

This is a repository copy of *Interpreting BOLD: towards a dialogue between cognitive and cellular neuroscience*.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/102498/

Version: Accepted Version

### Article:

Hall, C.N., Howarth, C., Kurth-Nelson, Z. et al. (1 more author) (2016) Interpreting BOLD: towards a dialogue between cognitive and cellular neuroscience. Philosophical Transactions B: Biological Sciences , 371 (1705). 20150348. ISSN 0962-8436

https://doi.org/10.1098/rstb.2015.0348

### Reuse

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

### Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Interpreting BOLD: towards a dialogue between cognitive and cellular neuroscience

### Catherine N. Hall<sup>1\*</sup>, Clare Howarth<sup>2\*</sup>, Zebulun Kurth-Nelson<sup>3\*</sup>, Anusha Mishra<sup>4\*</sup>

<sup>1</sup> School of Psychology, University of Sussex, Brighton, UK

<sup>2</sup>Department of Psychology, University of Sheffield, Sheffield, UK

<sup>3</sup>Max Planck UCL Centre for Computational Psychiatry and Ageing Research, London, UK

<sup>4</sup>Department of Neuroscience, Physiology and Pharmacology, London, UK

<sup>\*</sup>All authors contributed equally to this work

Corresponding Authors: Catherine N. Hall (catherine.hall@sussex.ac.uk) and Clare Howarth (c.howarth@sheffield.ac.uk)

Author contributions: CNH, CH, ZK and AM conceived of the idea, coordinated the meeting, researched the literature and drafted the manuscript. All authors gave final approval for publication.

We declare we have no competing interests.

Acknowledgements: We thank David Attwell, Katie Boyd, Charlotte Rae and Cassandra Gould for comments on the paper. Clare Howarth holds a Sir Henry Dale Fellowship jointly funded by the Wellcome Trust and the Royal Society (grant number:105586/Z/14/Z). Catherine Hall receives research funding from the Royal Society, the Physiological Society, the University of Sussex MRC Discovery Award and an Academy of Medical Sciences/Wellcome Trust Springboard Award.

Cognitive neuroscience depends on the use of blood oxygenation level-dependent (BOLD) functional magnetic resonance imaging (fMRI) to probe brain function. Although commonly used as a surrogate measure of neuronal activity, BOLD signals actually reflect changes in brain blood oxygenation. Understanding the mechanisms linking neuronal activity to vascular perfusion is, therefore, critical in interpreting BOLD. Advances in cellular neuroscience demonstrating differences in this neurovascular relationship in different brain regions, conditions or pathologies are often not accounted for when interpreting BOLD. Meanwhile, within cognitive neuroscience, increasing use of high magnetic field strengths and the development of model-based tasks and analyses have broadened the capability of BOLD signals to inform us about the underlying neuronal activity, but these methods are less well understood by cellular neuroscientists. In 2016, a Royal Society Theo Murphy Meeting brought scientists from the two communities together to discuss these issues. Here we consolidate the main conclusions arising from that meeting. We discuss areas of consensus about what BOLD fMRI can tell us about underlying neuronal activity, and how advanced modelling techniques have improved our ability to use and interpret BOLD. We also highlight areas of controversy in understanding BOLD and suggest research directions required to resolve these issues.

### 1. What do we know about BOLD?

A. An increase in the positive BOLD signal in adults generally represents a net increase in *neuronal activity*. This relationship between the BOLD signal and neuronal activity is extensively discussed in previous reviews [1, 2] and in this issue (Lecrux and Hamel, Uhlirova et al., and Turner in this issue). Box 1 provides an introduction to the origin of the BOLD signal.

**B.** Neurovascular coupling links neuronal activity to increases in blood vessel diameter. Release of neurotransmitters such as glutamate induce dilation of cerebral blood vessels by triggering the release of vasoactive signalling molecules from both neurons and glia (LeCrux and Hamel, and Uhlirova et al. in this issue; [3]). Much is now known about these signalling pathways, but their variability across brain areas and circuits is likely to lead to differences in neurovascular coupling properties and therefore also to variations in the relationship between BOLD and neuronal activity across different brain regions and experimental conditions (discussed below).

**C. BOLD** is a better indicator of processing within an area than the output of the area. Where synaptic activity and action potentials do not correlate, BOLD signals better reflect synaptic activity. This concept has been demonstrated elegantly by studies in both anaesthetised and awake primate cortex [4, 5] and anaesthetised rat cerebellum [6, 7] and is highlighted in the articles by Lohrenz et al., and Freeman and Li in this issue. Lohrenz et al. report that striatal dopamine release, which reflects prediction error, can become dissociated from BOLD signals under certain conditions. Presumably such dissociations occur when competing inputs (e.g the glutamatergic input from prefrontal cortex), rather than dopamine release alone, drive striatal synaptic activity and energy use. Energy budgets of both the cerebral cortex and cerebellum imply that synaptic activity consumes more ATP and oxygen than action potentials [8, 9], suggesting an evolutionary drive for vascular responses to be more closely correlated with local synaptic activity than with spiking output. In this issue, Freeman and Li further illustrate this correlation using binocular interaction in the visual cortex *in vivo*.  $O_2$  usage is strongly correlated with both synaptic input and spiking output during excitatory binocular facilitation, whereas during inhibitory binocular suppression,  $O_2$  levels only reflect synaptic input and are dissociated from the spiking output. This is probably because the output no longer correlates with the degree of synaptic activation, and therefore with net  $O_2$  use.

**D. BOLD** can be used to glean robust information about functional localisation within the brain. This is true, in particular, when spatially localised groups of neurons are concurrently active (e.g. sensory cortical maps; [10] and Knutsen et al. in this issue), and the testing conditions can be well-controlled. Increases in magnetic strength further allow increases in the spatial resolution of these maps (discussed below and in papers by Turner and Ugurbil in this issue).

*E. Early BOLD signals are more reliable reporters of neuronal activity than are late BOLD signals.* Vascular responses propagate outward from their site of initiation [11, 12], which means that late BOLD responses are less spatially localised to the initial site of neuronal activity than the first few seconds of the positive BOLD response (discussed below, and by Uhlirova et al. in this issue and in [13]).

#### BOX 1: What is the BOLD signal?

The brain has only limited storage of the energy substrates oxygen and glucose. Although the extra energy use associated with cognitive tasks utilized for BOLD studies is typically <10% [14], when there is an increase in energy demand by active neurons, a local increase in blood flow is required to supply the extra glucose and oxygen required. This is achieved through neurovascular coupling. Work over the past two decades has attempted to define the signaling mechanisms that give rise to neurovascular coupling (see main text, Lecrux and Hamel in this issue and [3]), which is exploited by the BOLD fMRI technique to visualise neuronal activation. BOLD contrast is dependent on the level of deoxyhaemoglobin in the blood [15]. The iron in deoxyhaemoglobin is paramagnetic. This causes inhomogeneities in the magnetic field, which dephases "spins" and thus decreases the MR signal. When neural activity increases, oxygen extraction from the blood increases, resulting in an increase in the amount of deoxyhaemoglobin in the blood and a faster dephasing. This effect alone would decrease the MR signal. However, within a few seconds, neurovascular signals increase blood flow and volume, bringing in more oxygenated blood and washing out deoxyhaemoglobin. Thus, there is a net decrease in deoxyhaemoglobin, resulting in a more homogenous magnetic field in which transverse relaxation occurs more slowly and the BOLD fMRI signal increases. Hence, BOLD signals reflect the net increase in blood oxygenation following neural activity, and represent the sum of the effects of oxygen consumption (which decreases BOLD) and blood flow increase (which increases BOLD). The resulting functional imaging signal is assumed to reflect neuronal activity. As these two effects of neural activity are in opposite directions, however, alterations in signalling between the brain and the vasculature can potentially seriously confound BOLD interpretation. For example, if neural activity fails to increase blood flow, BOLD could decrease instead of increase. Indeed, during development, there is an extended period when the effect of the increase in energy expenditure outweighs the effect of the increase in blood flow, and the BOLD signal is negative [16]. Such potential confounds of the BOLD signal are discussed further in the main text and in Lake et al. in this issue.

