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1 **Title:** Identifying traits for genotypic adaptation using crop models

2

3 **Running title:** Modelling traits for adaptation

4

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20

21 **Type of paper:** Review

22

23 **Significance statement**

24 We review state-of-the art genotypic adaptation modelling and suggest potential avenues to  
25 better realise the potential of model-based studies of genotypic adaptation for guiding  
26 future crop breeding efforts.

27

28 **Abstract**

29 Genotypic adaptation involves the incorporation of novel traits in crop varieties so as to  
30 enhance food productivity and stability and is expected to be one of the most important  
31 adaptation strategies to future climate change. Simulation modelling can provide the basis  
32 for evaluating the biophysical potential of crop traits for genotypic adaptation. This review  
33 focuses on the use of models for assessing the potential benefits of genotypic adaptation as  
34 a response strategy to projected climate change impacts. We first review some key crop  
35 responses to the environment as well as the role of models and model ensembles for  
36 assessing impacts and adaptation. Finally, we describe how crop-climate models can help  
37 focus the development of future-adapted crop germplasm in breeding programs. While  
38 recently published modelling studies have demonstrated the potential of genotypic  
39 adaptation strategies and ideotype design, we argue that for model-based studies of  
40 genotypic adaptation to be used in crop breeding, it is critical that modelled traits are better  
41 grounded in genetic and physiological knowledge. To this aim, two main goals need to be  
42 pursued in future studies: (1) a better understanding of plant processes that limit  
43 productivity under future climate change; and (2) a coupling between genetic and crop  
44 growth models –perhaps at the expense of number of traits analysed. Importantly, the latter  
45 may imply additional complexity [and likely uncertainty] in crop modelling studies. Hence,  
46 appropriately constraining processes and parameters in models and a shift from simply

47 quantifying uncertainty to actually quantifying robustness towards modelling choices are  
48 two key aspects that need to be included into future crop model-based analyses of  
49 genotypic adaptation.

50 Keywords: climate change, impacts, genotypic adaptation, ideotypes, crop models

51

## 52 **1. Introduction**

53 Agriculture is one of the most vulnerable sectors to changes in climates, due to its reliance  
54 on adequate environmental conditions for achieving high productivity (Huntingford *et al.*,  
55 2005). Crops are affected by shortages or excesses of water or excessively high or low  
56 temperatures during key periods of their growing cycle (Porter and Semenov, 2005).

57 Effects from adverse environmental conditions have been largely studied and reported by  
58 several authors, using combinations of models and data (Allen *et al.*, 2005; Boote *et al.*,  
59 2005). This understanding, in addition to well-constrained and skilful simulation models  
60 can provide insights on what could happen under future climate scenarios of higher  
61 temperatures, changing precipitation patterns and increased likelihood of extremes.

62

63 Although figures are varied, recent literature indicates that negative impacts are expected to  
64 affect the basic food basket (i.e. wheat, rice, maize and grain legumes), as well as major  
65 cash crops (i.e. sugarcane, coffee, cocoa) at moderate or low ( $\leq +3$  °C) levels of warming if  
66 no adaptation actions are taken (Lobell *et al.*, 2008; Porter *et al.*, 2014; Challinor *et al.*,  
67 2014b). Evidence from regional and local studies as well as global meta-analyses of  
68 modelling studies indicates that adaptation strategies are critical in countering any negative  
69 and/or capitalising positive effects that may arise as a result of climate change (Claessens *et*  
70 *al.*, 2012; Challinor *et al.*, 2014b). Adaptation strategies are likely the only means by which

71 food availability and stability can be maintained and/or increased so as to meet future food  
72 security needs. In fact, recent model-based global estimates indicate that even incremental  
73 adaptation strategies could result in mean yield increases of ~7 % at any level of warming  
74 (Porter *et al.*, 2014; Challinor *et al.*, 2014b). This suggests that substantial opportunities  
75 may exist if deeper (i.e. systemic and transformational) changes in cropping systems are  
76 implemented.

77

78 This review focuses on one such strategy, namely, genotypic adaptation. Genotypic  
79 adaptation involves the incorporation of novel traits in crop varieties so as to enhance food  
80 productivity and stability and, more broadly, also the design of crop ideotypes (i.e. crop  
81 plants with ideal traits) for future climates (Donald, 1968; Semenov and Stratonovitch,  
82 2013). Specifically, we review the use of models for the development of genotypic  
83 adaptation options. We first examine some important crop responses to key environmental  
84 factors. Secondly, we examine two aspects of climate impacts research: (1) the different  
85 approaches to climate change adaptation, and (2) the importance of models for developing  
86 adaptation options. We then describe existing models and provide recommendations so as  
87 to capitalise on the potential of using crop model ensembles for understanding crop  
88 responses and adaptation options under future climate scenarios. We finally describe how  
89 crop-climate models can help focus the development of future-adapted crop germplasm in  
90 breeding programs. In doing so, we review past experiences and recent trends in the crop  
91 modelling literature. We conclude by proposing a framework that mainstreams crop model-  
92 based analyses into future breeding strategies.

