Cholinergic Enhancement of Cell Proliferation in the Postnatal Neurogenic Niche of the Mammalian Spinal Cord

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INTRODUCTION

In the mammalian spinal cord, the area surrounding the central canal (CC) exhibits plasticity. First, ependymal cells display neural stem cell features in vitro as they are capable of neurosphere formation, self-renewal and differentiation into the three neural lineages [1, 2]. In vivo, spinal cord ependymal cells can proliferate under both normal and injured conditions and differentiate into astrocytes and oligodendrocytes following injury [3–5] and into neurons in a model of multiple sclerosis [6].

Second, there is a population of cells found either in the subependymal layer or interspersed with ependymal cells which possess a large cerebrospinal fluid-contacting process; thus, they are known as cerebrospinal fluid contacting cells (CSFcs). CSFcs express immunohistochemical markers of immature neurons or neurons involved in plasticity, such as Doublecortin, PSA-NCAM (polysialylated neuronal cell adhesion molecule) and growth-associated protein 43 [7, 8] and display electrophysiological properties consistent with them being neurons at different stages of maturity [7]. Their proliferative capability is less well understood.

Considering the stem cell potential of ependymal cells, understanding physiological factors influencing their function and manipulating

SIGNIFICANCE STATEMENT

Our work has shown that by, activating a specific neurotransmitter receptor, we can transform a population of spinal cord cells, the ependymal cells from being a quiescent stem cell population into one that can proliferate and thus enable spinal cord plasticity. Our findings are therefore of wide scientific interest to those involved in understanding adult neurogenesis and spinal cord function in health and disease. Perhaps most critically, these data will be of interest to those who are seeking to understand how we can harness the body’s natural ability to trigger cell proliferation and differentiation or even to suppress inappropriate proliferative responses.
their proliferation rate could provide an avenue to replenish spinal cord cellular pools that are depleted by injury or disease. Such a physiological factor, endogenous dopamine, promotes the generation of spinal motor neurons in lesioned adult zebrafish spinal cord [9].

Spinal cord regeneration in mammals is less successful and the influence of neurotransmitters is unknown; therefore, we determined whether CSFccs and/or ependymal cells were modulated by neurotransmitters, with a focus on acetylcholine (ACh). Neurotransmitters, including ACh, can regulate proliferation, neuronal differentiation and maturation in established areas of mammalian postnatal neurogenesis such as the subventricular zone of the forebrain and the subgranular zone in the hippocampal dentate gyrus [10]. For example, reducing cholinergic inputs to the olfactory bulb decreases numbers of newly born neurons [11], while their survival was improved by enhancing cholinergic signaling using the centrally acting reversible acetylcholinesterase inhibitor, donepezil [12]. In the dentate gyrus, newborn cells receive cholinergic inputs [13] and the normal survival, maturation, and integration of adult-born neurons is reliant on functional α7-containing nicotinic acetylcholine receptors (α7*nAChRs) [14]. In the subventricular zone, cholinergic neurons have recently been identified that enhance neural stem cell proliferation, thus cholinergic circuits may be critical during numerous stages of postnatal neurogenesis [15]. Cholinergic interneurons reside in the CC region of the spinal cord [16, 17] and thus are well placed to influence cells in the neurogenic niche.

Given the influence of ACh on cells in other postnatal neurogenic niches and the neural stem cell potential of ependymal cells, we hypothesized that these cells are influenced by activation of cholinergic receptors which modulate their proliferative capacity. We use electrophysiology in an in vitro spinal cord slice preparation to demonstrate that both ependymal cells and CSFccs respond to ACh. Furthermore, applications of a cholinergic modulator in vitro to organotypic slice cultures and in vivo combined with 5-ethyl-2'-deoxyuridine (EdU) labeling reveal that cholinergic stimulation enhances the proliferation of ependymal cells, but not CSFccs.