## 2. Cellular mechanisms underlying BOLD: how our incomplete understanding limits our ability to interpret BOLD fMRI.

That increased neuronal activity leads to increased blood flow has been known for over 100 years [16], but the relative contribution of different parenchymal and vascular cells, and of different synapses, to neurovascular coupling remains an area of active research (see [3] and [17] for recent summaries).

Α. Synaptic activity is linked to cerebral blood flow (CBF) via glutamate release. As the majority of neurons within the cortex are excitatory [18] and glutamate has been known for a long time to evoke vascular dilation [19], neurovascular coupling research has primarily focused on glutamate-evoked blood flow changes. This glutamate-mediated neurovascular coupling neatly links the most energetically expensive component of synaptic transmission, ion flux through post-synaptic glutamate receptors [8, 20, 21], with increased energy supply. Glutamate evokes vascular responses by stimulating neurons and astrocytes to produce vasodilatory second messengers and by raising the extracellular potassium concentration, as has been comprehensively reviewed ([3], Uhlirova et al. in this issue). In summary, glutamate-evoked calcium signals in excitatory and inhibitory neurons can dilate blood vessels via production of nitric oxide [22], prostaglandins [23] and vasoactive peptides [24]. Neuronal activity has also been shown to stimulate astrocytes to release  $K^{+}$  [25] and vasodilatory arachidonic acid metabolites such as prostaglandin E<sub>2</sub> and epoxyeicosatrienoic acids [25-30] to dilate vessels (reviewed in [3]). However, the extent to which astrocytes contribute to functional hyperemia in vivo remains open to debate [31-34]. Rather than triggering a rapid vascular response, synaptic activity-evoked astrocyte calcium increases have been proposed to mediate sustained vascular responses to prolonged stimulation [32, 35, 36]. Furthermore, astrocytes also appear to play a role in maintaining the resting tone of the vasculature [37-39], which could affect the size of the blood flow response following neuronal activity [40].

*B.* Vascular mediators of neurovascular coupling - smooth muscle cells and pericytes. Contractile cells on blood vessel walls respond to neuron or astrocyte-derived second messengers by contracting or relaxing, thus constricting or dilating vessels and altering cerebral blood flow. Classically blood flow was thought to be regulated solely by rings of smooth muscle on penetrating arterioles, but recent work has suggested that capillary blood flow can also be regulated by virtue of the contraction and relaxation of pericytes [41, 42]. While these findings are somewhat disputed [43, 44], this is primarily due to differences in the definitions of capillaries, arterioles, smooth muscle cells and pericytes (see [45] and Uhlirova et al. in this issue). In fact, the studies broadly agree that cerebral blood vessels from large to small calibre (down to 4  $\mu$ m) and even up to 4 branches from

diving arterioles can be regulated by neuronal activity [42, 44] due to the relaxation of vascular mural cells which express contractile proteins, including those having a "bump on a log" morphology and processes extending along and around the blood vessels [44, 46]. Current evidence suggests that pericytes on capillary regions at branching orders more than 4 from penetrating arterioles are less contractile and therefore less likely to actively contribute to neurovascular coupling ([44]; Uhlirova et al. in this issue).

Capillary-level blood flow regulation explains how individual cortical columns can be mapped from the haemodynamic response (e.g. whisker barrels [47]), despite the territory fed by individual arterioles not mapping onto these columns [48]. Regulation of blood flow at the capillary level therefore likely sets the spatial resolution of the BOLD signal [49], at least at early time points after stimulation (see below).

С. Upstream propagation of vasodilation. It is now well-established that in the brain [12] vascular responses propagate outward from their site of initiation, similar to what has been observed in the periphery [11]. The cells propagating this signal may vary, depending on the strength of stimulation, with evidence supporting signal propagation along both the vascular endothelium [50] and through the astrocytic syncytium [51]. This means that, in the case of dilation, the positive BOLD signal spreads over an increasingly larger volume of tissue over time and therefore provides progressively less information as to the initiation site and the neurons that were originally activated. This has been demonstrated using intrinsic optical imaging (IOS) of haemodynamic changes in rodent barrel cortex [47], which correlate with the positive BOLD response (Knutsen et al. in this issue), and using in vivo 2 photon imaging of individual blood vessels in cat visual cortex [52]. In the first study, IOS changes could discriminate between two independently stimulated whiskers 1.5s after stimulation onset, but not after 4s, suggesting a spread of dilation to neighbouring regions over time. In the second study [52], individual arterioles dilated not only to stimuli that activated the cortical column surrounding the arteriole, but also to stimuli that activated adjacent cortical columns despite a lack of neuronal activity in the area immediately surrounding the blood vessel i.e. the vascular response occurred over a larger tissue volume than the neuronal response. Thus, uncoupling between neuronal and vascular responses exists at the level of cortical columns. Furthermore, pial vessels responded to a wider range of stimuli than diving vessels, presumably due to summation of dilations from several diving vessels fed by the same surface artery [52]. Propagation of vascular dilation away from the site of neuronal activity therefore gradually reduces the spatial resolution of the BOLD signal, which is greatest in the capillary bed shortly after stimulus onset.

D. Central neuromodulatory pathways. The cerebral vasculature is innervated by nerve terminals from subcortical regions that can release vasoactive neurotransmitters such as acetylcholine (from the basal ganglia), serotonin, dopamine (from the ventral tegmental area) and noradrenaline (from the locus coeruleus) [53-56]. Changes in emotional and arousal state can alter the release of these neurotransmitters, resulting in altered vascular tone independent of a change in synaptic activity [57]. Furthermore, release of neurotransmitters from monoaminergic and cholinergic subcortical afferents can evoke both a neural and CBF response concurrently. For instance, release of acetylcholine from afferents originating in the basal forebrain [58, 59] alters cortical neuronal activity, but also concurrently evokes increases in CBF [60]. The vascular response appears to be partly mediated by local somatostatin- or neuropeptide Y-positive or GABAergic interneurons [61] and/or astrocytes [59]. Similarly, vasoconstrictions evoked by noradrenaline released by locus coeruleus neurons are thought to be important in optimizing neurovascular coupling [62], but direct effects of this pathway on both excitatory and inhibitory cortical neurons resulting in increases in cortical CBF have also been demonstrated [63]. It is not yet established whether the subcorticallyevoked changes in cortical activity and blood flow always occur in the same direction - does neurotransmitter release from these afferents always increase both neuronal activity and blood flow, or do the neuronal and vascular responses sometimes uncouple? Even when they respond in the same direction, is the relationship between neuronal activity and the vascular response causal or correlative? If it is purely correlative, e.g. due to parallel signalling from the subcortical region to cortical neurons and to the vascular cells, then the relationship may fall apart in pathological conditions or during different arousal states, thus complicating our interpretation of BOLD.

**E.** How does this limit our interpretation of BOLD? As so many different cell types and second messenger pathways are involved in neurovascular coupling, it should come as no surprise that the properties of neurovascular coupling vary across brain areas, developmental stages and experimental conditions. Direct evidence for regional differences in the relationship between neuronal activity and BOLD has been demonstrated in rodents by simultaneously recording electrical and vascular activity. For example, increased neuronal activity during seizures is associated with positive BOLD and increases in CBF in cortex, but negative BOLD and decreased CBF in subcortical structures such as the striatum, despite equivalent LFP sizes [64]. The mechanisms for such variation are unclear, but are likely to involve differences in both signalling pathways and vascular anatomy [65]. For example, the involvement of nitric oxide (NO) varies between brain areas, both in terms of the magnitude of its contribution and whether it acts to mediate or modulate neurovascular coupling (as discussed in [66]). There also appear to be regional differences in the involvement of metabotropic glutamate receptor (mGluR) subtypes, as the mGluR5 antagonist MPEP decreases stimulation-evoked BOLD responses more in rat subcortical regions than in neocortex [67].