93

94 **2. Key plant processes and crop responses to varying environmental factors**

95 In large areas, climate signals are discernible for many crops and regions even when  
96 aggregated growing season information is used (Fig. 1). Signals in such areas reflect crop  
97 plant responses to variations in weather and climate at local scale. Some of these responses  
98 are discussed in detail below.

99

100 [Figure 1 here]

101

102 A balance exists in the plant-soil-atmosphere interaction so as to allow enough carbon  
103 uptake for plant growth, prevent desiccation due to excess transpiration, and maintain  
104 canopy and leaf temperatures at near-optimum levels (Huntingford *et al.*, 2005; Lobell *et*  
105 *al.*, 2013). Stomatal conductance, a key factor regulating plant growth, is highly correlated  
106 with net photosynthesis (Wong *et al.*, 1979) and is affected by air moisture deficit (i.e.  
107 vapour pressure deficit –VPD), radiation intercepted, leaf temperatures, ambient CO<sub>2</sub>  
108 concentrations, and soil moisture. However, both temperature and air and soil moisture  
109 conditions operate against plant growth and also against each other in ways that are often  
110 difficult to understand.

111

112 Mean air temperatures drive canopy and leaf temperatures, which are determinant for  
113 photosynthesis. Photosynthetic efficiency varies with temperature in all crop species  
114 because it affects RuBisCO (Ribulose 1,5 biphosphate carboxylase oxygenase) activity, and  
115 in turn intercellular CO<sub>2</sub> concentration and stomatal conductance (Hew *et al.*, 1969; El-  
116 Sharkawy, 2014). Response of photosynthesis to temperatures varies by species (Fig. 2A).  
117 Mean temperatures also drive crop development rates and thus define crop duration (Fig.  
118 2B), which in turn affects total photosynthetically active radiation (PAR) intercepted –

119 linearly related to total biomass production. Daily extremes of temperature reduce crop  
120 yield mostly through damage to plant reproductive organs (Fig. 2C) (Peng *et al.*, 2004) and  
121 hastened senescence (Asseng *et al.*, 2011). However, complex responses and interactions  
122 occur throughout the cropping cycle. For an example: under optimal temperatures and  
123 water availability, photosynthesis and transpiration from leaves occur at normal rates;  
124 however, under high temperatures plants open their stomata to avoid heat stress, which  
125 increases within-leaf CO<sub>2</sub> concentrations and thus biomass accumulation (exception being  
126 made under high VPD conditions –dry air, as in such a case stomata would remain closed  
127 to avoid excessive transpiration). If the available soil water is limited, this induces  
128 desiccation and stomata are then closed. Drought causes desiccation and stomatal closure,  
129 but at the same time water is a direct input of photosynthesis and so the effects on carbon  
130 fixation are more direct than those of temperature. In addition, stomatal closure causes  
131 within-leaf CO<sub>2</sub> concentrations to decrease, thus decreasing inputs to photosynthesis, in  
132 some cases also increasing photorespiration (Kobza and Edwards, 1987). This causes lower  
133 biomass production and limits growth (Hew *et al.*, 1969; Huntingford *et al.*, 2005). Low  
134 light incidence (i.e. solar radiation) also reduces photosynthesis, whereas winds increase  
135 transpiration. Drought stress may be induced by increased osmotic pressure in saline soils.  
136 Many limiting conditions can occur simultaneously in a given site [e.g. Trnka *et al.*  
137 (2014)], thus making any prediction of their effect a challenging task.

138

139 [Figure 2 here]

140

141 The effects of increased CO<sub>2</sub> are beneficial for almost any food crop, with increased CO<sub>2</sub>  
142 concentrations thought to increase dry matter and thus yield (Leakey *et al.*, 2009).

143 However, there is contrasting experimental evidence on crop responses to enhanced CO<sub>2</sub>  
144 concentrations across varying degrees of soil water and air moisture availability (Long *et*  
145 *al.*, 2006; Tubiello *et al.*, 2007; Ainsworth *et al.*, 2008), despite advances in theoretical  
146 understanding (Ghannoum *et al.*, 2000; Leakey, 2009). Underlining experimental evidence  
147 on crop responses to elevated CO<sub>2</sub> concentrations is therefore needed, since most models  
148 incorporate effects in a fairly basic fashion –mainly through empirical factors to reduce  
149 assimilation. Particular attention must be placed on understanding the interactions between  
150 enhanced [CO<sub>2</sub>] and other environmental controls (particularly drought and high  
151 temperatures), as these remain only partially understood (White *et al.*, 2011; Asseng *et al.*,  
152 2013).