**Materials and Methods**

Full details are given in Supporting Information

**Animals**

Wistar rats (P9-adult) or CS7/B16 mice (P9-adult) of either sex were used in line with the UK Animals (Scientific Procedures) Act 1986 and ethical standards set out by the University of Leeds Ethical Review Committee. Every effort was made to minimize the number of animals used and their suffering.

**Slice Preparation**

Animals (9–28 days) were anaesthetized with sodium pentobarbitone (60 mg/kg) I.P. and perfused transcardially with ice-cold sucrose artificial CSF (aCSF) before cutting 300 μm thick transverse spinal cord slices using a vibrating microtome.

**Whole Cell Patch Clamp Electrophysiology**

Whole cell current clamp recordings were made at room temperature from CSFccs and ependymal cells. Neurobiotin (0.5 %; Vector Laboratories, Peterborough, UK, https://www.vectorlabs.com/uk/) and tetramethylrhodamine (0.02 %; Life Technologies, Paisley, UK, https://www.lifetechnologies.com/uk/en/home.html) were added to visualize the cells post-recording (see ref. 18).

ACh (3–10 mM) was pressure ejected locally; cholinergic antagonists were bath applied at a flow rate of approximately 4–6 ml/minute: these were the muscarinic antagonist atropine (5 μM, Sigma, Gillingham, UK, https://www.sigmaaldrich.com/uk/home.html), the nonselective nAChR antagonist, mecamylamine (MCA; 50 μM), the selective α7*nAChR antagonist at low nanomolar concentrations, methyllycaconitine (MLA; 20 nM), the non-α7*nAChR antagonist at low micromolar concentrations, dihydro-β-erythroidinone (DHβE; 1 μM), and the specific α7*nAChR positive allosteric modulator, PNU 120596 (10 μM; AbCam Biochemicals, Cambridge UK, http://www.abcam.com/). In some experiments, the following drugs were added to the aCSF: tetrodotoxin (TTX; 1 μM; Sigma) to block voltage-gated Na⁺ channels, D(-)-2-amino-5-phosphonopentanoic acid (D-AP5; 50 μM) to block NMDA receptors, and 2,3-dioxo-6-nitro-1,2,3,4-tetrahydrobenzo[f]quinoline-7-sulfonamide (NBQX; 20 μM) to block AMPA receptors. Unless stated, drugs were obtained from Tocris Brustik, UK, http://www.tocris.com/.

**Organotypic Spinal Cord Culture**

CS7/B16 mice (9–14 days) were anaesthetized with sodium pentobarbitone (60 mg/kg; intraperitoneal [I.P.]), perfused transcardially with sucrose aCSF and the thoracolumbar spinal cord was removed and transverse sections cut at 300 μm using a McIlwain tissue chopper. After 48 hours in culture, 1 μM EdU (Invitrogen, Paisley, UK) was added into the medium. For EdU detection in cultures, slices (protected from light) were incubated in 320 μl distilled water, 25 μl 2 M Tris buffer, 5 μl 10 mM copper sulphate, 5 μl Azide⁶⁹⁴, and 100 μl 0.5 M ascorbic acid for 30 minutes, then washed in Tris buffer (2 × 10 min). For EdU detection in sections, the same protocol was followed except 5 μl biotinylated azide (1 mM) was used instead of Azide⁶⁹⁴. This was detected by incubating
sections in Streptavidin Alexafluor 555 (1:1,000 in PBS + 0.1% Triton, Invitrogen) for 1 hour.

