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Supporting information for
Peroxisome protein import: A complex journey.

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Materials and Methods

Recombinant Protein Preparation:

PEX5C was prepared as previously described [1].

PEX5(1-304) (termed PEX5N) was prepared from the full-length PEX5 expression vector pET28b-*At*PEX5 [1] *via* restriction digest with *Hind*III to introduce four cuts in the PEX5 DNA. The largest fragment, corresponding to pET28b-PEX5(bp1-1159), was isolated by gel extraction, re-ligated with T4 DNA ligase, transformed into XL-10 Gold and confirmed *via* sequencing. A frame shift brought the C-terminal hexahistidine tag into the reading frame.

A colony of pET28b-PEX5N transformed BL21 STAR™ (DE3)/pRARE2 was inoculated into LB (5 mL) and incubated at 37 °C, 220 rpm for 16 h. This culture was diluted 1:500 into LB media (1 L) and incubated at 37 °C, 220 rpm until an OD₆₀₀ 0.4-0.6 was reached, then induced with IPTG (1 mM) and incubated for a further 20 h. Cell pellets were obtained *via* centrifugation (4,000 g, 5 min, 4 °C) from 1 L of culture. The cell pellet was re-suspended in 25 mL chilled lysis buffer (NaH₂PO₄ (50 mM), NaCl (300 mM), Glycerol (15 % v/v), β-metacapethanol (10 mM), pH 8.0) supplemented with 1 × Complete EDTA-free protease inhibitors (Roche) and lysed *via* sonication using a Soniprep 150 with a 10 mm diameter probe (8 Hz, 30 s on/off cycles, 10 min). The lysate was then cleared *via* centrifugation (20,000 g, 30 min, 4 °C). A slurry of Ni-NTA resin (1 mL) (Qiagen) in 50% EtOH was loaded into a column (ca. 0.5 mL Ni-NTA agarose beads) and washed with water (5 mL) and lysis buffer (3 × 5 mL). The resin was re-suspended in the supernatant fraction and incubated for 1 h at 4 °C with constant agitation. The mixture was loaded into a column and the supernatant allowed to flow through. The resin was washed with lysis buffer containing 50 mM imidazole (3 × 5 mL). His₆-PEX5N-His₆ was eluted in lysis buffer containing 250 mM imidazole (6 × 0.5 mL).

Co-Immunoprecipitation:

PEX5 immune serum was raised against amino acids ²³¹K-⁴⁵⁰D of *A. thaliana* PEX5 in rabbit, and was a kind gift from Dr Makoto Hayashi [2].

PEX5N and PEX5C (3 μM final concentration) were combined on ice in a final volume of 250 μL in PBS. The mixtures were incubated with anti-PEX5 N-terminal antibody (1 μL) with gentle agitation at 4 °C for 1 h. Protein A coupled beads (25 μL) (Sigma) were added and the mixture gently agitated at 4 °C for 1 h. Beads were separated by centrifugation (1000 g, 2 min, 4 °C), washed with PBS (3 × 1 mL), and bound proteins eluted from the resin in 2 ×

SDS PAGE sample buffer (1 × 25 μL). PEX5 constructs were detected by anti-polyhistidine immunoblotting as previously described [1].

		0	10
MGSSHHHHHH	SSGLVPRGSH	MAMRDLVNGG	AACAVPGSSS
20	30	40	50
SSNPLGALTN	ALLGSSSKTQ	ERLKEIPNAN	RSGPRPQFYS
60	70	80	90
EDQQIRSLPG	SELDQPLLQP	GAQGSEFFRG	FRSVDQNGLG
100	110	120	130
AAWDEVQQGG	PMPPTGPMFE	PVQPTFEGPP	QRVLSNFLHS
140	150	160	170
FVESSRGGIP	FRPAPVPVLG	LSQSDKQCIR	DRSSIMARHF
180	190	200	210
FADRGEFIN	SQVNALLSSL	DIDDGIQARG	HVPGRFRELD
220	230	240	250
DYWNESQAVV	KPNLHPADNW	AAEFNQHGMD	HGGPDSWVQS
260	270	280	290
FEQQHGVNGW	ATEFEQGQSQ	LMSSQMRSM	MQNIAAMEQT
300			
RKLAAALEHH	HHHH		

Figure 1. Amino acid sequence of His₆-PEX5N-His₆.

Domains are indicated as: hexahistidine (green), PEX5 sequence (bold), WX₃F/Y PEX14 binding motifs (highlight blue).

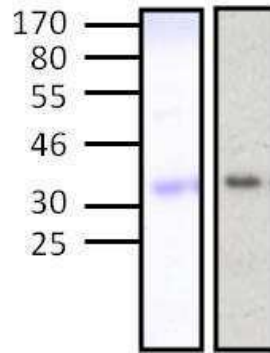


Figure 2. Analysis of purified PEX5N.

Left panel Coomassie stain, right panel α -polyhistidine immunoblot. His₆-PEX5N-His₆ (MW 36.7 kDa) is the band at ~35 kDa. Ladder denotes MW in kDa.

References:

- 1 Lanyon-Hogg, T., Hooper, J., Gunn, S., Warriner, S. L. and Baker, A. (2014) PEX14 binding to Arabidopsis PEX5 has differential effects on PTS1 and PTS2 cargo occupancy of the receptor. *FEBS Lett.* **588**, 2223-2229
- 2 Nito, K., Hayashi, M. and Nishimura, M. (2002) Direct interaction and determination of binding domains among peroxisomal import factors in *Arabidopsis thaliana*. *Plant Cell Physiol.* **43**, 355-366