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PNT1 is a C11 Cysteine Peptidase Essential for Replication of the Trypanosome Kinetoplast

Jaspreet S. Grewal1,2, Karen McLuskey1, Debanu Das3,4, Elmarie Myburgh1,2, Jonathan Wilkes1, Elaine Brown1,2, Leandro Lemgruber4, Matthew K. Gould5, Richard J. Burchmore1, Graham H. Coombs6, Achim Schnaufer5, Jeremy C. Mottram1,2

1Wellcome Trust Centre for Molecular Parasitology, Institute of Infection, Immunity and Inflammation, College of Medical, Veterinary and Life Sciences, University of Glasgow, Glasgow, G12 8TA, UK
2Centre for Immunology and Infection, Department of Biology, University of York, Wentworth Way, Heslington, York, YO10 5DD, UK
3Joint Center for Structural Genomics, http://www.jcsg.org
4Stanford Synchrotron Radiation Lightsourc, SLAC National Accelerator Laboratory, Menlo Park, CA94025, USA
5Institute of Immunology and Infection Research and Centre for Immunity, Infection, and Evolution, University of Edinburgh, Edinburgh EH9 3FL, UK
6Strathclyde Institute of Pharmacy and Biomedical Sciences, University of Strathclyde, Glasgow, G4 0RE, UK

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Correspondence: jeremy.mottram@york.ac.uk

ABSTRACT

The structure of a C11 peptidase PmC11 from the gut bacterium, *Parabacteroides merdae*, has recently been determined, enabling the identification and characterization of a C11 orthologue, PNT1, in the parasitic protozoon *Trypanosoma brucei*. A phylogenetic analysis identified PmC11 orthologues in bacteria, archaea, Chromerids, Coccidia and Kinetoplastida, the latter being the most divergent. A primary sequence alignment of PNT1 with clostripain and PmC11 revealed the position of the characteristic His-Cys catalytic dyad (H99 and C136), and an Asp (D134) in the potential S1 binding site. Immunofluorescence and cryo-electron microscopy revealed that PNT1 localizes to the kinetoplast, an organelle containing the parasite’s mitochondrial genome (kDNA), with an accumulation of the protein at or near the antipodal sites. Depletion of PNT1 by RNAi in the *T. brucei* bloodstream form was lethal both in *in vitro* culture and *in vivo* in mice and the induced population accumulated cells lacking a kinetoplast. In contrast, overexpression of PNT1 led to cells having mislocated kinetoplasts. RNAi depletion of PNT1 in a kDNA independent cell line resulted in kinetoplast loss but was viable, indicating that PNT1 is required exclusively for kinetoplast maintenance. Expression of a recoded wild type PNT1 allele, but not of an active site mutant restored parasite viability after induction *in vitro* and *in vivo* confirming that the peptidase activity of PNT1 is essential for parasite survival. These data provide evidence that PNT1 is a cysteine peptidase that is required exclusively for maintenance of the trypanosome kinetoplast.

INTRODUCTION

*Trypanosoma brucei* is a kinetoplastid protozoan parasite and the causative agent of human African trypanosomiasis (sleeping sickness) and nagana in cattle. Sleeping sickness causes widespread human morbidity and death in sub-Saharan Africa. Nagana leads to cattle mortality, lower meat and milk production and lower calving rate. The parasite has a complex life cycle spanning both the tsetse fly and the mammalian host. The metacyclic trypomastigote is transmitted to the mammalian host by the bite...
of the tsetse fly. Once inside the human host, these parasites transform into bloodstream form (BSF) trypanastigotes that divide and multiply in blood and lymph, and which is followed by invasion of the parasites to other organs and the central nervous system. The sleep-wake cycle gets disrupted and if the disease is left untreated, the infected individual enters coma and eventually dies (1). Approximately 60 million people are at risk of being infected worldwide with this disease (WHO fact sheet May 2015). Currently no vaccines are available and the drugs in use are becoming ineffective and are toxic (2,3). It is therefore imperative that new drug targets are identified against the protozoan parasite.

