparing the two models was not significant ( $p = 0.12$ ), showing either could be valid.

We tested whether observed effect sizes ( $\beta$ ) of the most associated SNPs from the combined Cox proportional hazard analysis were homogeneous across cases. There was some evidence for heterogeneity across the different datasets ( $rs2412208 I^2 = 59.1\%$ ,  $P = 0.02$ ) (webappendix table 7).

In the subset of 3439 ALS patients with clinical data information including site of onset information, Cox proportional hazards regression was modelled with this variable as additional covariate. The top rank SNP was  $rs2412208$  at  $1p36$  with the combined hazard ratio of  $1.19$  (95% CI:  $1.27-1.12$ ;  $p=5.11 \times 10^{-8}$ ) (webappendix Fig.2, Table 8) confirming association of *CAMTA1* locus with ALS survival identified by the larger sample size when this covariate was excluded from the model. The SNP  $rs139550538$  in *IDE* gene was less significant (HR  $1.51$ ; 95% CI:  $1.27-1.78$ ;  $p=2.24 \times 10^{-6}$ ) possible because the lower frequency of this SNP in a reduced sample. Additionally, in linear regression analysis testing bulbar versus spinal phenotypes, this variant was not significantly associated ( $p = 0.5$ ) (data not shown) indicating that the inclusion of site at onset as covariate in the Cox proportional hazard model was sufficient to decrease the strength of locus  $10q23$  association with survival.

Kaplan-Meier distribution of  $rs2412208$  genotypes indicated that risk allele G was associated (log-rank  $p=5 \times 10^{-6}$ ) with a shorter survival of 3.5 months corresponding to 19% increased rate of mortality compared to the TT homozygotes (webappendix Fig.3).

We examined previously reported candidate genes for ALS survival. SNP  $rs1541160$  in the *KIFAP3* gene was not significantly associated with survival in this study (HR  $1.04$ ; 95% CI:  $0.98, 1.1$ ;  $p = 0.423$ ) (webappendix Fig 4), confirming previous findings.<sup>15, 16</sup> SNP  $rs12608932$  in the *UNC13A* gene showed suggestive association with HR  $1.17$ ; 95% CI:  $1.1, 1.24$ ;  $p = 0.003$  but coverage for this SNP was limited to a reduced subset of patients ( $n = 3574$ ) (webappendix Fig. 5) and further studies on a larger scale are needed to validate the genetic effect of *UNC13A* as survival modifier. Of 105 SNPs tested in the *DAO* gene<sup>23</sup>, the top ranked was  $rs4623951$ , with a hazard ratio of  $1.07$  (95%CI:  $1.02-1.13$ ,  $p = 0.0053$ ).

## Discussion

We have identified two loci associated with survival in patients with ALS at genome wide significance in a large meta-analysis using Cox proportional hazards regression analysis. The discovery of gene variants within *IDE* and *CAMTA1* genes as survival modifiers in ALS is important both because of improved understanding of the disease process, and because the genes and associated pathways might become a target for therapy development. Furthermore, if gene variants have a large effect on survival, it is important to account for this in the design and analysis of clinical trials.

The effect size of the variants found is comparable to that of Riluzole, a drug shown to improve survival in ALS, for which the hazard ratio for those not taking Riluzole compared with those taking the drug is  $1.14$ . A weakness of this study is that the extent of Riluzole use was not available to include in the analysis. Generally, rates of prescription are higher in countries in which access to healthcare is free or reimbursed than in those where private insurance is required, and if such differences correlate with allele frequency differences, a spurious association might arise. We have mitigated against this both by accounting for differences in allele frequency by ancestry using principal components, and by accounting for differences in Riluzole prescription rates by performing a meta-analysis stratified by country.

The most associated polymorphism at the  $10q23$  locus was a low frequency variant within the *IDE* gene, a zinc metallopeptidase that degrades intracellular insulin and other peptides such as beta-amyloid. Tagged proxies for this polymorphism were in weak ( $r^2 < 0.4$ ) linkage disequilibrium and located in a neighbouring gene, *KIF11*, a motor kinesin-like protein involved in the spindle function during cell mitosis (figure 2A). The biological basis of this association is therefore unclear.