153

154 A large number of other factors exert control on plant growth and, particularly, on  
155 photosynthesis, biomass accumulation and yield. Leaf nitrogen (N) content is strongly and  
156 positively associated with carbon exchange rates (CER), radiation use efficiency (RUE) and  
157 total plant biomass (Sinclair and Horie, 1989). Similarly, low phosphorous (P) and  
158 potassium (K) contents can also lead to limited CER and biomass production (Longstreth  
159 and Nobel, 1980; Fredeen *et al.*, 1990). Limited availability of other nutrients (e.g. calcium,  
160 magnesium, sulphur, zinc, and iron, among others) can limit plant growth and reduce the  
161 nutritional quality of the harvested product, but research on their effects on plant processes  
162 is sparse. Responses to ozone concentrations (O<sub>3</sub>) are expected to negatively affect leaf area  
163 dynamics, light interception and biomass allocation and accumulation, but data scarcity has  
164 precluded accurate simulation of this process (Ewert and Porter, 2000). Understanding,  
165 parameterising, and evaluating many of these responses in models is essential for impacts  
166 science.

167

### 168 **3. Approaches for assessing climate impacts**

169 Methods to assess impacts can be classified in projection-based approaches and utility-  
170 based approaches. Utility-based approaches (also known as decision-based approaches)  
171 focus on making decisions that are robust against the known uncertainties. This is usually  
172 done by exploring the outcomes of decisions under a number of plausible scenarios and  
173 then choosing those decisions whose outcomes are not affected by the underlying  
174 uncertainties (Vermeulen *et al.*, 2013). Projection-based approaches (also known as predict-  
175 then-act approaches) are based on the use of models and data to produce projections of a  
176 given system's future state that can be used by decision makers. Projection-based  
177 approaches therefore focus on reducing uncertainties in order to provide decision-makers  
178 with information that can be directly used to make a decision. As with most of the  
179 modelling literature, this review focuses on projection-based approaches. In the following  
180 sections, a summary of related methods is provided. For further discussion on decision-  
181 based approaches the reader is referred to Vermeulen *et al.* (2013).

182

183 In projection-based frameworks, typically, global climate model projections for one or  
184 more given forcing scenarios are first scaled and/or bias-corrected to produce climate  
185 scenarios. Crop models are then forced using these climate scenarios to produce a range of  
186 projections that are then used to conceptualise and develop adaptation strategies to be tested  
187 or implemented at different scales (from global to the field) (Fig. 3). Modelling choices  
188 across the framework shown in Fig. 3 are thus varied and can produce differing responses,  
189 thus causing uncertainty. It is expected for almost all steps in the impact assessment process  
190 that uncertainty will increase, although it can be reduced via model calibration and

191 evaluation. The global meta-analysis of Challinor *et al.* (2014b) is particularly useful  
192 portraying some of the uncertainties to which impact projections are subjected.

193

194 [Figure 3 here]

195

#### 196 **4. The role of process-based models in estimates of climate change impacts and** 197 **adaptation**

198 The choice of both crop models and climate model projection types for climate change  
199 impact assessment varies substantially across modelling studies (White *et al.*, 2011).

200 Nevertheless, the vast majority of projection-based studies focus on a site-specific scale and  
201 use process-based simulation models, though a recent trend exists for regional-scale studies  
202 that use simple (yet process-based) or statistical models (Ramirez-Villegas and Challinor,  
203 2012). Rivington and Koo (2011) report the existence of 122 crop models –from which  
204 roughly a half are process-based. Due to the focus of this review, in this section, emphasis  
205 is placed on process-based models.

206

207 Process-based models are both the most diverse and the most complex of the two model  
208 types reviewed here and can themselves be divided into two categories according to scale  
209 and level of complexity: (i) regional-scale and (ii) field-scale. Regional-scale models have  
210 been designed to capitalise on large-scale crop-climate relationships and thus operate at  
211 scales commensurate with those of global and regional climate models (i.e. 25 – 100 km).  
212 Despite their reduced complexity, regional-scale models retain enough mechanistic detail in  
213 plant growth processes as to be used with reasonable confidence under future climate  
214 scenarios, including increased CO<sub>2</sub> concentrations, and higher rates of extreme temperature

215 and drought events (Challinor *et al.*, 2004, 2007). Conversely, field-scale crop models are  
216 tools aimed to simulate growth processes in plants so that technological changes and  
217 environmental effects at the farm level can be assessed (El-Sharkawy, 2005). Initially,  
218 field-scale models were conceived with the objective of being perfect and comprehensive,  
219 and able to reproduce all plant functions [the ‘universal model’ myth, see Sinclair and  
220 Seligman (1996)], though they rapidly evolved into approaches that were theoretically  
221 coherent, yet different in their implementation and purpose (Affholder *et al.*, 2012). While  
222 the choice of which processes to represent in detail, and the level of detail achieved for a  
223 given process is limited by an understanding of crop physiology derived from available  
224 data (Craufurd *et al.*, 2013), it is also governed by research focus and intended model use.  
225 The guiding principle for designing abstractions in such models is to “*Use the right level of*  
226 *description to catch the phenomena of interest. Don’t model bulldozers with quarks*”  
227 (Goldenfeld and Kadanoff, 1999).