Cysteine peptidases of parasitic protozoa are associated with important biological processes such as, invasion of the host cells (in case of intracellular parasites) and subsequent pathogenesis (4,5). Clan CD is comprised of peptidase families that have a protein fold similar to the caspase family (C14). Clan CD is exemplified by several important cysteine peptidases such as GPI8 (family C13), a component of the GPI:proteintransamidase peptidases such as GPI8 (family C13), a component of the GPI:proteintransamidase complex (6); metacaspase (family C14B) (7,8); separase (family C50) (9) and a relatively less characterized family C11 (clostripain) (10). In T. brucei the GPI:proteintranisamidase is required for anchoring proteins to the plasma membrane. Among these proteins the most prominent is VSG (variant surface glycoprotein), which forms a monolayer on the parasites surface and functions in the evasion of the host’s immune system (6). T. brucei has five metacaspases, including MCA4, which is a pseudopeptidase and virulence factor (11) and MCA2, which is a calcium-dependent enzyme associated with RAB11 positive endosomes and does not require processing for activation (12,13). Another T. brucei cysteine peptidase, separase, functions in segregation of mini-chromosomes and proper mitotic assembly (14).

Recently, the first crystal structure of a family C11 peptidase, PmC11, was determined from the gut bacterium Parabacteroides merdae (15). This structure facilitated the identification of an important T. brucei protein, PNT1 (Puf Nine Target 1) as a C11 orthologue and a potential cysteine peptidase. The PNT1 transcript was previously described, in the insect procyclic form of the parasite, as the target of the RNA binding protein PUF9 (16), with PNT1 localizing to the kinetoplast, a unique organelle containing the organism’s mitochondrial DNA. The kinetoplast DNA (kDNA) of T. brucei is composed of a few dozen maxicircles (23kb) and several thousand minicircles (~1kb) (17). The maxicircles encode essential mitochondrial proteins including the respiratory chain complex subunits. The minicircles encode guide-RNAs that function in editing the RNA encoded by the maxicircles. The division of the mitochondrial DNA is coordinated with cytokinesis (18) and the presence of essential genes on the kinetoplast makes it imperative for each of the T. brucei progeny cells to inherit a kinetoplast (19). Related species, T. equiperdum and T. evansi, do not have a fully functional kinetoplast and are, therefore, known as dyskinetoplastic (20). RNAi studies on kinetoplast regulatory genes have shown that loss of the kinetoplast in T. brucei is lethal; hence, the organelle is considered an important drug target (19,21). Our functional analysis suggests that PNT1 is a peptidase that plays an essential role in the maintenance of the kinetoplast, and that a catalytically active PNT1 is required for the survival of T. brucei. This work also demonstrates that depletion of PNT1 leads to the generation of an akinetoplastic cell line, whereas overexpression leads to formation of mislocated kinetoplasts thus, inhibitors of PNT1 could be of interest for drug discovery programmes.

**EXPERIMENTAL PROCEDURES**

*Phylogenetic analysis*—The peptidase C11 hidden Markov model (HMM) profile (ACC PF03415.9) was obtained from Pfam (www.pfam.org). The C11 peptidases are members of clan CD (www.merops.org), a group characterised by a doublet [H(G|A)] and triplet [D(A|S)C] motif accounting for the catalytic histidine/cysteine dyad (22). The UniProtKB FASTA set (04/29/2015) was downloaded from the UniProt site (www.uniprot.org). This sequence set was searched for peptides containing domains consistent with the model (conformant proteins) by use of the HMM search program of HMMER3 (http://hmmer.org/) using default parameters. The hits (full sequence E-value <0.0001) were filtered as follows: Sequences from prokaryotic and archaeal species were excluded; eukaryotic alignments were examined and only those with at least a minimum alignment to the cysteine containing...
motif [D(AIS)C] retained. Full peptide sequences for the identified eukaryotic sequences were subsequently retrieved from UniprotKB; and a representative sample of bacterial and archaeal C11 peptidase sequences were obtained from the MEROPS database (www.merops.org) and added to the sequence set.

The meta-method for assembling multiple sequence alignments was performed on the sequence set using the M-Coffee (23) mode of T-Coffee (24). The output alignment (color coded for alignment quality) was examined visually to determine the domain containing the doublet [H(G|A)] and triplet [D(AIS)C]. The alignment was trimmed to this domain, and an HMM profile generated (C11_euk), using the HMMER3 program hmmbuild. This HMM profile was used to search proteome sets of a variety of eukaryotic genomes for C11_euk conformant sequences. The domain sequences for the metacaspase TbMCA2 (Trypanosoma brucei Tb927.6.940, MEROPS family C14, also in clan CD) was subsequently retrieved from UniprotKB; and a MEROPS database (www.merops.org) and C11 peptidase sequences were obtained from the PNT1 RNAi cell line using gene replacement constructs (Matt Gould and Achim Schnaufer, unpublished). Briefly, two rounds of transfections were carried out to replace both the endogenous subunit γ alleles with γL262P mutant alleles. The cell lines were tested for kDNA independence by treating with 10 nM ethidium bromide for four passages, which resulted in generation of viable akinetoplastic cells.

