

This is a repository copy of Skp is a multivalent chaperone of outer membrane proteins.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/101494/

Version: Supplemental Material

Article:

Schiffrin, B, Calabrese, AN, Devine, PWA et al. (4 more authors) (2016) Skp is a multivalent chaperone of outer membrane proteins. Nature Structural and Molecular Biology, 23 (9). pp. 786-793. ISSN 1545-9993

https://doi.org/10.1038/nsmb.3266

(c) 2016, Nature American, Inc. This is an author produced version of a paper published in Nature Structural & Molecular Biology. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/



Example kinetic traces for (a) tOmpA, (b) PagP, (c) OmpF, (d) tBamA and (e) OmpT, in the absence of lipids, monitored by fluorescence emission spectroscopy. Assays were performed with OMP concentrations of 0.4 μM, in 0.24 M urea, 50 mM glycine-NaOH, pH 9.5, at 25 °C. At least three replicates are shown for each protein. (f) Crystal structure of OmpT, PDB:1178 (Vandeputte-Rutten, L. et al. *EMBO J* 20 5033-9, 2001). Tryptophan residues are shown in stick representation and highlighted in red. The data for OmpT are well described by a single exponential indicated by black dashed lines.



16-stranded OMPs require pre-incubation with a greater molar excess of Skp than 8-stranded OMPs to inhibit folding into synthetic liposomes.

Example kinetic folding data for (a) tOmpA, left panel and tOmpA in the presence of a 1:1, centre panel, or 2:1, right panel, molar ratio of Skp:tOmpA; (b) PagP, left panel and PagP in the presence of a 1:1, centre panel, or 2:1, right panel, molar ratio of Skp:tOmpA; (c) OmpF, left panel and OmpF in the presence of a 2:1, centre panel, or 4:1, right panel, molar excess of Skp; (d) tBamA, left panel and tBamA in the presence of a 2:1, centre panel, or 4:1, right panel, molar excess of Skp. OMP complexes were added to $diC_{11:0}$ PC liposomes and OMP folding was monitored by fluorescence spectroscopy. Final OMP concentrations were 0.4 μ M, with a molar lipid:protein ratio of 3200:1, in 0.24 M urea, 50 mM glycine-NaOH, pH 9.5. A minimum of three transients are shown in each panel. Single or double exponential fits to the data are indicated by black dashed lines (see Supplementary Table 2).



Complexes of Skp with OMPs have variable stoichiometries revealed by ESI-IMS-MS.

IMS driftscope plots of (a) Skp and Skp:OMP complexes with (b) tOmpA, (c) PagP, (d) OmpT, (e) OmpF or (f) tBamA. Peaks are labelled with their charge state. Charge states corresponding to Skp, 1:1 Skp:OMP and 2:1 Skp:OMP are labelled in red, green and blue, respectively. (g) CCS distributions (peak heights normalized to MS peak intensity) of 2:1 Skp:OMP complexes with OmpT (left), OmpF (middle), and tBamA (right) obtained from ESI-IMS-MS analyses. The charge state for each CCS distribution is indicated.



ESI-MS shows that two copies of Skp bind to full-length BamA.

(a) Non-covalent ESI mass spectrum of full-length BamA binding to Skp. The spectrum is annotated with yellow circles (BamA), red circles (Skp), green squares (1:1 Skp:BamA) and blue triangles (2:1 Skp:BamA). The most abundant charge state in each distribution is labelled. (b) IMS driftscope plot corresponding to the mass spectrum in (a). Charge states corresponding to Skp, BamA, 1:1 Skp:BamA, and 2:1 Skp:BamA are labelled in red, yellow, green and blue, respectively. The ions at *m/z* ~2000 arise from Skp subunits.



Supplementary Figure 5

Chemical cross-linking and SDS-PAGE/western blotting/MS analysis of Skp:OMP complexes.

(a) SDS-PAGE analysis of a mixture of Skp and OmpA incubated without (-) or with (+) cross-linker. Upon addition of cross-linker a new band appears at approx. 85 kDa, consistent with a cross-linked Skp:OmpA complex (Skp:OmpA: 89.3 kDa, Skp: 54.0 kDa, OmpA: 35.3 kDa). An additional band (triangle) may correspond to (Skp:OmpA)₂ (178.6 kDa), likely due to dimerization via the periplasmic domain as previously observed (Marcoux, J. et al. *Structure* 22 781-790, 2014). (b) Western blotting analysis with an anti-His antibody to confirm the location of His-tagged Skp-containing bands in the cross-linked Skp:OmpA sample. Higher molecular weight bands in the cross-linked sample (> 250 kDa) are therefore likely due to intermolecular OmpA cross-links. (c-d) Peptides identified from excised and digested bands from SDS-PAGE gels of cross-linked Skp:OmpA complexes consistent with (c) a 1:1 Skp:OmpA complex (the molecular weight band at ~90 kDa in a,b), (d) a larger Skp:OmpA complex (the molecular weight band labelled with a triangle in a,b). (e) SDS-PAGE analysis of a mixture of Skp and BamA incubated without (-) or with (+) cross-linker. Note that upon addition of cross-linker, no band appears at the expected mobility for a 1:1 Skp:BamA complex (*, 142.5 kDa). Instead, a single high molecular weight band is observed which corresponds to a higher order Skp:OMP assembly, consistent with the 2:1 Skp:BamA complex observed by ESI-MS (196.5 kDa). (f) Western blotting analysis with an anti-His antibody showing the location of His-tagged Skp-containing bands in the cross-linked Sample in (e) (> 250 kDa) are therefore likely due to intermolecular weight band abelled with a triangle in a,b). (d) a larger Skp:OMP assembly, consistent with (c) a 1:1 Skp:BamA complex veight band is observed which corresponds to a higher order Skp:OMP assembly, consistent with the 2:1 Skp:BamA complex observed by ESI-MS (196.5 kDa). (f) Western blotting analysis with an anti-His antibody showing the location of His-tagged Skp-containing bands in the



(a) Theoretical CCS, and (b) backbone RMSD (red line) and radius of gyration (R_g) (light blue line) calculated for the initial 10 ns of a 100 ns molecular dynamics simulation in the gas-phase. (c) The starting model Skp structure used for the MD simulation (PDB: 1U2M (Walton, T.A. & Sousa, M.C. *Mol Cell* **15** 367-74, 2004), with missing residues in chains B and C modelled from chain A) (Supplementary Data Set 1). Skp subunits are colored green, blue and yellow. (d,e) Structures of Skp after a simulation time of (d) 0.2 ns and (e) 10 ns (Supplementary Data Set 2). The CCS values of the collapsed Skp structures after a simulation time of 10 ns and 100 ns (38.0 ± 1.8 nm² and 37.3 ± 1.9 nm², respectively) (see also Supplementary Table 5) agree favorably with the modal CCS of Skp at the lowest observed charge state (37.9 ± 0.6 nm²) in native-MS experiments (Fig. 4a).



Molecular dynamics simulations of Skp in solvent.

(a) Starting structure used for explicit solvent MD simulation of Skp (PDB: 1U2M3, with missing residues in chains B and C modelled from chain A), shown from the side (left) and bottom (right) (**Supplementary Data Set 1**). (b) Example structure of Skp in an 'open' conformation (t = 7.5 ns), shown from the side (left) and bottom (right) (**Supplementary Data Set 3**). (c) Radius of gyration of Skp over the course a MD simulation in explicit solvent. Structural collapse of initially extended chains of (d) tOmpA and (e) tBamA simulated with an implicit solvent model (see Online Methods).