228

#### 229 **4.2.1. Designing models for extensibility and correctness**

230 There are three key aspects involved in the development and use of well-established  
231 process-based crop models – (1) the modelling of biophysical processes, (2) the selection  
232 and maintenance of technical methodologies, and (3) collaborative community support.  
233 Modelling biophysical processes involves choosing the right abstractions to map the  
234 interactions of genotype, management and environment to phenotypic traits of interest. The  
235 selection of technical methodology involves choosing programming languages, software  
236 environments, data formats, collaboration software, computing hardware, and protocols for  
237 maintaining model quality (e.g., automated testing) and uncertainty (e.g., model  
238 ensembles). Collaborative community support includes communication between developers

239 of the model, between the modelling team and other expert modelling groups, and between  
240 model developers, users and the wider community of stakeholders (such as farmers,  
241 consultants and policy makers).

242

243 These key modelling aspects have been traditionally undertaken within individual research  
244 groups, often using ad hoc procedures –although with exceptions [e.g. the International  
245 Consortium for Agricultural Systems Applications, ICASA, White *et al.* (2013)]. However,  
246 two relatively recent developments have had a significant impact on the design and  
247 development of process-based crop models. First, a significant increase in available  
248 computer processing power has enabled ever-increasing complexity in the processes being  
249 modelled. ‘Next generation’ frameworks spanning processes from gene expression to  
250 climate change are becoming available (Holzworth *et al.*, 2014). Second, the rapid adoption  
251 of online tools has enabled global collaborative model development (McLaren *et al.*, 2009)  
252 and inter-comparison [AgMIP; Rosenzweig *et al.* (2013)], and changed expectations  
253 regarding the availability of model source code and data.

254

255 Contemporary process-based crop models are increasingly being used to combine sub-  
256 components (such as different crop types and genotypic traits) in novel ways. These models  
257 are typically not developed in isolation, but are the refinement and integration of pre-  
258 existing algorithms, data, and models [Fig. 1 in Holzworth *et al.* (2014)]. In addition, they  
259 are developed and tested in a variety of programming languages and computing  
260 environments, utilizing agronomical and climate data provided in a wide variety of formats.

261

262 This increased complexity of processed-based crop modelling, and the global, cross-  
263 disciplinary nature of model development, assessment, and use, has led to modelling groups  
264 adopting more formal techniques to support their research. In particular, to facilitate  
265 scientific reproducibility, sharing, inter-comparison and integration of sub-models and data,  
266 the crop modelling community is increasingly relying on tools and techniques from the  
267 software development community. The use of support tools such as wikis, source code  
268 version control and issue tracking (as in the GLAM, DSSAT and APSIM communities),  
269 online user interfaces (Hochman *et al.*, 2009), and the adoption of modular source code  
270 frameworks, is becoming more frequent. For example, the current APSIM process-based  
271 crop modelling framework (Holzworth *et al.*, 2014) employs (1) a modular software  
272 structure that allows components to be combined in novel ways at runtime, and to be  
273 improved and tested in isolation, (2) XML configuration files allowing model parameters  
274 and custom logic to be shared in a standardized way, and (3) the integration of scripting  
275 language control (including the R and C# languages) that facilitates quick prototyping and  
276 sharing of model logic.

277

278 While such developments are significant steps towards improved model sharing,  
279 uncertainty analysis, and code correctness, more work needs to be done. Automated testing,  
280 source code version control, and modular model structure are not yet ubiquitous process-  
281 based modelling practices. Standardization of common parameter names and their  
282 definitions would facilitate more complete model intercomparisons. Significant gains can  
283 be achieved through the adaptation of the software design patterns process (Gamma *et al.*,  
284 1994) to document key crop modelling components such as biophysical processes, model  
285 structure, ensemble design, and model intercomparison, in a form independent of any

286 specific implementation or programming language. The development of such patterns  
287 would help reduce the reinvention of solutions, encourage the use of state of the art  
288 procedures, and provide a community platform for crop model improvement.

289

#### 290 **4.2.2. The use of ensembles for informing impacts and adaptation**

291 The aforementioned increase in the complexity and number of models, along with  
292 significant advances in the climate models used to drive regional-scale yield projections,  
293 has led to greater confidence in our model projections. However, increasing model detail  
294 has meant that uncertainty in projections is not being reduced [see, for example, Knutti &  
295 Sedlacek (2012)]. In addition, model simplifications (such as regional scale process-based,  
296 statistical, and niche-based models) have introduced their own uncertainties in terms of  
297 spatio-temporal scaling and specificity, and the inter-related lineage of process-based crop  
298 models complicates assessments of model uncertainty. As a result, an emphasis on  
299 quantifying the uncertainty in projected yields has become prevalent (Iizumi *et al.*, 2009;  
300 Asseng *et al.*, 2013). Crop predictions based on single parameter sets or single model  
301 output values are no longer good enough.