Furthermore, many enzymes involved in neurovascular coupling are developmentally regulated [68], and neuronal activity elicits vascular constriction [69-71] corresponding to negative BOLD responses in infants [72, 73]. The mechanisms underlying neurovascular coupling in a given situation will vary depending on expression patterns of key enzymes and receptors across different brain areas, as well as the local and global circuit dynamics engaged by a given task. For example, different subtypes of inhibitory interneurons presumably alter neurovascular coupling not only by release of different vasoactive molecules, but also by regulating the activity of the local network ([66]; Lecrux and Hamel in this issue), affecting glutamate release and second messenger production in excitatory neurons and astrocytes. The activity of these circuits and the reactivity of blood vessels can also be affected by global brain state as expressed in different levels of monoaminergic and cholinergic tone.

In short then, variability in neurovascular coupling mechanisms mean that the BOLD signal evoked by a given net increase in neuronal activity not only could vary in magnitude across different experimental conditions, ages, brain areas and brain states, but also could reflect different balances of circuit activity. How all these factors interact to generate the BOLD signal from changes in underlying neuronal activity has only been well-characterised during primary sensory processing, mostly in anaesthetised animals. Much further work is required to understand exactly how the neurovascular coupling relationship varies across the brain and therefore what exactly BOLD can inform us about underlying patterns of neuronal activity.

**F.** Changes in neurovascular coupling and MRI signals due to disease processes. In a number of neurological conditions, including stroke, glioma, subarachnoid haemorrhage, cortical spreading depolarizations, brain injury, hypertension, and neurodegenerative diseases such as Alzheimer's and Parkinson's disease, neuro-glio-vascular coupling may be altered or dysfunctional [74-77], altering the relationship between neuronal activity and changes in blood oxygenation. In this case, a decreased BOLD response to a task (as compared to healthy brain) may reflect a decrease in neural response, a decrease in neurovascular coupling, or both.

Neurodegenerative diseases may affect BOLD responses in various ways. There may be altered brain levels of neuromodulators/vasoactive molecules in the brain (e.g. the changes in dopamine levels that occur in Parkinson's disease) which could either directly impact BOLD signals or modulate the cellular and molecular mechanisms underlying neurovascular coupling. Reactive astrogliosis, which occurs in almost all neurological diseases [78, 79], is characterized by changes in the morphology and protein expression patterns of astrocytes [80, 81]. In particular, there are deficits in proteins involved in calcium dynamics, neurotransmitter sensing and K<sup>+</sup> buffering, all of which are

important in neurovascular signalling, and dysfunction in astrocyte-mediated neurovascular coupling in disease has been reported [82].

Pathological changes affecting BOLD responses are not restricted to changes in neurovascular coupling. Pathology can result in changes to magnetic resonance parameters, for example oedema and changes in diffusion, which will affect the number of water molecules and hence protons in the region of interest [83, 84], can generate artefacts or, when interpreted correctly, could serve as an important biomarker (Lake et al. in this issue). Furthermore, in the case of cerebrovascular pathology, the BOLD response may be altered in the absence of neuronal dysfunction, such as after acute injury, when a complete remapping of the vascular response may occur (see Lake et al., in this issue). Our lack of knowledge of how neurovascular coupling is altered in disease and pathology impacts our ability to accurately interpret BOLD data.

**G. Regional differences in BOLD signal magnitude.** To further complicate interpretation of BOLD signals, regional differences in the size of BOLD responses have been observed in different brain areas. In the hippocampus, for example, the relative difficulty in observing task-related activations may be due to cognitive factors (i.e. greater hippocampal activation at rest), vascular factors (a decreased capillary density in some subregions compared to neocortex) or altered patterns of neurovascular coupling ([65] and discussed above). Notably, human neocortical BOLD signals are expected to vary by as much as 40% across different cortical regions, simply due to the varying orientation of the vasculature relative to the magnetic field in the highly folded cortical structure ([85], further refined by Gagnon et al. in this issue). It will be important for future experiments to further investigate the regional variations in signal magnitudes and the underlying reasons for these, and to build these findings into fMRI analysis methods.

# 3. How can advances in fMRI/BOLD experimental design and analysis strengthen inferences of neuronal activity from BOLD signals?

**A. Model-based analyses.** BOLD generally reflects neuronal activity and is reasonably colocalised with the site of neuronal activation. Based on this understanding, early studies used BOLD to identify brain areas or networks where activity is modulated during a particular function [86, 87]. Reassuringly, there is an extremely high correspondence between response properties measured in fMRI and those measured from invasive electrical recordings or from homologous brain areas in nonhuman primates, for a very wide range of properties including visual object selectivity, orientation tuning, motor somatotopy and many others. fMRI has thus enabled a very detailed study of functional specialization, including those more difficult to study in non-human animals such as emotion, psychiatric disorders, abstract reasoning, autobiographical memory, meta-cognition, complex social cognition, language and consciousness [88]. However, such research concerning functional specialization failed to address questions of mechanism.

A key advance in going beyond early designs was to postulate formal models of brain computation, and test whether properties of these models are encoded in the brain (Lohrenz et al. in this issue, [89, 90]). By revealing *where* these quantities are encoded, we additionally gain linkages to other levels of neuroscience. For example, if a reward prediction error is sent from the midbrain to the striatum, we can form hypotheses about how plasticity at synapses in the striatum should work. As more aspects of the model can be linked to plausible neural computational mechanisms, and competing models eliminated, we can conclude whether our model is an accurate explanation of how the brain computes.

Many experiments in model-based neuroimaging have used psychological models with very few parameters that incorporate particular structural hypotheses. A recent, more open-ended approach is to use models with many parameters that are learned from large amounts of data, perhaps in a manner analogous to the brain. In particular, the success of deep neural networks in many engineering problems [91] has generated interest in these networks as models of the brain (Kriegeskorte et al. in this issue). For example, deep neural networks trained on natural images can better explain cell firing properties in monkey inferotemporal cortex than any other existing models [92]. These methods are now beginning to be applied to fMRI data [93, 94].

**B. Multivariate analyses.** In a similar vein, there has been a growing use of multivariate methods (Turner, in this issue), which trade spatial specificity for increased sensitivity, by looking for a signal in the pattern of activation across multiple voxels. These methods allow precise identification of *what* is encoded, in addition to some (albeit reduced) information about where it is encoded. By giving up spatial specificity, a resulting benefit is a minimized impact of co-registration errors between the unique brains of each participant. Unlike univariate methods, multivariate methods have the potential to capture information that exists only in the relationships between different voxels. This is important given that distributed representations are likely a central feature of information encoding by the brain.

A popular variety of multivariate analysis, called representational similarity analysis (RSA), goes a step further in abstraction by disregarding information about what particular patterns over voxels actually encode the signal of interest, and only considering the similarities between different patterns [95]. Thus, neural recordings made in different modalities can be analyzed in a common space, for example to combine the spatial resolution of fMRI with the temporal precision of MEG [96]. Neural recordings can even be analyzed in a common space with non-neuronal sources of information, such

as subjective reports [97] or the output of the deep neural networks mentioned above. RSA is particularly well suited to be combined with model-based approaches like deep neural networks which have many free parameters. Although the neural networks are too complex to map directly onto brain imaging data, the similarity structure of the neural network output can be compared directly to the similarity structure of BOLD data.