**Immunohistochemistry**

Immunofluorescence was performed with antibodies against choline acetyl transferase (ChAT; goat, 1:500; Millipore, Watford, UK, http://www.merckmillipore.com/GB/), cluster of differentiation 24 (CD-24; rat, 1:500; BD, Biosciences, Oxford UK, http://www.bdbiosciences.com/eu/home), polycystic kidney disease 2-like 1 (PKD2L1, rabbit, Abcam, Biochemicals), PanQKI (mouse, UC Davis/NIH Neuromab Facility, Davis, CA, http://neuromab.ucdavis.edu/, N147/6, 1:2), glial fibrillary acidic protein (GFAP, mouse, UC Davis/NIH Neuromab Facility N206A/8 1:100), Na+/K+ ATPase (NKA1; rabbit, Epitomics, Burlingame, CA, http://www.epitomics.com/, 1:1,000), (beta tubulin III (Tuj1, chicken, 1:500 Neuromics, Edina, MN, http://www.neuromics.com/antibodies), Sox2 (goat, 1:1,000 Santa-Cruz), and NeuN (mouse, Millipore, 1:1,000). Antibodies were detected with appropriate Alexa488 conjugated secondary antibodies.

**Image Capture and Manipulation**

Sections were imaged using a Zeiss LSM510 Meta laser scanning confocal microscope equipped with argon (λex = 488 nm) and He-Ne (λex = 543 nm) lasers. Images were captured using Carl Zeiss ZEN software and images adjusted for brightness, contrast, and intensity using CorelDraw16 software. Figures are single plane confocal images.

**Analysis of Data**

For electrophysiology, changes in membrane potential were recorded following ACh application and all data expressed as mean ± SE. Drug effects were determined using paired t-tests. When the CSFcs were separated into different subtypes, one-way analysis of variance (ANOVA) with post hoc Bonferroni tests were used to determine the effects of ACh and two-way ANOVAs with post hoc Bonferroni tests to determine effects of cholinergic modulators on the ACh responses (n = no. of cells).

EdU-positive cells and cells also immunopositive for the specific antibodies were counted in the white and grey matter and the CC region (within 10 μm of the abluminal edge of the ependymal cells) directly through visualization down a microscope (×40 magnification). Cells were mapped, counted, and checked by two investigators. Counts are given as mean number of EdU-positive cells per 300 μm slice in cultured slices or per 50 μm section for in vivo treatment (n = no. of slices or sections, N = no. of animals). All data are expressed as means ± SE and for statistical analysis, one-way ANOVAs with post hoc Bonferroni tests determined differences in the numbers of proliferating cells between control and PNU treated slices and percentages of colocalization.
Figure 2. Acetylcholine has profound depolarizing effects on ependymal cells and cerebrospinal fluid contacting cell (CSFcCs). (A–D): Responses of ependymal cells and CSFcCs to depolarizing and hyperpolarizing current pulses (A and B, ± 100 pA; C and D, ± 10 to −30 pA). Arrows denote synaptic potentials. These responses, together with the morphology of the cells that had been filled with neurobiotin (Ai–Ci) and rhodamine (Di), were used for identification of cell types. Scale bars = 10 μm. (Aii–Dii): Responses of cells to pressure application of acetylcholine (ACh); arrows denote puff applied. (E): Overlay of the responses to ACh in different cells to show the difference in amplitude of the depolarization. (F): Responses to ACh recorded in the presence (tetrodotoxin [TTX]) and absence (control) of TTX. Abbreviations: ACh, acetylcholine; CSFcC, cerebrospinal fluid contacting cell; TTX, tetrodotoxin.
RESULTS

Cholinergic Structures Closely Appose Both Ependymal Cells and CSFcCs

ChAT immunopositive structures were closely apposed to ependymal cells identified as CD24 +ve [19] (Fig. 1A, 1B) and CSFcCs as PKD2L1 +ve [20] (Fig. 1A, 1C), indicating that cholinergic terminals are ideally located to influence both cell types in this CC region.

Ependymal Cells and CSFcCs Are Acutely Depolarized by Activation of Cholinergic Receptors

Ependymal cells and CSFcCs were distinguished by their voltage responses to current injections and the morphology revealed by intracellular dye-loading. Whole cell patch clamp recordings in acute spinal cord slices revealed that ependymal cells had a low input resistance (IR, 96.3 ± 12.6 MΩ, n = 79 cells) and no spontaneous activity (Fig. 2A). Ependymal cells also exhibited gap junction coupling between cells, illustrated by transfer of neurobiotin into neighbouring cells, as expected from previous studies [18]. Dye filling confirmed ependymal cells by a lack of process into the CC (Fig. 2Ai). Ependymal cells were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai). CSFcCs were categorized into three subtypes: subtype 1 cells were defined by a lack of process into the CC (Fig. 2Ai).