302

303 Consequently, projecting crop responses under future climate scenarios requires careful  
304 treatment of issues related to parameter uncertainty, structural uncertainty (model  
305 discrepancy), algorithmic uncertainty (code uncertainty), parametric variability,  
306 experimental uncertainty (observation error), and interpolation uncertainty (Kennedy and  
307 O'Hagan, 2001; Challinor *et al.*, 2009a). While accounting for all of these uncertainty  
308 sources is critical for the robust use environmental models in general, the tendency for crop  
309 models to be developed using information from one spatial scale, and applied at another,

310 means that crop modellers must pay particular attention to parameter, structural, and  
311 interpolation uncertainty. An assessment of 178 published studies on climate change  
312 impacts (sourced by searching the keywords ‘climate change impacts’ in  
313 <http://scholar.google.com> in June 2014) indicates that field-scale, regional-scale process-  
314 based models, and statistical models are used at a variety of spatial scales (Fig. 4). For  
315 field-scale process based models, the fact that ca. 50 % of studies use the models at scales  
316 other than those for which the models were originally designed suggests some potential for  
317 model vs. study scale mismatch or even model misuse (Fig 4A). While mathematically one-  
318 dimensional models can be used across different spatial scales, remarkably, virtually no  
319 study using field-scale process-based models at scales beyond individual fields assesses  
320 parameter uncertainty or parameter scaling issues [Fig. 4B, e.g. Iizumi *et al.* (2014)]. More  
321 importantly, the implications of model misuse, including the use of models that lack key  
322 processes and scale mismatches, may impact further estimates of adaptation (Challinor *et*  
323 *al.*, 2014a; Lobell, 2014). This is of particular importance since about one in every three  
324 studies does not conduct model evaluation regardless of the type of model used (Fig. 4C).

325

326 [Figure 4 here]

327

328 In the last ten years, the critical task of quantifying and accounting for the full range of  
329 uncertainty sources in models has been recognized by the weather, climate, and  
330 hydrological communities (Stainforth *et al.*, 2005; Beven, 2006). However, there has been  
331 limited applied appreciation for these issues in the crop modelling community besides  
332 quantifying parameter (Iizumi *et al.*, 2009; Tao and Zhang, 2013) and structural uncertainty  
333 in impacts projections (Ruane *et al.*, 2013; Asseng *et al.*, 2013). While many crop-climate

334 impact studies include some treatment of modelling uncertainty (e.g. by using various  
335 future climate projections, crop parameters, and crop models), sampling of the entire model  
336 and parameter space is fundamentally incomplete, and is likely to underestimate the  
337 importance of uncertainty in model-based projections of impacts and adaptation. Therefore,  
338 in order for the crop modelling community to move towards ensembles that better sample  
339 the uncertainty space and provide useful information for food security assessments,  
340 platforms that allow model, parameter and input transferability between groups and regions  
341 so as to facilitate ensemble simulations for both site- and regional-scale assessments need  
342 to be developed (also see **Sect. 4.2.1**). Additionally, characterising the crop model space  
343 [e.g. Angulo *et al.* (2013)] and better understanding of parameter and process scaling  
344 (Iizumi *et al.*, 2014) will ultimately allow for a better understanding and sampling of the  
345 model and parameter uncertainty space.

346

## 347 **5. Design of genotypic adaptation strategies using crop models**

### 348 **5.1. The importance of genotypic adaptation**

349 Genotypic adaptation is expected to be one of the most important adaptation strategies to  
350 future climate change (Challinor *et al.*, 2009b; Semenov and Stratonovitch, 2013). For  
351 instance, Challinor *et al.* (2014b) indicated that switching from currently grown to better-  
352 adapted varieties that are cultivated elsewhere or stored at genebanks ('cultivar  
353 adjustment') is a more effective adaptation strategy than adjusting planting dates,  
354 improving irrigation and enhancing fertilisation (**Fig. 5**). In addition, increased evidence  
355 exists that climate change stresses can, to a large extent, be managed or completely offset  
356 through the breeding of new "climate-smart" cultivars with improved yield potential and  
357 stability (Ortiz *et al.*, 2008; Semenov and Stratonovitch, 2013). Progress in crop breeding