С. Towards insulating cognitive interpretations from the physiological details of *neurovascular coupling.* The use of model-based neuroimaging and multivariate analyses enables our conclusions to be quite robust against the details of neurovascular coupling. For example, a trained neural network closely matches the similarity structure of BOLD recorded in some parts of the brain. The conclusions that can be drawn from this do not depend on whether BOLD arises from spiking or local field potentials, and are not altered if BOLD is affected by pathology or neuromodulators. Effectively, methods whose interesting mechanics are at a more abstract level protect us, to a greater or lesser degree, from the underlying physiology compared to older analysis methods. For example, if multivoxel pattern analysis (MVPA) can discriminate between two remembered stimuli during memory retrieval based on the patterns of BOLD activation during encoding, we can be relatively confident that these patterns of activation relate to the neural signature rather than changes in vascular reactivity. It is worth noting that use of these analyses may minimise the impact of variations in neurovascular coupling created by disease states on the generation of false positives, but diseases could still increase the false negative rate (for example, by reducing the ability of neurons to activate a vascular response).

**D.** *fMRI adaptation.* In contrast to multivariate methods that probe the encoding of stimuli or model quantities across multiple voxels, fMRI adaptation can reveal encoding at a within-voxel level. When a stimulus is shown twice in succession, the BOLD response to the second presentation is often decreased compared to the first presentation. Such a decrease can also be observed if the second stimulus is related but not identical to the first stimulus, which is called cross-stimulus suppression. This effect is thought to arise at least in part from the diminished response of individual neurons to the second stimulus (Barron et al., in this issue).

This is particularly important because multivariate methods depend on a heterogeneous distribution of neuronal response properties across voxels. If every voxel contains 50 cells encoding stimulus A and 50 cells encoding stimulus B, it will be difficult to decode A vs. B even using the most sensitive multivariate analysis [98]. fMRI adaptation bypasses this problem by relying on the sensitivity of individual neurons to repeated activation (Barron et al., in this issue), and has yielded a wealth of new insights in cognitive neuroscience [99-102].

Interestingly, although fMRI adaptation aims to address similar questions of fine-grained representation as MVPA, cognitive interpretations of adaptation are much less likely to be insulated from physiological variation in neurovascular coupling. The underlying mechanism driving the adaptation is still not fully understood (as discussed by Barron et al. in this issue). The relationship between neuronal and vascular responses could be impacted by neuromodulators or other factors, which might produce false positives. Any such influences might also be heterogeneous across the brain. This is an important area for further study.

*E. High field strength.* Meanwhile, developments on the hardware side have increased both imaging resolution and our confidence in the results, by enabling single subject analyses. As discussed by Turner in this issue, high field MRI (7T and above) has many advantages over the lower field strengths which have predominated to date. High field MRI provides improved signal to noise, which can be traded for temporal or spatial resolution. The resulting smaller voxel sizes allow the resolution of structural details that were not previously visible, and the observed cortical microstructure can allow identification of different brain regions in each subject's brain. The ability to compare structure to function in individual subjects eliminates the need for both spatial smoothing of BOLD signals and comparison with standardised brains, allowing finer details of functional organisation to be resolved. Higher field strength favors detection of signals from capillaries (Turner, in this issue). In combination with improved spatial resolution, this has enabled detailed imaging of cortical layers [103-105] as studies are now more able to detect the local, capillary-level vascular changes that better map onto sites of neuronal activity. As discussed above, however, propagating dilation from the capillary bed can reduce this spatial resolution over time.

**F. Functional connectivity and resting state fMRI.** fMRI has facilitated, both in humans and animals, the study of functional interactions between distant brain areas [106-108]), something that was challenging to achieve with unit recordings. Initially, in studies focusing on functionally-evoked neural activity, spontaneous fluctuations observed in fMRI signals were regarded as noise (Mitra and Raichle, in this issue). However there is increasing evidence that brain areas displaying correlated spontaneous fluctuations represent the same regions active during task-related fMRI ([109, 110], reviewed by Ugurbil in this issue) and therefore constitute functional networks known as 'resting state networks' (RSNs). For example, fMRI signals in the posterior cingulate cortex, medial prefrontal cortex and angular gyrus fluctuate together during spontaneous activity and play a coordinated role in imagination, episodic memory and theory of mind. More recently, temporal lags between propagating fMRI signals during resting state have also been investigated to identify the direction of connectivity between brain areas co-activated closely in time (Mitra and Raichle, in this issue). Our understanding of RSNs, both in space and time, is expected to improve as higher field strengths with

better signal-to-noise ratio are employed, e.g. by making it possible to detect more components with possibly new functional significance [111].

Understanding network organization also provides clues about the structure of computations in the brain. For example, some brain regions, called hubs, are functionally connected to many other regions, suggesting they may coordinate computations at a broader scale [112, 113]. This interpretation is further supported by distinct patterns of expression in hub regions of genes supporting metabolism and inter-cortical connections (Vertes et al. in this issue). Such studies can therefore assist investigations into disruptions of functional connectivity in neurodegenerative and psychiatric disorders [107].

The spatial and temporal relationship of RSNs is assumed to arise from activation of neural connections between brain regions and recent data demonstrate that electrophysiological recordings can predict RSNs in the human brain to some extent [114]. However, it is unclear whether these patterns of neural activity solely reflect functional connectivity, or could be influenced by the vasculature. For example, regional differences in the haemodynamic response could contaminate RSNs [115-118], while concurrent modulation of neurons and the vasculature in different brain regions by ascending monoaminergic and cholinergic projection systems could create apparent connectivity in the absence of direct functional connections (discussed above; [63, 119]) when these systems are differently engaged, for example during regulation of arousal states [120, 121]. Although it is possible that vasomotion intrinsic to the vasculature might distort RSN signals [122], functional connectivity patterns have recently been successfully separated from vasomotion by applying low band-pass filters [123]. Furthermore, neurons tend to generate random spontaneous activity during 'rest' (meaning the absence of task-evoked activity) due to their intrinsically excitable nature [124], giving rise to small haemodynamic responses. Such fluctuations might represent actual functional connections between neurons [125], or may be due to spontaneous noise (which nevertheless may be computationally beneficial due to stochastic resonance [126]). More studies employing multimodal approaches, both in animal models and humans, are required to test how reliably fMRI based RSNs reflect the underlying spontaneous neural activity and the nature of the relationship between them.

**G. Multi-modal Imaging Approaches.** Inferences from BOLD experiments can be confirmed or augmented by using a multi-modal imaging approach. fMRI results have been replicated using EEG or MEG (often with spatial specificity through source localization algorithms) while magnetic resonance spectroscopy (MRS) has been used to add detail of neurotransmitter contributions to circuit-level mechanisms [100]. In addition to applying MRS, Barron et al. [100], applied transcranial direct current stimulation with fMRI adaptation to test, at the cellular population level, hypotheses

based on BOLD experiments. Performing multi-modal MRI enhances the information we can obtain from a study involving only BOLD. For example, by combining BOLD and arterial spin labelling, allowing measurement of blood oxygen level dependent signals and cerebral blood flow, respectively, the coupling between blood flow and brain energy metabolism can be calculated [127]. As the amplitude of the BOLD response is dependent on baseline physiology, i.e. oxygen extraction fraction, cerebral blood volume and hematocrit levels, it is difficult to interpret changes in BOLD in situations where physiology may be varied. This has led to the development of calibrated BOLD fMRI [128] in which a calibration scan (typically involving a hypercapnic breathing challenge) is applied with the aim of accounting for physiological variations (Blockley et al. [129] review the different methods of performing this calibration).