Transfection of the parasites—1 x 10^7 log phase BSF cells were suspended in 100 µL of ice-cold Amaxa Human T-Cell solution (Amaxa, Koeln, Germany) and transfection carried out according to the manufacturer’s protocol. Briefly, 10 µg of linearized DNA plasmid was added to the cells and transected with the Amaxa Human Nucleofector II, using program X-001. The cells were transferred to pre-warmed HMI 11 medium and plated out in 24-well plates at 1:20 and 1:40 dilutions. The selection antibiotic was added after 6 h of transfection and cells incubated at 37 °C. Transfectants were selected after a week. F_Fo ATPase subunit γL262P mutants (20) were generated in the PNT1 RNAi cell line using gene replacement constructs (Matt Gould and Achim Schnaufer, unpublished). Briefly, two rounds of transfections were carried out to replace both the endogenous subunit γ alleles with γL262P mutant alleles. The cell lines were tested for kDNA independence by treating with 10 nM ethidium bromide for four passages, which resulted in generation of viable akinetoplastic cells.

Analysis of trypanosome growth after induction of RNAi—The PNT1 RNAi cell lines
were seeded in triplicate at 1 × 10^5 cells per well in 24 well plates (HMI 11 hygromycin-phleomycin media) in the presence and absence of tetracycline (1 µg ml^-1). The cell numbers were counted using a haemocytometer every 24 h and cell seeded at 1 × 10^5 cells for the subsequent time point. Infections in mice were carried out by initially inoculating a donor mouse with 1 × 10^5 parasites and monitoring blood parasitemia over 72 h. Four experimental mice were infected with 4 × 10^5 parasites harvested from the donor mouse. Doxycycline was added to the drinking water of two of the mice on the day of inoculation. Parasitaemia in the blood of the mice was calculated daily using a haemocytometer after drawing the blood by tail venepuncture and diluting the blood in 0.83% ammonium chloride. Cumulative growth curves were plotted for the in vivo and in vitro cell counts using Prism6 (GraphPad Software). The experiments were done in triplicate for in vitro and in duplicate for in vivo experiments.

**Real time PCR**—2 × 10^5 BSF cells were grown for 24 h in the presence and absence of tetracycline, harvested and RNA was prepared using the RNeasy kit (Qiagen). First strand cDNA was prepared using 4 µg of RNA for each sample (Invitrogen Superscript: First strand synthesis kit). The cDNA was treated with RNase H (Invitrogen) to degrade the template RNA. PCR was performed using SYBR green PCR master mix (Applied Biosystems) and GPI8 was used as an endogenous control. Oligonucleotides OL4119: CACTTGGGTGGGCTCTCTTTC and OL4120:CAGCGGCTTCGACAATGG were used for amplifying PNT1, and OL4118:AGCGCGTGATGACAGTGAAG to OL4122:CGAAGCGCATTTGGATAGC and used for amplifying PNT1, and OL4119: CACTTGGGTGGGCTCTCTTTC and OL4120:CAGCGGCTTCGACAATGG were used for amplifying PNT1, and OL4118:AGCGCGTGATGACAGTGAAG to OL4122:CGAAGCGCATTTGGATAGC and used for amplifying PNT1.

**SDS-PAGE and Western blotting of T. brucei cell lysates**—1 × 10^7 BSF parasites were lysed in sample buffer (NuPAGE sample buffer) and analysed on a 4-12% NuPAGE Novex Bis-Tris gel using MES buffer. Subsequently, the samples were electro blotted on nitrocellulose membrane (GE healthcare) and probed with the respective antibodies. The PNT1HA cell line was probed with anti-HA antibody (1:200, Roche). The PNT1Myc cell line was probed with anti-Myc antibody (1:2000, Millipore). Anti-EF1-alpha antibody (1:5000, Millipore) was used for the loading controls.