358 demonstrates the scales of potential yield gains. In Africa, two decades of maize breeding  
359 have led to mean genetic gains of 14 kg ha<sup>-1</sup> year<sup>-1</sup> under drought and 40 kg ha<sup>-1</sup> year<sup>-1</sup>  
360 under optimum conditions (Badu-Apraku *et al.*, 2013). Similarly, global mean wheat  
361 breeding gains in the last 25 years are about 100 kg ha<sup>-1</sup> year<sup>-1</sup> under drought and 25 kg ha<sup>-1</sup>  
362 year<sup>-1</sup> under optimum conditions (Gourdji *et al.*, 2013). For rice, genetic gains have been  
363 estimated in 45 kg ha<sup>-1</sup> year<sup>-1</sup> for Brazilian upland systems in the period 2002-2009  
364 (Breseghello *et al.*, 2011), whereas in irrigated rice in Asia solely the release of the semi-  
365 dwarf rice variety IR8 produced an increase of almost 70 % in rice potentials during the  
366 1950s and 1960s (Peng *et al.*, 2008).

367

368 [Figure 5 here]

369

370 Under future climate scenarios, ideotype design appears as key strategy to drive breeding  
371 decisions, since breeding towards a crop ideotype is more efficient than breeding to remove  
372 undesired characteristics one at a time (Peng *et al.*, 2008). Crop ideotypes are idealised  
373 plant types that have the greatest effectiveness in producing dry matter and yield under  
374 given environmental conditions (Donald, 1968). Defining a crop ideotype involves a  
375 definition of the physical-morphological (e.g. height, maximum leaf size, leaf thickness and  
376 positioning) and physiological (e.g. stomatal conductance, photosynthetic efficiency)  
377 characteristics of a given crop plant, that would allow such a plant to respond well under  
378 certain conditions (e.g. in a drought-prone environment). Breeding programmes are  
379 currently challenged with having to set priorities based on climate change impacts  
380 projections [see e.g. Cairns *et al.* (2013)]. Decisions of which traits to breed and by when  
381 would varieties need to hold such traits are expected to be largely influenced by the type

382 (e.g. increase in mean, increase in extreme events), direction (e.g. drier and warmer, wetter  
383 and warmer), and extent (how warmer, how drier) of the projected climatic changes in a  
384 given area (Stamp and Visser, 2012). Many breeding programs, however, already work  
385 towards achieving crop ideotypes for different agro-environmental zones (Berry *et al.*,  
386 2007; Peng *et al.*, 2008). Hence, progress towards better future food security prospects of  
387 increased food availability and stability through breeding better adapted crop varieties  
388 seems, at least in principle, possible to achieve.

389

## 390 **5.2. The potential role of crop models for developing genotypic adaptation options**

391 Process-based crop models can help make informed decisions with regards to genotypic  
392 adaptation options and ideotype design both under current and future climates (Baenziger *et al.*  
393 *al.*, 2004; Banterng *et al.*, 2004). The main challenge, however, is to carefully interpret  
394 modelling outcomes so as to provide information that is of use for breeders. Recent  
395 experiences in the use of crop model simulated ideotypes for crop breeding in rice as well  
396 as existing model-based investigations of genotypic adaptation and ideotype design reveal  
397 encouraging results with regards to increasing food availability and stability in the context  
398 of climate change adaptation.

399

400 Under current climates, probably the most notable example of ideotype design for  
401 increasing yield potential is the New Plant Type (NPT) proposed and developed by the  
402 International Rice Research Institute (IRRI) and the subsequent establishment of the *super*  
403 rice program in China inspired by the NPT (Cheng *et al.*, 2007; Peng *et al.*, 2008). IRRI's  
404 NPT had its origins on the work of Dingkuhn *et al.* (1991), who used a process-based  
405 growth simulation model to propose a rice ideotype. Based upon model simulations, they

406 hypothesised that 25 % productivity gains could be achieved by increasing the length of the  
407 grain filling phase, maintaining high concentration of nitrogen in the leaves, increasing the  
408 vertical gradient of nitrogen in the foliage (so that top leaves have more N, and lower  
409 leaves have less), enhancing leaf growth in early stages and reduced leaf growth in later  
410 stages, larger panicles but reduced tillering capacity (i.e. lower number of panicles), more  
411 assimilates in the stems and longer life span and larger size of flag leaves (Dingkuhn *et al.*,  
412 1991). Since morphological characteristics are easier to select for in breeding trials, a more  
413 precise definition of these was done in a subsequent study (Khush, 1995) (Fig. 6). Two  
414 breeding cycles then led to the development of NPT varieties that outyielded check  
415 varieties (Peng *et al.*, 2008). Following IRRI's promising results, the *super* rice program in  
416 China was established (Cheng *et al.*, 2007). In addition to what had been proposed by  
417 IRRI's NPT breeding program, a more specific definition of the position and size of the  
418 flag leaves and an optimisation of photosynthetic efficiency were done. Newly developed  
419 *super* rice varieties reportedly outyielded commonly cultivated rice hybrids by 15-25 % in  
420 many regions of China (Peng *et al.*, 2008). Further research and development of ideotype  
421 rice cultivars and hybrids is currently being pursued both in China and internationally by  
422 IRRI.