Although in general we must rely on using non-invasive methods (such as fMRI) in human subjects, it has been possible in some patient groups to undertake invasive recordings. Microelectrode recordings of single neurons in the human brain have allowed access to cell response properties, supporting inferences of some BOLD experiments (e.g. the existence of grid cells in entorhinal cortex; [130]), while invasive chemical recordings of neurotransmitters such as dopamine and serotonin have expanded hypotheses based on BOLD results to implicate specific neurotransmitters in cognitive processes ([131], Lohrenz et al. in this issue). Such invasive recordings can, of course, only be obtained, with consent, from patients undergoing neurosurgery for other reasons, so such methods are understandably not widely available as a tool for research.

While in humans we are mostly restricted to estimating neural activity from an indirect, non-invasive measurement such as the BOLD signal, in animal studies the use of invasive techniques allows us to concurrently measure neural activity and haemodynamics. This approach enables a thorough investigation of neurovascular coupling and the mechanisms which relate neural activity to the BOLD signal. A wealth of information has already been acquired from detailed, invasive animal studies [132] and our understanding of how BOLD fMRI signals relate to neural signals is set to increase as newly developed techniques, such as genetically encoded fluorescent proteins and improved hardware, are increasingly coupled with existing imaging approaches, such as wide field optical mapping techniques (Ma et al. in this issue). The key is to use these data from animal studies to aid our interpretation of non-invasive signals in humans. In this issue, Uhlirova et al. propose that by developing models that estimate non-invasive imaging signals from the population activity of specific neurons, we can establish a "ground truth" that can be used to estimate neural activity from non-invasive data in humans. Multi-modal imaging is a powerful tool for increasing our understanding of the relationship between neural activity and BOLD fMRI signals, and vice versa.

As discussed above, an exciting development is recent work combining fMRI with information from gene expression databases such as the Allen Institute, in order to understand functional differences between brain regions and networks detected using fMRI (Vertes et al. in this issue). Interestingly, these results suggest differences in energy use between different functional nodes, and these hubs may be a useful future target for neurovascular coupling studies, to investigate if neurovascular coupling properties vary with net energy demand of a brain region. Functional changes associated with specific genes can also be investigated by stratifying subjects according to genotype and testing the effect of genetic variation.

### 4. Future directions to enable better interpretation of BOLD signals

To fully understand the physiological basis of the BOLD signal, and therefore be able to optimise experimental design and interpretation, we need a complete understanding of the processes that generate it at the molecular, cellular, vascular and voxel level. While, as discussed above, we are still some way from this complete understanding, there are certain areas where research should be focussed to best increase our ability to confidently interpret BOLD:

I: To account for regional differences in BOLD signals, bottom-up modelling of BOLD responses based on vascular anatomical networks could be extended to human vascular anatomy. Incorporation of predicted differences in signal strength due to vascular anatomy alone could then be incorporated into fMRI analysis software.

II: Physiological research on the properties of neurovascular coupling should investigate the cell types responsible, modulation by neurotransmitter projection systems (and therefore brain states), and how consistent these properties are across different brain regions, from neocortex to sub-cortical structures.

III: fMRI analysis tools could allow more flexibility for modulation of the haemodynamic response function, to allow the effects of variations in neurovascular coupling properties on the BOLD response amplitude and time course to be reflected in fMRI analyses.

IV: Multi-modal experiments in humans and animals should investigate the physiological basis of phenomena such as resting state BOLD and fMRI adaptation that are increasingly being used to infer cognitive functions, but are understudied at the physiological level.

### Conclusion

Recent research has gone some way to elucidating the cellular mechanisms that underlie the BOLD signal, revealing a multitude of cell types and signalling pathways that coordinate the vascular response to neuronal activity. These pathways link blood flow increases primarily with increased synaptic activity and, by defining the spread of the vascular response from the site of neuronal activity, set the minimum spatiotemporal resolution that could be achieved with BOLD.

Within these pathways, however, there is a huge potential for variation in the properties of neurovascular responses between different brain regions, at different developmental stages, in conditions of altered local or global circuit dynamics, or during disease states, as reflected in the observed uncoupling of neural activity and BOLD in subcortical structures and alterations in predicted BOLD between human neocortical areas. Caution is therefore required when interpreting BOLD signals that could be affected by such factors, particularly using experimental paradigms (such as resting state fMRI and fMRI adaptation) that have been less studied using multimodal methods that allow measurement of both neuronal and vascular responses. Future research should characterise these multimodal responses across the wide variety of paradigms currently used in fMRI research, to establish when BOLD does and does not provide useful information about underlying neuronal activity.

Developments in fMRI analysis and hardware are increasing the potential for fMRI/BOLD to inform us about cognitive processes. Model-based fMRI analyses partly mitigate the effect of uncertain underlying physiology by testing whether a given predicted neuronal response is present in the spatial or temporal BOLD signature, while increases in magnetic field strength both reduce the requirement for spatial smoothing and allow prioritisation of the signal from the blood vessels closest to the site of neuronal activity.

If we can incorporate our increasing understanding of cellular processes and the limits they impose on BOLD signals into fMRI analysis methods, we will enhance the power of these analyses and thus enable greater understanding of the cognitive processes that are monitored using BOLD. Continued dialogue between cellular and cognitive neuroscientists, as initiated by the 2016 Royal Society Theo Murphy meeting, will be critical to achieving this ambition.

### References

- 1. Attwell, D. and C. Iadecola, *The neural basis of functional brain imaging signals.* Trends Neurosci, 2002. **25**(12): p. 621-5.
- Buxton, R.B., *The physics of functional magnetic resonance imaging (fMRI)*. Rep Prog Phys, 2013. **76**(9): p. 096601.
- Attwell, D., et al., *Glial and neuronal control of brain blood flow.* Nature, 2010. 468(7321): p. 232-243.
- 4. Logothetis, N.K., et al., *Neurophysiological investigation of the basis of the fMRI signal.* Nature, 2001. **412**(6843): p. 150-7.
- 5. Goense, J.B. and N.K. Logothetis, *Laminar specificity in monkey V1 using high-resolution SE-fMRI.* Magn Reson Imaging, 2006. **24**(4): p. 381-92.
- Mathiesen, C., et al., Modification of activity-dependent increases of cerebral blood flow by excitatory synaptic activity and spikes in rat cerebellar cortex. J Physiol, 1998. 512 (Pt 2): p. 555-66.
- Thomsen, K., N. Offenhauser, and M. Lauritzen, *Principal neuron spiking: neither necessary nor sufficient for cerebral blood flow in rat cerebellum.* J Physiol, 2004. 560(Pt 1): p. 181-9.
- 8. Howarth, C., P. Gleeson, and D. Attwell, *Updated energy budgets for neural computation in the neocortex and cerebellum.* J Cereb Blood Flow Metab, 2012. **32**(7): p. 1222-32.
- Hall, C.N., et al., Oxidative phosphorylation, not glycolysis, powers presynaptic and postsynaptic mechanisms underlying brain information processing. J Neurosci, 2012. 32(26): p. 8940-51.
- 10. Bednar, J.A. and S.P. Wilson, *Cortical Maps*. Neuroscientist, 2015.
- 11. Bagher, P. and S.S. Segal, *Regulation of blood flow in the microcirculation: role of conducted vasodilation.* Acta Physiol (Oxf), 2011. **202**(3): p. 271-84.
- 12. Chen, B.R., et al., *High-speed vascular dynamics of the hemodynamic response.* Neuroimage, 2011. **54**(2): p. 1021-30.
- 13. Yu, X., et al., *Direct imaging of macrovascular and microvascular contributions to BOLD fMRI in layers IV-V of the rat whisker-barrel cortex.* Neuroimage, 2012. **59**(2): p. 1451-60.
- 14. Scholvinck, M.L., C. Howarth, and D. Attwell, *The cortical energy needed for conscious perception.* Neuroimage, 2008. **40**(4): p. 1460-8.