CSFcCs were also significantly decreased in MLA from 0.5 mV to 7.3 mV (p < 0.001) and all CSFcCs, regardless of subtype, 2 and 3 (p < 0.001) for subtype 2 CSFcCs and subtype 3 CSFcCs (p < 0.001) was significantly decreased in MLA from 2.9 mV to 0.2 mV (n = 6 animals) than those of both subtype 2 and subtype 3 CSFcCs (p < 0.001; Fig. 2E). The response to ACh was likely to be a direct effect on ependymal cells and CSFcCs as the responses were not significantly decreased in the presence of TTX, AP5, and NBQX (p < 0.05; n = 3–5 cells; Fig. 2F).

Cholinergic Responses of Ependymal Cells and CSFcCs Are Mediated by nAChRs

Atropine (5 μM), the muscarinic selective antagonist, had no significant effect on ACh responses in either cell type (regardless of CSFc subtype; Fig. 3A). The nicotinic-selective antago-
Activation of α7*nAChRs Increases Proliferation of Ependymal Cells In Vivo

To determine whether the α7*nAChR-mediated enhancement of proliferation in slice culture could be extended to the whole animal and could also be observed in adults, the effects of in vivo application of the selective α7*nAChR modulator PNU 120596 on cell proliferation in the CC, grey, and white matter of the spinal cord were tested. Sections were...
analyzed from thoracic and lumbar spinal cord regions and the locations of EdU-positive cells were mapped (Fig. 5).

In the CC area (Fig. 5E), the average numbers of EdU-positive cells per section in control animals (N = 4 animals) injected with saline were 1.0 ± 0.2, (n = 35 sections) in the thoracic region and 1.4 ± 0.3 (n = 40 sections) in the lumbar region. In comparison with control sections, injection of PNU 120596 significantly increased (ρ < 0.0001) the average number of EdU-labeled cells per section in the CC area in both the thoracic (3.0 ± 0.2, n = 40 sections, N = 4 animals) and lumbar (4.6 ± 0.4, n = 40 sections) regions.

In the grey matter (Fig. 5F), not including the CC region already counted, the average numbers of EdU-labeled cells per section in control animals were 11.9 ± 1.2 (n = 35 sections) in thoracic sections and 15.2 ± 0.8 (n = 40 sections) in lumbar sections. Injection of PNU 120596 caused a significant (ρ < 0.0001) increase in the number of EdU-labeled cells in the grey matter of both the thoracic (15.9 ± 1.3 cells per section, n = 40 sections) and lumbar (21.8 ± 1.3 cells per section, n = 40 sections) spinal cord compared with control. In the white matter, (Fig. 5F) control animals had EdU-labeled cells in both the thoracic (30.6 ± 2.0 cells per section, n = 35 sections) and lumbar (37.8 ± 2.7 cell per section, n = 40 sections) regions. Once again, injection of PNU 120596 caused a significant (ρ < 0.0001) increase in EdU-labeled cells in the white matter of thoracic (42.3 ± 2.0 cells per section, n = 40 sections) and lumbar (49.5 ± 1.9 cells per section, n = 40 sections) regions.