423

424

[Figure 6 here]

425

426 Under future climates, on the contrary, to the knowledge of the authors, no breeding  
427 program is currently breeding a model-designed plant type; although the WHEAt and  
428 barley Legacy for Breeding Improvement (WHEALBI) appears as a new (started in early  
429 2014) promising initiative. Nevertheless, the number of studies investigating genotypic

430 adaptation through introduction of novel traits and ideotype design has been increasing in  
431 the last decade. All these studies point to the same direction: that in most situations  
432 genotypic adaptation can offset climate change-related losses and even boost crop yields.  
433 For instance, studies for wheat indicate that climate-ready varieties would outyield  
434 currently cultivated varieties by 25-65 % under future climates (Semenov *et al.*, 2014), and  
435 similar figures have been reported for other crops such as groundnut, sorghum and maize  
436 (Fig. 7). These figures are, however, contingent on two key modelling aspects:

437 (i) *The ability of the model to correctly simulate processes that are relevant in future*  
438 *climate scenarios.* The fact that all existing models have been subjected to varying  
439 degrees of evaluation mostly against agronomic trial data [e.g. Asseng *et al.* (2013),  
440 Bassu *et al.* (2014)] and many individual model components (e.g. water balance,  
441 photosynthesis response) are often assessed independently has increased confidence  
442 on the capabilities of models to simulate crop responses under varying environmental  
443 conditions, including climate change. Recent literature, however, indicates shifting  
444 climate distributions and increased likelihood of extreme events (Battisti and Naylor,  
445 2009; Trnka *et al.*, 2014) and this may result in additional and/or different processes  
446 constraining future crop yields as compared to present-day conditions. Indeed, a  
447 recent review identified that only a handful ( $\leq 6$ ) of crop models currently used in  
448 impact and adaptation studies simulate CO<sub>2</sub> impacts on canopy temperature (by  
449 computing a soil-plant-atmosphere energy balance), a key process under climate  
450 change (White *et al.*, 2011). It is thus not clear whether models already include  
451 sufficient detail so as to simulate any additional processes that may arise from  
452 projected climate change. This has in turn resulted in the need for additional field  
453 experiments in which novel conditions and their interaction are evaluated and then

454 tested in multi-model intercomparison frameworks (Rosenzweig *et al.*, 2013). While  
455 these initiatives are clearly a way forward, individual-study assessments of processes  
456 and their interactions in single-model and multi-model ensemble simulations as well  
457 as more complete descriptions of model limitations with respect to key missing  
458 processes are warranted in future genotypic adaptation studies [see e.g. Fig. 1 in  
459 Singh *et al.* (2014)]. Achieving a better representation of future-climate relevant  
460 processes will ensure that model-based analyses are more realistic.

461 (ii) *The correct separation between model parameters that influence yield as a function of*  
462 *crop physiology and those with large impact on simulated yield only due to model*  
463 *specification.* That is, the possibility of relating model parameters to the effect of  
464 alleles on given loci or genes controlling key traits (Luquet *et al.*, 2012). Importantly,  
465 there is a tight link between such a relationship and model complexity –an  
466 overarching issue in climate impacts prediction, because overly simplistic models are  
467 unlikely to capture physiological responses with enough level of detail for use in crop  
468 breeding (Luquet *et al.*, 2012), but overly complex models are more difficult to  
469 constrain at the scales typical of climate prediction frameworks (Challinor *et al.*,  
470 2009a). Work towards linking quantitative trait loci information and process-based  
471 crop growth modelling, however, shows promise. For example, Chenu *et al.* (2009)  
472 used a gene-to-phenotype modelling approach that included a genetic model and a  
473 process-based crop model to simulate the impact of leaf and silk elongation traits (as  
474 derived from genetic data) on maize yield across different environments. Despite  
475 some success, however, the lack of a more thorough consideration of genetic effects  
476 [beyond those related to crop development (Messina *et al.*, 2006; Challinor *et al.*,  
477 2009b)] on yield and genotype-by-environment interactions in genotypic adaptation

478 studies suggests that appropriate frameworks need to be established [e.g. Chenu *et al.*  
479 (2009); Cooper *et al.* (2005)].