**Immunofluorescence**—DAPI staining was carried out using 10^6 cells from exponential phase cultures. The cells were washed with vPBS (22,28), added to a 12-well glass slide (Thermo Fisher) pre-coated in poly-L-lysine (0.1% w/v in H2O, Sigma), and allowed to adhere for 4 min. Cells were fixed with of 4% para-formaldehyde (PFA) for 5 min. The wells were washed twice with PBS (pH 7.4) and excess PFA was quenched with 100 mM glycine for 10 min. The cells were permeabilized with 0.1% Triton X-100 for 10 min. The wells were washed three times with PBS and overlaid with a drop of SlowFade Gold Antifade Reagent with DAPI (Invitrogen). Coverslips were added and the slides were kept at 4°C until required. Staining of PNT1HA was performed using 1 x 10^5 cells from an exponential phase culture. The cells were resuspended in 200 µl PBS followed by addition of 300 µL of 4% PFA to fix overnight at 4°C. The cells were washed with PBS and resuspended in 100 mM glycine for 15 min. The cells were then centrifuged at 600 g for 10 min and resuspended in 0.1% Triton X-100 (Sigma) for 5 min. Cells were washed twice with PBS and incubated on ice for 1 h in 1% BSA (in PBS). The cells were centrifuged for 600 g for 10 min and incubated with anti-HA antibody (1:200, Rat monoclonal, Roche) for 1 h on ice. After washing with 0.1% BSA (PBS) cells were resuspended in anti-Rat Alexa-488 (1:2000, Molecular Probes) and incubated in the dark on ice for 1 h. The cells were washed with 0.1% BSA (PBS) before 0.5 µl DAPI (100 µg/ml stock) was added to the cells and incubated for 5 min. The cells were washed once with 0.1% BSA (PBS) and once with PBS. The final cell pellet was resuspended in final volume of PBS (30 – 50 µL) and stored at 4°C until ready to image. The staining of the PNT1Myc cell line was also carried out as described above using an anti Myc antibody conjugated to Alexa Fluor 488 (Millipore) at 1:500 dilution. Stained cells were spread on a slide and overlaid with a square coverslip. The images were acquired on a DeltaVision Core microscope using excitation wavelength of 405-489 nm and images were analysed and processed in Adobe Photoshop and FIJI.

**Transmission electron microscopy**—Cells were washed in PBS, fixed with 2.5% glutaraldehyde and 4% paraformaldehyde in 0.1 M sodium cacodylate buffer (pH 7.2), and post-fixed with 1% osmium tetroxide and 2.5% potassium ferrocyanide in 0.1 M sodium cacodylate buffer (pH 7.3) for 45 min in the dark.
Samples were en bloc stained with 2% aqueous uranyl acetate, dehydrated in a series of acetone solutions (30, 50, 70, 90 and 100%) and finally embedded in Epon resin. Collected ultrathin sections were stained with lead citrate prior to observation in a Tecnai T20 transmission electron microscope (FEI, Netherlands). All images were analysed and processed in Adobe Photoshop and FIJI (30). Acquired images were processed and the intensity plot profile acquired for kinetoplast density analysis. Statistical analyses were performed using Prism6, with one-way ANOVA analysis. Immuno-gold labelling of cells was done by fixing cells overnight in 4% paraformaldehyde in phosphate buffer, and dehydrating in ascending ethanol solutions. The material was progressively infiltrated with LR White resin, and polymerization was carried out in gelatine capsules under ultraviolet light. Formvar-coated nickel grids with ultrathin sections were incubated with blocking solution (3% BSA, 0.02% Tween 20, in phosphate buffer, pH 8) for 1 h. The grids were then incubated with the primary antibody anti-HA (1:200 rat monoclonal, Roche) diluted in blocking buffer. The grids were then incubated with protein A conjugated to immunogold (Aurion, Netherlands) for 1 h. After several washes with phosphate buffer, the grids were stained with uranyl acetate and lead citrate prior to observation in a Tecnai T20 transmission electron microscope (FEI, Netherlands).

RESULTS

Phylogeny of the C11 cysteine peptidase family—To determine the phylogenetic distribution of C11 proteins, the UniProt database was queried with a bespoke C11 Hidden Markov Model (HMM) domain profile. This analysis identified clostripain-like proteins in a wide range of bacterial and archaeal species and in a limited range of eukaryotic species, which could be divided into distinct clades (branches) (Fig 1A). One clade, containing a subset of species from Chromerida, Coccidia and Chlorophyta, is monophyletic with the archaeal and bacterial species, as exemplified by *P. merdae* PmC11 (15) and *Clostridium histolyticum* clostripain, and suggests a lateral gene transfer event from a non-eukaryotic species to a plastid-containing organism ancestral to these phyla (31). The sparse distribution of the C11 domain is indicative of frequent loss from genomes in successive rounds of evolution, as there is no evidence of C11 proteins among species of Babesia or Plasmodium, whereas C11 proteins are found in other apicomplexan genera including Toxoplasma, Eimeria and Neospora. A second clade (kin) is present in kinetoplastids and is atypical in comparison to other eukaryotic species and bacterial/archaeal domains. The C11 domain is found in *T. brucei* (PNT1), Leishmania and Bodo saltans, but not in *Phytomonas, Leptomonas* or *Paratrypanosoma*, indicating secondary loss of the PNT1 gene in kinetoplastid groups.