Identity of Proliferating Cells in the CC

As both CSFCs and ependymal cells were shown to be acutely depolarized by activation of α7* nAChRs, we next established which type of cells were proliferating in both cultured slices and in vivo. Using antibodies for CD24 or NKAx1 [22] as markers for ependymal cells and PKD2L1 as a marker of CSFCs, dual labeling for EdU was carried out on treated slices. Colocalization of anti-PKD2L1 and EdU labeling was not observed in either condition (n = 13 slices, N = 6 animals for cultured slices; Fig. 4C and n = 15 sections, N = 5 animals for in vivo treatment; Fig. 6A); however, colocalization of anti-CD24 or NKAx1 and EdU labeling was always seen in both cultured slices and after in vivo treatment (n = 13 slices, N = 6 animals for cultured slices; Fig. 4C and n = 16 sections, N = 5 animals for in vivo treatment; Fig. 6B, 6C). Furthermore, EdU in the ependymal layer always colocalized with Sox2, a transcription factor associated with ependymal cells [23], in both the in vivo experiments (Fig. 6D) and cultured spinal cord slices. Taken together, the presence of Sox2 in the nucleus and CD24 and NKAx1 in the membranes surrounding all EdU-positive cells, is consistent with activation of α7* nAChRs selectively initiating proliferation in ependymal cells and not CSFCs.

The identity of those cells in the CC that had proliferated in vivo was further tested using dual labeling techniques after in vivo treatment with PNU 120596. EdU-labeled cells surrounding the CC contained immunoreactivity for the transcription factor Sox2, consistent with a stem cell identity ([24]; Fig. 6D). Furthermore, EdU-labeled cells did not contain immunoreactivity for class III β tubulin (Tuj1) a marker for immature and mature neurons ([25]; Fig. 6E), the reactive glial cell marker GFAP ([26]; Fig. 6F) or the oligodendrocyte marker PanQKI ([27, 28]; Fig. 6G). There were never any neuronal nuclei marker (NeuN)
Figure 5. In vivo administration of the cholinergic modulator PNU 120596 increased the number of 5-ethynyl-2'-deoxyuridine (EdU)-labeled cells in both thoracic and lumbar regions. (A–D): Representative diagrams showing the distribution of EdU-labeled cells in control sections in thoracic (A) and lumbar (B) regions compared with PNU 120596 treated sections at thoracic (C) and lumbar (D) levels. (Ai–Di): Confocal images of the EdU-labeled cells surrounding the central canal (CC) from the representative diagrams above. Scale bars = 20 μm. (E, F): Average number of EdU-labeled cells per 50 μm section ± SE in the area of the CC (E), grey and white matter (F) of thoracic and lumbar spinal cord sections in control and PNU 120596 treated animals. ****, p < 0.0001. Abbreviation: EdU, 5-ethynyl-2'-deoxyuridine.
Figure 6. Following in vivo PNU 120596 administration, 5-ethynyl-2'-deoxyuridine (EdU)-labeled cells surrounding the central canal (CC) are proliferating, yet undifferentiated. (A–D): Confocal images following PNU 120596 application in vivo showing EdU-labeled cells surrounding the CC (Ai) are distinct from the population of PKD2L1 immunoreactive cerebrospinal fluid contacting cells (Aii) as seen in the merged image (Aiii). EdU-labeled cells (Bi, Ci) are colocalized with immunoreactivity for the ependymal cell markers CD24 (Bii) and NKA α1 (Cii) as shown in the merged images (Biii, Ciii). EdU-labeled cells contain immunoreactivity for the transcription factor Sox2 (EdU [Dii], Sox2 [Diii], merged image [Diii]; boxed area shows a higher magnification of a dual-labeled cell). EdU-labeled cells show no colabeling with the neuronal marker Tuj1 (Ei–Eiii), the astrocyte marker glial fibrillary acidic protein (Fi–Fiii) or the oligodendrocyte marker PanQKI (Gi–Giii). Scale bars = 10 μm. Abbreviations: EdU, 5-ethynyl-2-deoxyuridine; GFAP, glial fibrillary acidic protein; NKAα1, Na⁺/K⁺ ATPase α1.
immunopositive neurons (labeling mature neurons) in the vicinity of the CC (not shown). The colocalization of EdU with Sox2, CD24, and NKAx1 normally expressed by ependymal cells, but not GFAP nor PKD2L1 which are expressed by other ependymal layer cell types, is consistent with the EdU positive ependymal layer cells being ependymal cells with stem cell characteristics.