- 15. Ogawa, S., et al., *Brain magnetic resonance imaging with contrast dependent on blood oxygenation.* Proc Natl Acad Sci U S A, 1990. **87**(24): p. 9868-72.
- 16. Roy, C.S. and C.S. Sherrington, *On the Regulation of the Blood-supply of the Brain.* J Physiol, 1890. 11(1-2): p. 85-158 17.
- 17. Hillman, E.M., *Coupling mechanism and significance of the BOLD signal: a status report.*Annu Rev Neurosci, 2014. **37**: p. 161-81.
- Braitenberg V., S.A., *Cortex: Statistics and geometry of neuronal connectivity, 2nd ed.* 2nd ed. 1998: Berlin: Springer.
- Fergus, A. and K.S. Lee, *Regulation of cerebral microvessels by glutamatergic mechanisms.* Brain Res, 1997. **754**(1-2): p. 35-45.
- Harris, J.J., R. Jolivet, and D. Attwell, *Synaptic energy use and supply*. Neuron, 2012. **75**(5):
   p. 762-77.
- 21. Attwell, D. and S.B. Laughlin, *An energy budget for signaling in the grey matter of the brain.* J Cereb Blood Flow Metab, 2001. **21**(10): p. 1133-45.
- 22. Ma, J., et al., *Regional cerebral blood flow response to vibrissal stimulation in mice lacking type I NOS gene expression.* Am J Physiol, 1996. **270**(3 Pt 2): p. H1085-90.
- 23. Niwa, K., et al., *Cyclooxygenase-2 contributes to functional hyperemia in whisker-barrel cortex.* J Neurosci, 2000. **20**(2): p. 763-70.
- 24. Cauli, B., et al., *Cortical GABA interneurons in neurovascular coupling: relays for subcortical vasoactive pathways.* J Neurosci, 2004. **24**(41): p. 8940-9.
- 25. Filosa, J.A., et al., *Local potassium signaling couples neuronal activity to vasodilation in the brain.* Nat Neurosci, 2006. **9**(11): p. 1397-1403.
- 26. Zonta, M., et al., *Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation.* Nat.Neurosci., 2003. **6**(1): p. 43-50.
- 27. Mulligan, S.J. and B.A. Macvicar, *Calcium transients in astrocyte endfeet cause cerebrovascular constrictions.* Nature, 2004. **431**(7005): p. 195-199.
- 28. Gordon, G.R., et al., *Brain metabolism dictates the polarity of astrocyte control over arterioles.* Nature, 2008. **456**(7223): p. 745-749.
- 29. Metea, M.R. and E.A. Newman, *Glial cells dilate and constrict blood vessels: a mechanism of neurovascular coupling.* J.Neurosci., 2006. **26**(11): p. 2862-2870.
- 30. Mishra, A., A. Hamid, and E.A. Newman, *Oxygen modulation of neurovascular coupling in the retina.* Proc Natl Acad Sci U S A, 2011. **108**(43): p. 17827-31.

- 31. Nizar, K., et al., In vivo stimulus-induced vasodilation occurs without IP3 receptor activation and may precede astrocytic calcium increase. J.Neurosci., 2013. **33**(19): p. 8411-8422.
- 32. Schulz, K., et al., *Simultaneous BOLD fMRI and fiber-optic calcium recording in rat neocortex.* Nat Methods, 2012. **9**(6): p. 597-602.
- 33. Takata, N., et al., Cerebral blood flow modulation by Basal forebrain or whisker stimulation can occur independently of large cytosolic Ca2+ signaling in astrocytes. PLoS One, 2013.
   8(6): p. e66525.
- 34. Bonder, D.E. and K.D. McCarthy, *Astrocytic Gq-GPCR-linked IP3R-dependent Ca2+ signaling does not mediate neurovascular coupling in mouse visual cortex in vivo.* J.Neurosci., 2014. **34**(39): p. 13139-13150.
- 35. Rosenegger, D.G. and G.R. Gordon, *A slow or modulatory role of astrocytes in neurovascular coupling.* Microcirculation, 2015. **22**(3): p. 197-203.
- 36. Calcinaghi, N., et al., *Metabotropic glutamate receptor mGluR5 is not involved in the early hemodynamic response.* J Cereb Blood Flow Metab, 2011. **31**(9): p. e1-10.
- 37. Rosenegger, D.G., et al., *Tonic Local Brain Blood Flow Control by Astrocytes Independent of Phasic Neurovascular Coupling.* J Neurosci, 2015. **35**(39): p. 13463-74.
- 38. Kur, J. and E.A. Newman, *Purinergic control of vascular tone in the retina*. J Physiol, 2014.
  592(3): p. 491-504.
- 39. Kim, K.J., et al., *Astrocyte contributions to flow/pressure-evoked parenchymal arteriole vasoconstriction.* J Neurosci, 2015. **35**(21): p. 8245-57.
- 40. Blanco, V.M., J.E. Stern, and J.A. Filosa, *Tone-dependent vascular responses to astrocytederived signals.* Am J Physiol Heart Circ Physiol, 2008. **294**(6): p. H2855-63.
- 41. Peppiatt, C.M., et al., *Bidirectional control of CNS capillary diameter by pericytes*. Nature, 2006. **443**(7112): p. 700-704.
- 42. Hall, C.N., et al., *Capillary pericytes regulate cerebral blood flow in health and disease.* Nature, 2014. **508**(7494): p. 55-60.
- 43. Fernández-Klett, F., et al., *Pericytes in capillaries are contractile in vivo, but arterioles mediate functional hyperemia in the mouse brain.* Proc.Natl.Acad.Sci.U.S.A, 2010. 107(51): p. 22290-22295.

- 44. Hill, R.A., et al., Regional Blood Flow in the Normal and Ischemic Brain Is Controlled by Arteriolar Smooth Muscle Cell Contractility and Not by Capillary Pericytes. Neuron, 2015.
  87(1): p. 95-110.
- 45. Attwell, D., et al., *What is a pericyte?* J Cereb Blood Flow Metab, 2016. **36**(2): p. 451-5.
- 46. Hartmann, D.A., et al., *Pericyte structure and distribution in the cerebral cortex revealed by high-resolution imaging of transgenic mice.* Neurophotonics, 2015. **2**(4): p. 041402.
- 47. Sheth, S.A., et al., *Columnar specificity of microvascular oxygenation and volume responses: implications for functional brain mapping.* J Neurosci, 2004. **24**(3): p. 634-41.
- 48. Blinder, P., et al., *The cortical angiome: an interconnected vascular network with noncolumnar patterns of blood flow.* Nat Neurosci, 2013. **16**(7): p. 889-97.
- 49. Hamilton, N.B., D. Attwell, and C.N. Hall, *Pericyte-mediated regulation of capillary diameter: a component of neurovascular coupling in health and disease.* Front Neuroenergetics, 2010. **2**.
- 50. Chen, B.R., et al., *A critical role for the vascular endothelium in functional neurovascular coupling in the brain.* J Am Heart Assoc, 2014. **3**(3): p. e000787.
- 51. Xu, H.L. and D.A. Pelligrino, *ATP release and hydrolysis contribute to rat pial arteriolar dilatation elicited by neuronal activation.* Exp Physiol, 2007. **92**(4): p. 647-51.
- 52. O'Herron, P., et al., *Neural correlates of single-vessel haemodynamic responses in vivo.* Nature, 2016.
- 53. Krimer, L.S., et al., *Dopaminergic regulation of cerebral cortical microcirculation*. Nat Neurosci, 1998. **1**(4): p. 286-9.
- 54. Hamel, E., *Cholinergic modulation of the cortical microvascular bed.* Prog Brain Res, 2004.
  145: p. 171-8.
- 55. Hamel, E., *Perivascular nerves and the regulation of cerebrovascular tone.* J Appl Physiol (1985), 2006. **100**(3): p. 1059-64.
- 56. Cohen, Z., G. Molinatti, and E. Hamel, *Astroglial and vascular interactions of noradrenaline terminals in the rat cerebral cortex.* J Cereb Blood Flow Metab, 1997. **17**(8): p. 894-904.
- 57. Pisauro, M.A., A. Benucci, and M. Carandini, *Local and global contributions to hemodynamic activity in mouse cortex.* J Neurophysiol, 2016. **115**(6): p. 2931-6.
- 58. Vaucher, E. and E. Hamel, *Cholinergic basal forebrain neurons project to cortical microvessels in the rat: electron microscopic study with anterogradely transported*

*Phaseolus vulgaris leucoagglutinin and choline acetyltransferase immunocytochemistry.* J Neurosci, 1995. **15**(11): p. 7427-41.