Active PNT1 is essential for *Trypanosoma brucei* viability—PNT1 is the ortholog of PmC11 in *T. brucei* (Fig. 1A). A primary sequence alignment of PmC11, clostripain and PNT1 (Fig. 1B) led to the identification of the conserved His-Cys catalytic dyad (H99 and C136, respectively). PmC11 is somewhat shorter than the other two proteins but large sections of all three overlay well, covering most of the PmC11 residues with particularly good alignment around the active site. This suggests that the secondary structural elements identified in PmC11 will form the core structural elements of clostripain and PNT1.

Attempts to obtain soluble recombinant PNT1, for testing protease activity *in vitro*, were unsuccessful. However, it was possible to investigate peptidase activity and its importance in parasite survival indirectly. Initially, a PNT1 RNAi line was generated in the bloodstream-form (BSF) of *T. brucei*, where induction by tetracycline resulted in loss of cell viability in the *in vitro* culture (Fig. 2A). PNT1 was also observed to be essential *in vivo* with doxycycline induction leading to loss of detectable parasitaemia in mice after 96 h (Fig. 2B).

To test whether catalytically active PNT1 is required for the survival of the parasites, a recoded version of PNT1, containing a C-terminal HA tag (PNT1HA, which is refractory to RNAi) was constitutively expressed in the PNT1 RNAi line. Expression of the PNT1HA recoded protein compensated for the loss of endogenous PNT1 and restored viability of tetracycline induced parasites both *in vitro* culture (Fig. 2C) and *in vivo* (Fig. 2D). A cell line in which the recoded PNT1HA had the predicted active site cysteine mutated to serine PNT1HA C136S was not viable after induction *in vitro* (Fig. 2E) and *in vivo* (Fig. 2F). Quantitative RT-PCR confirmed the depletion of PNT1 mRNA in each of the cell lines (insets of Fig’s.
2A, C and E) and Western blotting confirmed that PNT1HA\textsuperscript{C136S} was expressed at a similar level to PNT1HA (Fig. 3A). Another interesting observation from the Western blots was the absence of processed fragments of recoded PNT1HA suggesting that PNT1 might not require processing for activity. This is in contrast to PmC11, which requires intramolecular processing for full activation (15). This observation is further supported by the primary sequence alignment of PNT1 with PmC11 and clostripain, where the cleavage site Lys (K147) in PmC11 aligns with a reported cleavage site Arg (R190) in clostripain (32), whereas in PNT1 there is an Ala at the equivalent position (Fig. 1B). Additionally, phylogenetic analysis (Fig. 1A) shows that trypanosomatid C11 orthologues are quite divergent from bacterial and archaea C11 family members. These results suggest that PNT1 is different from PmC11 in terms of activation and that the peptidase activity of PNT1 is essential for viability of \textit{T. brucei}.

\textit{PNT1} localizes on the kinetoplast—\textit{PNT1} localizes to the kinetoplast in BSF parasites and was enriched either within or near the antipodal sites, which are protein assemblies located on opposite sides of the kDNA disc involved in minicircle replication (33, 34). This localization was more prominent in dividing kinetoplasts (Fig. 3B). Immunogold-labelling of the PNT1HA cell line confirmed that PNT1HA is located on the kinetoplast (Fig. 3C (i, ii)). The status of the kinetoplast in PNT1-depleted cells was investigated using TEM, which showed a lower electron density of the kinetoplast in RNAi-induced cells compared to uninduced cells (Fig's. 3(iii, iv). This indicates that PNT1 depletion results in a reduction of material within the kinetoplast. It has been demonstrated that RNAi-mediated depletion of proteins closely associated with the kinetoplast, can cause the loss of kDNA (35, 36). In order to observe the effects of downregulating PNT1 on number of nuclei and kinetoplasts, the \textit{PNT1} RNAi line was stained with DAPI at different time points after induction. Depletion of PNT1 by RNAi resulted in increased number of cells lacking a kinetoplast (akinetoplastic, 1N0K) after 2 days (Fig. 3D). The number of 1N0K cells increased from 16% on day 2 to 46% on day 4, after RNAi induction. This increase in akiinetoplastic cells suggests that PNT1 is involved in the maintenance of the kinetoplast in \textit{T. brucei}. Interestingly, inducible overexpression of a Myc-tagged PNT1 (PNT1Myc) leads to formation of an additional kinetoplast anterior to the nucleus, a phenotype similar to that observed in the procyclic form of \textit{T. brucei} (16). In addition, 36% of the induced PNT1Myc cell line had a mislocated kinetoplast after 48h of induction (Fig. 4A, B). PNT1 was also localized on these incorrectly positioned kinetoplasts (Fig. 4B (lower panel)).