As the newly proliferated cells in the CC region of the intact spinal cord seem to be maintained in a state of readiness as stem cells, we hypothesized that signals to induce differentiation were absent and the α7nAChR activation could be strongly promoting proliferation over differentiation. As preparation of the spinal cord cultures necessarily involves damage to the system again, we hypothesized this may be sufficient to trigger such differentiation. In control conditions, cells in the CC were neither NeuN nor Tuj1 immunoreactive, while the proportions of EdU-labeled cells that were also labeled for GFAP (2.4 ± 1.2 %) or PanQKI (7.3 ± 2.4 %) were quite low. PNU 120596 application resulted not only in an increase in proliferation but also significantly (p = 0.02) increased the proportion of cells that were colabeled for PanQKI to 15.1 ± 2.6 % (Fig. 4F). There were still no cells labeled for EdU and either NeuN or Tuj1 nor was there a significant change in the level of colabeling for EdU and GFAP (4.0 ± 1.0 %). This suggests that presence of an α7nAChR potentiator results in the proliferating cells preferentially differentiating along an oligodendrocyte lineage.

Identity of Proliferating Cells in the Other Spinal Cord Regions

In the grey and white matter of the spinal cord, the numbers of cells colocalized for EdU and either NeuN or GFAP were negligible. These were not changed by the application of PNU 120596. However, there were moderately high percentages of EdU immunoreactive cells that were also colabeled for PanQKI in thoracic (grey matter 24.0% ± 6.7%, white matter 21.8% ± 3.4%) and lumbar (grey matter 12.9% ± 3.3%, white matter 17.4% ± 2.2%) sections. These proportions were significantly (p = 0.0001) increased by PNU 120596 in both thoracic (grey matter 51.1% ± 5.3%, white matter 36.1% ± 4.2%) and lumbar (grey matter 34.6% ± 5.7%, white matter 32.8% ± 3.6%) sections (Fig. 7). This indicates that the newly proliferated cells in the intact white and grey matter of the spinal cord also preferentially express oligodendrocyte markers.

**DISCUSSION**

This study provides the first evidence that cell proliferation in the neurogenic niche of the postnatal mammalian spinal cord is significantly enhanced by the potentiation of endogenous ACh through activation of α7nAChRs in both organotypic slice cultures and in vivo. Although both ependymal cells and CSFcs of the mammalian spinal cord responded robustly to ACh in acute spinal cord slices, our data revealed it was only the ependymal cells that were capable of cholinergic receptor induced proliferation. In addition, α7nAChR stimulation resulted in proliferation of cells in both the white and grey matter. There was a significant increase in the proportion of newly proliferated cells that expressed oligodendrocyte markers.

**Cholinergic Responses of Ependymal Cells and CSFcs**

The combination of immunohistochemistry demonstrating the presence of cholinergic structures closely apposing ependymal cells and CSFcs and electrophysiology demonstrating maintenance of the ACh response in TTX, suggests that the ACh response is mediated by receptors located directly on the membrane of both cell types. The cholinergic structures may be either synaptic terminals or dendrites; in other central nervous system (CNS) regions, both dendrites and axon terminals have the capacity to release acetylcholine [29]. There could be an alternative source of ACh or choline within the CSF [30], which both cell types contact. The variation in the size of the ACh response in different CSF subtypes is likely to correspond to a difference in receptor density or proportions of receptors mediating the response.

Both antagonisms of the cholinergic response by DH/E and potentiation of the response by PNU 120596 are in agreement with in situ hybridization studies, which reveal the expression of α7nAChR [31] and non-α7nAChR subunits including α2–4 and β2 [32]. In addition, an α7nAChR subunit reporter mouse made by the Gensat project (Gensat 2012) showed some GFP expressing CSFcs. As DH/E is a preferential antagonist at α4*αChRs and β2*αChRs [33, 34], it is likely that α4/β2*αChRs could be contributing to the cholinergic response in both ependymal cells and CSFcs, however, this needs further confirmation. The indication of the presence of α7nAChR antagonists was further supported by the antagonism of PNU 120596 potentiation by MLA in both acute and cultured slices.