The most associated 87 variants in the *CAMTA1* gene ( $p \leq 10^{-4}$ ) map to a small 90 kb region within intron 3-4 (figure 2B) encompassing the CG-1 DNA-binding domain. The CG-1 motif is a functional domain with a nuclear localisation signal and transcriptional regulation properties that extends from exon 3 to exon 7 (6825092 to 7640553 base pair; GRCh37/hg19 Assembly) within *CAMTA1*. Intragenic *CAMTA1* micro-rearrangements disrupting a CG-1 DNA binding domain have been reported to co-segregate with non-progressive congenital cerebellar ataxia (NPCA) and gait instability in several unrelated families<sup>24, 25</sup> Common

variants within *CAMTA1* have been reported to be associated with variation in human episodic memory.<sup>26</sup> Mutant *CAMTA1* knock-out mice, disrupted in the CG-1 domain, show severe ataxia and neuronal atrophy approximating the phenotype of haploinsufficiency observed in NPCA patients.<sup>27</sup> Furthermore, the identification of the consensus sequences of the DNA-binding site of CG-1 domain combined with expression analyses in *CAMTA1* knock-out mice have shown more than 80 neural related genes regulated by *CAMTA1*.<sup>27</sup> The finding of a gene involved in cerebellar disease in ALS is not surprising, given that *ATXN2* trinucleotide repeat expansion causes spinocerebellar ataxia or ALS,<sup>28</sup> the finding of *C9orf72* pathology in the cerebellum of ALS patients,<sup>29</sup> and the discovery of abnormal eye gaze in ALS patients.<sup>30, 31</sup> Thus there is increasing evidence of a relationship between ALS and cerebellar degeneration that is currently under-recognised, in the same way as the relationship between ALS and frontotemporal dementia remained undetected until recently.

Strength of this study is the large sample size. Use of the Cox proportional hazard model allowed us to include the nearly 27% of patients still alive (n = 1131) and means the analysis is not biased by restricting the analysis to a linear regression in those who have died. Weaknesses of our study include the difficulty of imputing low frequency variants; several significant or suggestive findings were of rare alleles. Better imputation panels and disease specific imputation panels will improve imputation of rare genetic variation.

In conclusion, we have identified genetic variants statistically significantly associated with survival. The promise of this research is not only to improve our understanding of the biology of the disease and suggest biological targets for pharmaceutical intervention to extend the survival time, but also to use genetic risk scores as an adjunct to clinical trials to account for the genetic contribution to survival.

## References

1. Swinnen B, Robberecht W. The phenotypic variability of amyotrophic lateral sclerosis. *Nature reviews Neurology* 2014; **10**(11): 661-70.
2. DeJesus-Hernandez M, Mackenzie IR, Boeve BF, et al. Expanded GGGGCC hexanucleotide repeat in noncoding region of C9ORF72 causes chromosome 9p-linked FTD and ALS. *Neuron* 2011; **72**(2): 245-56.
3. Renton AE, Majounie E, Waite A, et al. A hexanucleotide repeat expansion in C9ORF72 is the cause of chromosome 9p21-linked ALS-FTD. *Neuron* 2011; **72**(2): 257-68.
4. van Es MA, Veldink JH, Saris CG, et al. Genome-wide association study identifies 19p13.3 (UNC13A) and 9p21.2 as susceptibility loci for sporadic amyotrophic lateral sclerosis. *Nature genetics* 2009; **41**(10): 1083-7.
5. Shatunov A, Mok K, Newhouse S, et al. Chromosome 9p21 in sporadic amyotrophic lateral sclerosis in the UK and seven other countries: a genome-wide association study. *The Lancet Neurology* 2010; **9**(10): 986-94.
6. Fogh I, Ratti A, Gellera C, et al. A genome-wide association meta-analysis identifies a novel locus at 17q11.2 associated with sporadic amyotrophic lateral sclerosis. *Human molecular genetics* 2014; **23**(8): 2220-31.
7. Pupillo E, Messina P, Logroscino G, Beghi E. Long-term survival in amyotrophic lateral sclerosis: a population-based study. *Annals of neurology* 2014; **75**(2): 287-97.
8. Al-Chalabi A, Lewis CM. Modelling the effects of penetrance and family size on rates of sporadic and familial disease. *Human heredity* 2011; **71**(4): 281-8.
9. Turner MR, Parton MJ, Shaw CE, Leigh PN, Al-Chalabi A. Prolonged survival in motor neuron disease: a descriptive study of the King's database 1990-2002. *Journal of neurology, neurosurgery, and psychiatry* 2003; **74**(7): 995-7.
10. Chio A, Mora G, Leone M, et al. Early symptom progression rate is related to ALS outcome: a prospective population-based study. *Neurology* 2002; **59**(1): 99-103.
11. Wijesekera L, Mathers S, Talman P, et al. Natural history and clinical features of the flail arm and flail leg ALS variants. *Neurology* 2009; **72**(12): 1087-94.
12. Byrne S, Elamin M, Bede P, et al. Cognitive and clinical characteristics of patients with amyotrophic lateral sclerosis carrying a C9orf72 repeat expansion: a population-based cohort study. *The Lancet Neurology* 2012; **11**(3): 232-40.
13. Landers JE, Melki J, Meininger V, et al. Reduced expression of the Kinesin-Associated Protein 3 (KIFAP3) gene increases survival in sporadic amyotrophic lateral sclerosis. *Proceedings of the National Academy of Sciences of the United States of America* 2009; **106**(22): 9004-9.
14. Diekstra FP, van Vught PW, van Rheenen W, et al. UNC13A is a modifier of survival in amyotrophic lateral sclerosis. *Neurobiology of aging* 2012; **33**(3): 630.e3-8.
15. van Doormaal PT, Ticozzi N, Gellera C, et al. Analysis of the KIFAP3 gene in amyotrophic lateral sclerosis: a multicenter survival study. *Neurobiology of aging* 2014; **35**(10): 2420.e13-4.
16. Traynor BJ, Nalls M, Lai S-L, et al. Kinesin-associated protein 3 (KIFAP3) has no effect on survival in a population-based cohort of ALS patients. *Proceedings of the National Academy of Sciences* 2010; **107**(27): 12335-8.
17. Brooks BR. El Escorial World Federation of Neurology criteria for the diagnosis of amyotrophic lateral sclerosis. Subcommittee on Motor Neuron Diseases/Amyotrophic Lateral Sclerosis of the World Federation of Neurology Research Group