Overexpression of PNT1 led to an increase of 1N2K cells from 12% to 39% after 24 h of induction, compared to 12% observed in uninduced cells (Fig. 4C). However, the number of 1N2K cells in the induced culture decreased to 14% and 8% at 48 h and 72 h time points, respectively. The \textit{T. brucei} cell cycle is a well-co-ordinated event in which the kinetoplast is placed posterior to the nucleus during cytokinesis. In cells overexpressing PNT1, mislocated kinetoplasts found anterior to the nucleus were observed in cells at all stages of the cell cycle. The decrease in the number of kinetoplasts might be due to continuous cytokinesis of cells having mislocated kinetoplasts. PNT1 overexpression by tetracycline induction also leads to loss of cell viability in BSF \textit{T. brucei} (Fig. 4D). Importantly, cell death is observed when PNT1 is either downregulated or overexpressed, indicating that an optimal amount of PNT1 peptidase activity is essential for viability of the parasite.

\textit{PNT1} is required exclusively for maintenance of the kinetoplast—A kinetoplast-independent \textit{PNT1} RNAi cell line was generated to investigate whether PNT1 is required exclusively for kinetoplast maintenance or whether it has other essential mitochondrial functions. It has recently been demonstrated that an L262P mutation in the \textit{F1}\gamma subunit of the \textit{F1}-\textit{F0} ATPase complex allows BSF \textit{T. brucei} to survive loss of the kinetoplast (20). The \textit{PNT1} RNAi line was modified to replace both of the wild-type \textit{\gamma} alleles with L262P mutant alleles. The \textit{PNT1} RNAi was then induced with tetracycline. No growth defect was observed after induction (Fig. 5A, left panel) despite greater than 90% depletion of the \textit{PNT1} transcript (Fig. 5A, right panel). Additionally, depletion of PNT1 resulted in a 100% akiinetoplastic cell population after 4 days of induction. Thus, PNT1 is redundant in the absence of a requirement for the kinetoplast for cell viability (Fig. 5B). Taken together, these data indicate that PNT1 is exclusively required for kinetoplast maintenance and becomes
redundant in the absence of a cellular requirement for the organelle.

**DISCUSSION**

The structure of a bacterial C11 protein (PmC11) (15) allowed the catalytic dyad of the PNT1 orthologue to be identified and probed. Despite being larger than PmC11, a basic primary sequence alignment suggests that the secondary structural elements identified in PmC11 will form the core structural elements of PNT1 (Fig 1B). Our data and others (16) indicate that PNT1 is present at the ends of the kinetoplast, which could be the antipodal sites where newly replicated minicircles reattach to the network for repairing discontinuities during replication (such as gap filling and ligation) (37,38). Several proteins have been identified at the antipodal sites of the kinetoplast including helicases (33,39), a minicircle origin binding protein p38 (40), DNA polymerase β (41), and a DNA primase PRI1 (42), among others. Depletion of p38 (40), PIF8 (33) and PRI1 (42) also result in loss of kDNA, a similar phenotype to PNT1 depletion; however, there is no evidence to suggest that these proteins require proteolytic processing whereas it is the peptidase activity of PNT1 that is essential. To date, only one other protease, HslVU (43) has been found to regulate kDNA maintenance in *T. brucei*. Unlike PNT1, downregulation of HslVU leads to over-replication of minicircles and formation of large kDNA networks. Moreover, HslVU localizes throughout the mitochondrion, suggesting that HslVU could have other roles in the organelle, whereas PNT1 is required exclusively for kDNA maintenance. The appearance of an additional kinetoplast in an abnormal position observed after PNT1 overexpression is reminiscent of the ‘ancillary’ kinetoplasts reported after ablation of, DNA polymerase IC (44) or the acyl carrier protein (45), and of those observed in candidate gametes of *T. brucei* (46). Phylogenetic analysis of PNT1 orthologues suggests that kinetoplastid PNT1 genes are very divergent from orthologues of other organisms. The sequence alignment of PNT1 with PmC11 revealed the presence of conserved structural elements, but also showed novel elements indicating that they have evolved to perform some kinetoplastid-specific function. Not all kinetoplastids have PNT1: *Phytomonas* and *Leptomonas* appear to lack a PmC11 orthologue, which might indicate that there are alternative regulatory mechanisms for kDNA replication in these organisms. It is also interesting to note that *T. evansi* and *T. equiperdum* have a PNT1 gene, even though these organisms are dyskinetoplastic (47). It is possible that PNT1 is required for maintenance of minicircles in these organisms, although there are no data currently to support this hypothesis. Further work on the identification of PNT1 substrates would provide exciting new insights into the mechanism of kinetoplast replication and segregation in *T. brucei* as well as guiding inhibitor and possibly drug discovery.