**Functional Implications of Cholinergic Modulation of Cells in the Neurogenic Niche**

The importance of this cholinergic influence on the CC neurogenic niche is maintained into adulthood since it was observed in both cultured slices obtained from juvenile animals and fully mature mice, using the in vivo preparation. The higher number of proliferating cells in the control cultured spinal cord slices compared with the in vivo situation is likely due to the fact that slice culture is more representative of an injury state in which increased proliferation is observed [4, 5]. This further indicates that cholinergic enhancement of proliferation can occur in physiological and pathological situations. Indeed, the reduction of proliferation by application of only α7nAChR antagonists in culture suggests that these receptors may be necessary and sufficient for injury induced proliferation to occur.

There are a number of functional implications to consider with regards to the effects of the cholinergic modulators on ependymal cells and CSFcs. The fact that PNU 120596 has an effect in cultured slices suggests that the source of ACh is local and maintained in this reduced preparation. An initial consideration is whether the two possible sources of ACh, axodendritic, or CSF, are mediating different functional effects. For example, structures containing ACh closely apposing both cell types could be important for ensuring that newly generated cells are integrated within the spinal cord circuitry, responding to neighboring cells and contributing to specific pathways. This idea is similar to that recently proposed in the subventricular zone where newly identified cholinergic neurons are sufficient to control neurogenic proliferation [15]. In the case of CSFcs, it has previously been hypothesized that these cells act as sensory neurons, sensing the composition of the CSF [35], and this is
supported as they can respond to changes in pH and ATP [7].

This study, however, provides evidence that they are far more complex than simple CSF sensors; they could be modulated by other neurons. The origin of the cholinergic structures influencing either ependymal cells or CSFcCs is currently unknown, although it could be speculated that they are from partition or CC cluster cholinergic interneurons located in lamina X [36]. One possible source is a small cluster of cholinergic interneurons in the CC region that provide c boutons onto motoneurons [16]; their close proximity to the ependymal cell layer would enable release from either dendrites or axon terminals. Given the central location of ependymal cells and CSFcCs within the spinal cord, they are well placed to receive inputs regarding sensory, autonomic and motor functions. Alternatively or indeed additionally, ACh or choline within the CSF [37] could act in a paracrine manner coordinating a widespread effect on

**Figure 7.** Following in vivo PNU 120596 administration, 5-ethynyl-2'-deoxyuridine (EdU)-labeled cells in the white and grey matter preferentially become oligodendrocytes. (A) Average percentage ± SE of 5-ethynyl-2'-deoxyuridine (EdU) and PanQKI colabeled cells per 50 µm section in the grey matter (gm) and white matter (wm) of thoracic and lumbar spinal cord sections in control and PNU 120596-treated animals. *, p < 0.05; **, p < 0.01; ***, p < 0.001. (B, C): Confocal images showing examples of EdU and PanQKI-IR colocalization in grey (Bi–Biii) and white (Ci–Ciii) matter. Scale bars = 20 µm. Abbreviations: EdU, 5-ethynyl-2'-deoxyuridine; PNU, PNU 120596; WM, white matter.
populations of ependymal cells and CSFccs throughout the spinal cord. This is less likely due to the fact that MLA reduced proliferation in the culture, where CSF is washed out.