- 59. Chedotal, A., et al., Light and electron microscopic immunocytochemical analysis of the neurovascular relationships of choline acetyltransferase and vasoactive intestinal polypeptide nerve terminals in the rat cerebral cortex. J Comp Neurol, 1994. **343**(1): p. 57-71.
- 60. Hotta, H., et al., *Control of cerebral cortical blood flow by stimulation of basal forebrain cholinergic areas in mice.* J Physiol Sci, 2011. **61**(3): p. 201-9.
- 61. Kocharyan, A., et al., *Specific subtypes of cortical GABA interneurons contribute to the neurovascular coupling response to basal forebrain stimulation.* J Cereb Blood Flow Metab, 2008. **28**(2): p. 221-31.
- 62. Bekar, L.K., H.S. Wei, and M. Nedergaard, *The locus coeruleus-norepinephrine network optimizes coupling of cerebral blood volume with oxygen demand.* J Cereb Blood Flow Metab, 2012. **32**(12): p. 2135-45.
- 63. Toussay, X., et al., *Locus coeruleus stimulation recruits a broad cortical neuronal network and increases cortical perfusion.* J Neurosci, 2013. **33**(8): p. 3390-401.
- 64. Mishra, A.M., et al., *Where fMRI and electrophysiology agree to disagree: corticothalamic and striatal activity patterns in the WAG/Rij rat.* J Neurosci, 2011. **31**(42): p. 15053-64.
- 65. Ekstrom, A., *How and when the fMRI BOLD signal relates to underlying neural activity: the danger in dissociation.* Brain Res Rev, 2010. **62**(2): p. 233-44.
- 66. Duchemin, S., et al., *The complex contribution of NOS interneurons in the physiology of cerebrovascular regulation.* Front Neural Circuits, 2012. **6**: p. 51.
- 67. Sloan, H.L., et al., *Regional differences in neurovascular coupling in rat brain as determined by fMRI and electrophysiology.* Neuroimage, 2010. **53**(2): p. 399-411.
- 68. Harris, J.J., C. Reynell, and D. Attwell, *The physiology of developmental changes in BOLD functional imaging signals.* Dev Cogn Neurosci, 2011. **1**(3): p. 199-216.
- 69. Kozberg, M.G., et al., *Resolving the transition from negative to positive blood oxygen leveldependent responses in the developing brain.* Proc Natl Acad Sci U S A, 2013. **110**(11): p. 4380-5.
- 70. Kozberg, M. and E. Hillman, *Neurovascular coupling and energy metabolism in the developing brain.* Prog Brain Res, 2016. **225**: p. 213-42.

- 71. Zehendner, C.M., et al., *Developmental switch in neurovascular coupling in the immature rodent barrel cortex.* PLoS One, 2013. **8**(11): p. e80749.
- 72. Anderson, A.W., et al., *Neonatal auditory activation detected by functional magnetic resonance imaging.* Magn Reson Imaging, 2001. **19**(1): p. 1-5.
- Born, A.P., et al., *Visual cortex reactivity in sedated children examined with perfusion MRI (FAIR).* Magn Reson Imaging, 2002. 20(2): p. 199-205.
- 74. Girouard, H. and C. Iadecola, *Neurovascular coupling in the normal brain and in hypertension, stroke, and Alzheimer disease.* J Appl Physiol (1985), 2006. 100(1): p. 328-35.
- 75. Nelson, A.R., et al., *Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease.* Biochim Biophys Acta, 2016. **1862**(5): p. 887-900.
- 76. Zlokovic, B.V., *Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders.* Nat Rev Neurosci, 2011. **12**(12): p. 723-38.
- 77. Dalkara, T. and L. Alarcon-Martinez, *Cerebral microvascular pericytes and neurogliovascular signaling in health and disease.* Brain Res, 2015. **1623**: p. 3-17.
- 78. Barres, B.A., *The mystery and magic of glia: a perspective on their roles in health and disease.* Neuron, 2008. **60**(3): p. 430-40.
- 79. Anderson, M.A., Y. Ao, and M.V. Sofroniew, *Heterogeneity of reactive astrocytes.* Neurosci Lett, 2014. **565**: p. 23-9.
- Hamby, M.E., et al., Inflammatory mediators alter the astrocyte transcriptome and calcium signaling elicited by multiple G-protein-coupled receptors. J Neurosci, 2012. 32(42): p. 14489-510.
- 81. Zamanian, J.L., et al., *Genomic analysis of reactive astrogliosis*. J Neurosci, 2012. **32**(18): p. 6391-410.
- 82. Mishra, A. and E.A. Newman, *Inhibition of inducible nitric oxide synthase reverses the loss of functional hyperemia in diabetic retinopathy.* Glia, 2010. **58**(16): p. 1996-2004.
- 83. Krings, T., et al., *Factors related to the magnitude of T2\* MR signal changes during functional imaging.* Neuroradiology, 2002. **44**(6): p. 459-66.
- 84. Kim, S.G. and S. Ogawa, *Biophysical and physiological origins of blood oxygenation leveldependent fMRI signals.* J Cereb Blood Flow Metab, 2012. **32**(7): p. 1188-206.

- 85. Gagnon, L., et al., *Quantifying the microvascular origin of BOLD-fMRI from first principles with two-photon microscopy and an oxygen-sensitive nanoprobe.* J Neurosci, 2015. 35(8): p. 3663-75.
- 86. Zeki, S., et al., *A direct demonstration of functional specialization in human visual cortex.* J Neurosci, 1991. **11**(3): p. 641-9.
- 87. Kanwisher, N., *Functional specificity in the human brain: a window into the functional architecture of the mind.* Proc Natl Acad Sci U S A, 2010. **107**(25): p. 11163-70.
- 88. Gazzaniga, M.S., *The Cognitive Neurosciences*. 4 ed. 2009, Cambridge, MA: The MIT Press.
- 89. O'Doherty, J.P., *Lights, camembert, action! The role of human orbitofrontal cortex in encoding stimuli, rewards, and choices.* Ann N Y Acad Sci, 2007. **1121**: p. 254-72.
- 90. Mars, R.B., et al., *Model-based analyses: Promises, pitfalls, and example applications to the study of cognitive control.* Q J Exp Psychol (Hove), 2012. **65**(2): p. 252-67.
- 91. LeCun, Y., Y. Bengio, and G. Hinton, *Deep learning*. Nature, 2015. **521**(7553): p. 436-44.
- 92. Yamins, D.L., et al., *Performance-optimized hierarchical models predict neural responses in higher visual cortex.* Proc Natl Acad Sci U S A, 2014. **111**(23): p. 8619-24.
- 93. Khaligh-Razavi, S.M. and N. Kriegeskorte, *Deep supervised, but not unsupervised, models may explain IT cortical representation.* PLoS Comput Biol, 2014. **10**(11): p. e1003915.
- 94. Guclu, U. and M.A. van Gerven, *Deep Neural Networks Reveal a Gradient in the Complexity of Neural Representations across the Ventral Stream.* J Neurosci, 2015. 35(27): p. 10005-14.
- 95. Kriegeskorte, N., M. Mur, and P. Bandettini, *Representational similarity analysis connecting the branches of systems neuroscience.* Front Syst Neurosci, 2008. **2**: p. 4.
- 96. Cichy, R.M., D. Pantazis, and A. Oliva, *Resolving human object recognition in space and time.* Nat Neurosci, 2014. **17**(3): p. 455-62.
- 97. Charest, I., et al., *Unique semantic space in the brain of each beholder predicts perceived similarity.* Proc Natl Acad Sci U S A, 2014. **111**(40): p. 14565-70.
- 98. Dubois, J., A.O. de Berker, and D.Y. Tsao, *Single-unit recordings in the macaque face patch system reveal limitations of fMRI MVPA.* J Neurosci, 2015. **35**(6): p. 2791-802.
- 99. Barron, H.C., R.J. Dolan, and T.E. Behrens, *Online evaluation of novel choices by simultaneous representation of multiple memories.* Nat Neurosci, 2013. **16**(10): p. 1492-8.
- 100. Barron, H.C., et al., Unmasking Latent Inhibitory Connections in Human Cortex to Reveal Dormant Cortical Memories. Neuron, 2016. **90**(1): p. 191-203.