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**FIGURE LEGENDS**

**FIGURE 1.** Phylogenetic analysis of the clan CD C11 peptidase family. A. Unrooted neighbour joining tree of the C11 family from bacteria and archaeal species (Bact/Arch), Coccidia (Cocc), Chromerida (Chro), Chlorophyta (Chl) and Kinetoplastids (Kin; A. Trypanosoma, B. Leishmania/Endotrypanum/Crithidia, C. Bodonida). Clostripain (+)Uniprot ID: P09870, PmcC11(*)Uniprot ID: A7A9N3, and PNT1 (#)Uniprot ID:Q385B4 are marked. Key: 1; K2PE90, Trypanosoma cruzi, 2; A0A061IWY5 T. rangeli, 3; F9W8X5 T. congolense, 4; Tb927.11.6550 T. brucei, 5; G0UBL1 T. vivax, 6; Q4QH06 Leishmania major, 7; AOF50010278.1 2 2 2134 Endotrypanum monterogei, 8; CfaC1 13 0800 Crithidia fasciiculata, 9; BS39455 Bodo saltans, 10; EQ973215 Bacteroides fragilis, 11; GL883824 Paraprevotella xylaniphila, 12; AHHG01000029 Elizabethkingia anophelis, 13; P09870 Clostridium histolyticum, 14; ADLW01000010 Dysgonomonas mossii, 15; AE010299 Methanosarcina acetivorans, 16; SNAP00000001341 Eimeria tenella, 17; B6ABL5/Cryptosporidium muris, 18; MER037012 Cryptosporidium parvum, 19; Cve1 15101 Chromera velia, 20; A0A086LCT5 Toxoplasma gondii, 21; V4ZRY6 Toxoplasma gondii, 22; F0VJH3 Neospora caninum, 23; U6G6U0 Eimeria praecox, 24; MER494436 Virella brassicaformis, 25; D8TW66 Volvox carteri, 26; A89P11 Chlamydomonas reinhardtii, 27; A0A0D2MVH2 Monoraphidium neglectum, 28; NC 018015 Thermococcus sp. CL1, 29; ABYK01000024 Arthrospira maxima, 30; DS990529 Acidipulifundum boonei, 31; A7A9N3 Parabacteroides merdae, 32; AJ437303) Trypanosoma brucei MCA2 (outgroup). B. Primary amino acid sequence alignment of PmcC11 (from Parabacteroides merdae), PNT1 (from T. brucei) and clostripain (from Clostridium histolyticum) with identical residues highlighted in grey shading. The assigned secondary structure from the crystal structure of PmC11 (15) is mapped onto its sequence with the position of the PmC11 catalytic dyad and auto-catalytic cleavage site (K147) highlighted by a red star and a red triangle, respectively. Connecting loops are colored gray/black, the main β-sheet is in orange, with other strands in olive, α-helices in blue and the nonapeptide linker of clostripain underlined in red. Sequences around the catalytic site of clostripain, PmcC11, and PNT1 are aligned where identical residues are highlighted in grey.