Both sources of ACh could mediate effects on the proliferation, maturation, and survival of both cell types. With regard to CSFccs, our study did not reveal α7*nAChR mediated proliferation of CSFccs, although these cells were highly responsive to activation of these receptors. It is also possible that cholinergic receptor activation of CSFccs or indeed other intermediary cells could release factors which in turn contribute to the ependymal cell proliferation. However, our evidence from the recordings in acute spinal cord slices suggests that proliferation of ependymal cells could be due to a direct activation of cholinergic receptors on these cells. Cholinergic stimulation of ependymal cells is likely to induce an increase in intracellular Ca2+, as observed in other non-neuronal CNS cells [38, 39], leading to a number of downstream effects. Interestingly, following increases in intracellular Ca2+, the coupled nature of ependymal cells would enable propagation of Ca2+ waves as observed in radial glial cells during embryonic neurogenesis and neural progenitor cells in the postnatal subventricular zone [40, 41]. Ependymal cells of the spinal cord proliferate to maintain the ependymal cell population under normal conditions [2]. This study demonstrates that cholinergic modulation enhances this proliferation and as this system is readily accessed pharmacologically, this provides an avenue for therapeutic intervention.

Implications for New Avenues in Spinal Cord Regeneration

An important finding in the CC was the strong colocalization of EdU with the transcription factor Sox2. Sox2 is considered a marker of neural stem cells since it inhibits neuronal differentiation and maintains progenitor characteristics [24], which is also evident in the spinal cord ependymal cell population [42]. Thus, enhancing the actions of α7*nAChRs leads to increased production of neural stem cells. We did not find evidence of differentiation of the newly proliferated cells in the intact CC region of spinal cord since EdU was not colocalized with markers of neurons or glial cells, but our short recovery period is likely to have precluded this. Alternatively, signals for appropriate differentiation may have been absent in the CC of spinal cords. However, in the cultured spinal cord slices, we found that a higher proportion of the EdU positive CC cells was also colabeled with anti-PanQKI in PNU 120596 treated animals compared with control. This suggests that in this spinal cord slice culture, differentiation along the oligodendrocyte lineage is more prominent when α7*nAChRs are activated. In fact, it is known that mammalian spinal ependymal cells can differentiate in some conditions, indeed they are the only spinal cells capable of generating progeny of multiple fates [5]. In a mouse model of multiple sclerosis, ependymal cells not only proliferated but also migrated and differentiated into neurons [6]. A recent study in zebrafish was the first to demonstrate that a neurotransmitter (dopamine) could promote spinal motor neuron generation in the lesioned spinal cord [9]. Therefore, similar to other CNS regions [43], identification of appropriate signals to direct differentiation and improve survival and integration of endogenously produced neural precursors will be amongst the next steps towards spinal repair.

Another important finding was the increased number of EdU-labeled cells in the white and grey matter of the spinal cord following PNU 120596 administration. These cells also expressed Sox2, indicating the potential to increase stem cell production outside of the CC. Indeed, PNU treatment not only increased EdU labeling, but resulted in an increased proportion of EdU-labeled cells being immunoreactive for PanQKI. While the identity of the cells contributing to the α7*nAChR induced proliferation in the white and grey matter remains to be determined, one potential source could be the meninges [44]. It is clear though that the α7*nAChR could be a viable target to increase the numbers of oligodendrocytes in the spinal cord, perhaps following injury or in demyelinating diseases such as multiple sclerosis.

Summary

In conclusion, this study is the first to demonstrate that neurotransmitter mediated manipulation can enhance spinal cord cell proliferation in mammals. One likely source of these cells is the ependymal cell layer [4, 5], consistent with our observations here. The factors determining the fates of these cells require further elucidation, but our data provide new potential for future therapeutic interventions since the cholinergic system can be manipulated by drugs that cross the blood brain barrier.

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Author Contributions

L.A. and J.D.: design of some experiments, collection and/or assembly of data, data analysis and interpretation, manuscript writing; L.A. and J.D.: collection and/or assembly of data, data analysis and interpretation, manuscript writing; I.J.E. and L.N.: collection and/or assembly of data, data analysis and interpretation; S.A.D.: collection and/or assembly of data, data analysis and interpretation, manuscript writing, final approval of manuscript.

Disclosure of Potential Conflicts of Interest

The authors indicate no potential conflicts of interest.


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