- Boorman, E.D., et al., *Two Anatomically and Computationally Distinct Learning Signals Predict Changes to Stimulus-Outcome Associations in Hippocampus.* Neuron, 2016. **89**(6): p. 1343-54.
- 102. Garvert, M.M., et al., *Learning-induced plasticity in medial prefrontal cortex predicts preference malleability.* Neuron, 2015. **85**(2): p. 418-28.
- 103. Zhao, F., et al., Cortical layer-dependent BOLD and CBV responses measured by spin-echo and gradient-echo fMRI: insights into hemodynamic regulation. Neuroimage, 2006. 30(4): p. 1149-60.
- 104. Olman, C.A., et al., *Layer-specific fMRI reflects different neuronal computations at different depths in human V1.* PLoS One, 2012. **7**(3): p. e32536.
- 105. Augustinack, J.C., et al., *Detection of entorhinal layer II using 7Tesla [corrected] magnetic resonance imaging.* Ann Neurol, 2005. **57**(4): p. 489-94.
- 106. Fox, M.D. and M.E. Raichle, *Spontaneous fluctuations in brain activity observed with functional magnetic resonance imaging.* Nat Rev Neurosci, 2007. **8**(9): p. 700-11.
- 107. van den Heuvel, M.P. and H.E. Hulshoff Pol, *Exploring the brain network: a review on resting-state fMRI functional connectivity.* Eur Neuropsychopharmacol, 2010. 20(8): p. 519-34.
- 108. Gozzi, A. and A.J. Schwarz, *Large-scale functional connectivity networks in the rodent brain.* Neuroimage, 2016. **127**: p. 496-509.
- 109. Biswal, B., et al., *Functional connectivity in the motor cortex of resting human brain using echo-planar MRI.* Magn Reson Med, 1995. **34**(4): p. 537-41.
- 110. Smith, S.M., et al., *Correspondence of the brain's functional architecture during activation and rest.* Proc Natl Acad Sci U S A, 2009. **106**(31): p. 13040-5.
- 111. Koopmans, P.J., et al., *Whole brain, high resolution spin-echo resting state fMRI using PINS multiplexing at 7 T.* Neuroimage, 2012. **62**(3): p. 1939-46.
- 112. Bullmore, E. and O. Sporns, *Complex brain networks: graph theoretical analysis of structural and functional systems.* Nat Rev Neurosci, 2009. **10**(3): p. 186-98.
- 113. He, Y. and A. Evans, *Graph theoretical modeling of brain connectivity.* Curr Opin Neurol, 2010. 23(4): p. 341-50.
- 114. Tewarie, P., et al., *Predicting haemodynamic networks using electrophysiology: The role of non-linear and cross-frequency interactions.* Neuroimage, 2016. **130**: p. 273-92.

- Schippers, M.B., R. Renken, and C. Keysers, *The effect of intra- and inter-subject variability* of hemodynamic responses on group level Granger causality analyses. Neuroimage, 2011.
   57(1): p. 22-36.
- 116. Smith, S.M., et al., *The danger of systematic bias in group-level FMRI-lag-based causality estimation.* Neuroimage, 2012. **59**(2): p. 1228-9.
- 117. Smith, S.M., et al., *Network modelling methods for FMRI*. Neuroimage, 2011. 54(2): p. 875-91.
- 118. Wen, X., G. Rangarajan, and M. Ding, *Is Granger causality a viable technique for analyzing fMRI data?* PLoS One, 2013. **8**(7): p. e67428.
- 119. Vaucher, E., D. Linville, and E. Hamel, *Cholinergic basal forebrain projections to nitric oxide synthase-containing neurons in the rat cerebral cortex.* Neuroscience, 1997. **79**(3): p. 827-36.
- Scholvinck, M.L., et al., *Neural basis of global resting-state fMRI activity*. Proc Natl Acad Sci U S A, 2010. **107**(22): p. 10238-43.
- 121. Chang, C., et al., *Tracking brain arousal fluctuations with fMRI*. Proc Natl Acad Sci U S A, 2016. 113(16): p. 4518-23.
- 122. Rayshubskiy, A., et al., Direct, intraoperative observation of ~0.1 Hz hemodynamic oscillations in awake human cortex: implications for fMRI. Neuroimage, 2014. 87: p. 323-31.
- 123. Bumstead, J.R., et al., *Cerebral functional connectivity and Mayer waves in mice: Phenomena and separability.* J Cereb Blood Flow Metab, 2016.
- 124. Desai, N.S., L.C. Rutherford, and G.G. Turrigiano, *Plasticity in the intrinsic excitability of cortical pyramidal neurons.* Nat Neurosci, 1999. **2**(6): p. 515-20.
- 125. Scholvinck, M.L., K.J. Friston, and G. Rees, *The influence of spontaneous activity on stimulus processing in primary visual cortex.* Neuroimage, 2012. **59**(3): p. 2700-8.
- 126. McDonnell, M.D. and D. Abbott, *What is stochastic resonance? Definitions, misconceptions, debates, and its relevance to biology.* PLoS Comput Biol, 2009. **5**(5): p. e1000348.
- 127. Leontiev, O. and R.B. Buxton, *Reproducibility of BOLD, perfusion, and CMRO2 measurements with calibrated-BOLD fMRI.* Neuroimage, 2007. **35**(1): p. 175-84.
- 128. Davis, T.L., et al., *Calibrated functional MRI: mapping the dynamics of oxidative metabolism.* Proc Natl Acad Sci U S A, 1998. **95**(4): p. 1834-9.

- 129. Blockley, N.P., et al., *A review of calibrated blood oxygenation level-dependent (BOLD) methods for the measurement of task-induced changes in brain oxygen metabolism.* NMR Biomed, 2013. **26**(8): p. 987-1003.
- 130. Jacobs, J., et al., *Direct recordings of grid-like neuronal activity in human spatial navigation.* Nat Neurosci, 2013. **16**(9): p. 1188-90.
- 131. Kishida, K.T., et al., Subsecond dopamine fluctuations in human striatum encode superposed error signals about actual and counterfactual reward. Proc Natl Acad Sci U S A, 2016. 113(1): p. 200-5.
- 132. Martin, C., *Contributions and complexities from the use of in vivo animal models to improve understanding of human neuroimaging signals.* Front Neurosci, 2014. **8**: p. 211.