**FIGURE 2.** Growth of PNT1 RNAi cell lines in vitro and in vivo. A and B, PNT1 RNAi cell line. C and D PNT1 RNAi cell line expressing recoded PNT1HA. E and F, PNT1 RNAi cell line expressing recoded PNT1HA C136S. A, C and E Representative growth curves show cumulative cell counts after induction of RNAi with tetracycline in vitro (mean of three independent experiments) (inset: PNT1 mRNA depletion monitored by qRT-PCR). B, D and F, 4 mice were inoculated intraperitoneally with 4 x 10⁸ parasites and RNAi was induced with doxycycline in 2 mice at the time of inoculation. Uninduced mice were culled when the parasitemia levels were higher than 10⁸ cells/ml. The parasitemia in blood samples were plotted over time.

**FIGURE 3.** Phenotype analysis of PNT1 RNAi, PNT1HA and PNT1Myc cell lines. A, Expression of recoded PNT1HA in the PNT1 RNAi cell line. Lysates of PNT1 RNAi cells expressing recoded PNT1HA (lanes 1 and 2) or recoded PNT1HA C136S (lanes 3 and 4) before (lanes 1 and 3) and after (lanes 2 and 4) tetracycline induction were immunoblotted and probed with anti-HA antibody. EF1α was used as loading control. B, Immunofluorescence of PNT1HA expressed in tetracycline induced PNT1 RNAi cell line (lower panel) shows localization within or near antipodal sites. Uninduced PNT1 RNAi cell lines (upper panel) were used as a control. The cells were stained with anti-HA antibody (green) and DAPI (blue). A stained, nearly duplicated kinetoplast (yellow box) is shown in magnified view. Scale bar; 5µm. C, (i, ii) Immuno gold labelling of PNT1HA cell line using anti-HA antibody after 72 h of RNAi induction. Scale bar; 100 nm. (iii, iv) Electron micrographs of PNT1 RNAi uninduced (iii) and 72 h tetracycline induced (iv) cells (scale bar; 150 nm). D, PNT1 RNAi cells induced with tetracycline (Tet+) were stained with DAPI and the number of nuclei (N) and kinetoplasts (K) per cell was counted (in 200 cells) at the indicated time points and plotted as a percentage of the total. Each histogram represent mean ±SD (n=3).
FIGURE 4. Overexpression of PNT1. A, Upper panel, Western blot showing expression of PNT1Myc using anti-Myc antibody at different time points after tetracycline induction (lanes 2, 3 and 4 represent induced samples after 24, 48 and 72 h respectively). A 24 h uninduced PNT1Myc sample in lane 1 served as negative control. EF1α was used as the loading control. Lower panel, Percentage of cells having anterior kinetoplasts after tetracycline induction at different time points (quantitation from 200 cells, histogram represents mean ± SD (n=3)). B, Representative images showing the DAPI-stained, PNT1Myc-tagged cell line before (top panel) and after (middle panel) induction with tetracycline for 24 h. Bottom panel, Staining of PNT1Myc cell line with anti-Myc antibody conjugated to Alexa fluor 488 (green). Scale bar; 5µm. C, Counts of DAPI stained kinetoplast (K) and nuclei (N) of the PNT1Myc cell line showing a higher percentage of 1N2K cells at 24 h after induction. The number of 1N2K cells fall after 48 and 72 h (n=3, mean±SD). D, cumulative growth curve of PNT1Myc cell line showing reduced cell viability after PNT1 overexpression.

FIGURE 5. PNT1 is exclusively required for kinetoplast function. A, Left panel, cumulative growth curve of PNT1 γL262P RNAi line over time ± tetracycline (n=3). Right panel, the mRNA levels of PNT1 transcript were evaluated in the induced and uninduced γL262P PNT1 RNAi cell line using qRT-PCR. Each histogram represents the mean ± SD (n=3). B, Loss of kinetoplast in γL262P PNT1 RNAi cell line after addition of tetracycline to downregulate PNT1. The cells were grown for 120 h ± tetracycline and stained with DAPI. Scale bar; 5µm.
Figure 1
Figure 2
Figure 3
Figure 4

A. Western blot analysis of Myc and EF1-α levels with molecular weight markers (kDa).

B. Fluorescence microscopy images showing PNT1.Myc (Tet+) cells with DIC and DAPI staining.

C. Bar graph illustrating the percentage of cells having extra kinetoplasts over time (24 h, 48 h, 72 h) for different treatments (1N0K, 2N2K, 1N2K, 1N1K).

D. Graph depicting cumulative cell number over time (0 to 144 h) for Tet- and Tet+ conditions.
Figure 5