on Neuromuscular Diseases and the El Escorial "Clinical limits of amyotrophic lateral sclerosis" workshop contributors. *Journal of the neurological sciences* 1994; **124 Suppl**: 96-107.

18. Brooks BR, Miller RG, Swash M, Munsat TL. El Escorial revisited: revised criteria for the diagnosis of amyotrophic lateral sclerosis. *Amyotrophic lateral sclerosis and other motor neuron disorders : official publication of the World Federation of Neurology, Research Group on Motor Neuron Diseases* 2000; **1**(5): 293-9.

19. Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS genetics* 2009; **5**(6): e1000529.

20. Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate genotype imputation in genome-wide association studies through pre-phasing. *Nature genetics* 2012; **44**(8): 955-9.

21. Aulchenko YS, Struchalin MV, van Duijn CM. ProbABEL package for genome-wide association analysis of imputed data. *BMC bioinformatics* 2010; **11**(1): 134.

22. Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* 2010; **26**(17): 2190-1.

23. Cirulli ET, Lasseigne BN, Petrovski S, et al. Exome sequencing in amyotrophic lateral sclerosis identifies risk genes and pathways. *Science* 2015; **347**(6229): 1436-41.

24. Shinawi M, Coorg R, Shimony J, Grange D, Al-Kateb H. Intragenic CAMTA1 deletions are associated with a spectrum of neurobehavioral phenotypes. *Clinical genetics* 2015; **87**(5): 478-82.

25. Thevenon J, Lopez E, Keren B, et al. Intragenic CAMTA1 rearrangements cause non-progressive congenital ataxia with or without intellectual disability. *Journal of medical genetics* 2012; **49**(6): 400-8.

26. Huentelman MJ, Papassotiropoulos A, Craig DW, et al. Calmodulin-binding transcription activator 1 (CAMTA1) alleles predispose human episodic memory performance. *Human molecular genetics* 2007; **16**(12): 1469-77.

27. Long C, Grueter CE, Song K, et al. Ataxia and Purkinje cell degeneration in mice lacking the CAMTA1 transcription factor. *Proceedings of the National Academy of Sciences* 2014; **111**(31): 11521-6.

28. Elden AC, Kim H-J, Hart MP, et al. Ataxin-2 intermediate-length polyglutamine expansions are associated with increased risk for ALS. *Nature* 2010; **466**(7310): 1069-75.

29. Al-Sarraj S, King A, Troakes C, et al. p62 positive, TDP-43 negative, neuronal cytoplasmic and intranuclear inclusions in the cerebellum and hippocampus define the pathology of C9orf72-linked FTL and MND/ALS. *Acta neuropathologica* 2011; **122**(6): 691-702.

30. Donaghy C, Thurtell MJ, Piro EP, Gibson JM, Leigh RJ. Eye movements in amyotrophic lateral sclerosis and its mimics: a review with illustrative cases. *Journal of Neurology, Neurosurgery & Psychiatry* 2011; **82**(1): 110-6.

31. Proudfoot M, Menke RA, Sharma R, et al. Eye-tracking in amyotrophic lateral sclerosis: A longitudinal study of saccadic and cognitive tasks. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration* 2015; (0): 1-11.