480

481 [Figure 7 here]

482

483 We suggest that in order to mainstream crop-model based analyses of genotypic adaptation  
484 into breeding programmes, more research as well as a framework on the coupling of crop  
485 and genetic models is needed. Fig. 8, based on the work of Chenu *et al.* (2009) and Cooper  
486 *et al.* (2005), is an attempt to such a framework, through which we expect model-based  
487 analyses of genotypic adaptation can incorporate genetic information from breeding  
488 programs and, in turn, retrieve *ex-ante* assessments of genotypic responses across  
489 environments [also see Yin *et al.* (2003, 2004)]. As a starting point, traits that have constant  
490 QTLs (and hence constant model parameters) across environments have to be determined.  
491 Modular crop models can then be coupled with ‘plug-and-play’ parameterisations of  
492 relevant characteristics for which genetic information is available, with appropriate  
493 sensitivity testing to ensure realism. Genetic model simulations of crosses between  
494 promising parental lines can then yield crop model parameters and be run through an  
495 ensemble of crop models at one or more environments. The resulting crop model  
496 simulations can then be used to select promising phenotypes and the process repeated for  
497 various steps in the breeding cycle (Fig. 8).

498

499 [Figure 8 here]

500

501 Additionally, simulation of genotypic adaptation (including ideotype design) for projected  
502 weather conditions of uncertain nature means that additional principles may be needed in  
503 order to develop robust projections of adaptation. In particular, appropriately constraining  
504 processes and parameters in models across scales [cf. Iizumi *et al.* (2014)] and a shift from  
505 simply quantifying uncertainty to actually quantifying robustness (i.e. the relationship  
506 between uncertainty and the climate change signal) towards modelling choices [cf.  
507 Ramirez-Villegas and Challinor (2014)] are two key aspects that need to be included into  
508 crop model-based analyses of genotypic adaptation. Two key initiatives toward these aims  
509 include the AgMIP (Rosenzweig *et al.*, 2013) and FACCE-MACSUR  
510 (<http://www.macsur.eu>) projects.

511

## 512 **Conclusions**

513 The challenges ahead with regards of developing genotypic adaptation strategies that can  
514 then be implemented in breeding programs are substantial. On one hand, climate change  
515 impacts are projected to pose significant challenges to agriculture and genotypic adaptation  
516 strategies are critical for responding to such challenges. On the other hand, uncertainties in  
517 climate and crop modelling are substantial and poorly explored in studies of genotypic  
518 adaptation to future climates that use process-based simulation models, particularly at field  
519 scales. While uncertainties need to be better understood and quantified (see **Sect. 5**), it is  
520 important to note that a shift in focus from solely quantifying output variance to  
521 quantifying robustness is required in order so as to facilitate assessments and interpretation  
522 of confidence levels in crop model-based projections of genotypic adaptation. In addition to  
523 this, it is critical that genotypic adaptation options are grounded in genetic and  
524 physiological knowledge that can be mainstreamed in real-world breeding programs. To

525 this aim, while recently published studies have demonstrated the potential of genotypic  
526 adaptation strategies and ideotype design, two main goals need to be pursued in future  
527 studies: (1) a better understanding of driving processes under future climate change; and (2)  
528 a coupling between genetic and crop growth models –perhaps at the expense of number of  
529 traits analysed. Importantly, the latter may imply additional complexity [and likely  
530 uncertainty] in crop modelling studies. Therefore, modularity in crop models as well as  
531 individual component testing against observational data would be critical components in  
532 any attempts to simulate crop breeding strategies under future climate scenarios.

533

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544

#### **References**

**Affholder F, Tittone P, Corbeels M, Roux S, Motisi N, Tixier P, Wery J.** 2012. Ad Hoc Modeling in Agronomy: What Have We Learned in the Last 15 Years? *Agron. J.* **104**, 735–748.

**Ainsworth EA, Leakey ADB, Ort DR, Long SP.** 2008. FACE-ing the facts: inconsistencies and interdependence among field, chamber and modeling studies of elevated [CO<sub>2</sub>] impacts on crop yield and food supply. *New Phytologist* **179**, 5–9.

**Allen LH, Boote KJ, Vara Prasad P V, Thomas JMG, Vu JC V.** 2005. *Hazards of Temperature on Food Availability in Changing Environments (Hot-FACE): Global Warming Could Cause Failure of Seed Yields of Major Food Crops*. Broomfield, Colorado, USA.

**Angulo C, Rötter R, Trnka M, Pirttioja N, Gaiser T, Hlavinka P, Ewert F.** 2013. Characteristic “fingerprints” of crop model responses to weather input data at different spatial resolutions. *European Journal of Agronomy* **49**, 104–114.

**Asseng S, Ewert F, Rosenzweig C, et al.** 2013. Uncertainty in simulating wheat yields under climate change. *Nature Climate Change* **3**, 827–832.

**Asseng S, Foster IAN, Turner NC.** 2011. The impact of temperature variability on wheat yields. *Global Change Biology* **17**, 997–1012.

**Badu-Apraku B, Oyekunle M, Menkir A, Obeng-Antwi K, Yallou CG, Usman IS, Alidu H.** 2013. Comparative Performance of Early-maturing Maize Cultivars Developed in Three Eras under Drought Stress and Well-watered Environments in West Africa. *Crop Science* **53**, 1298.

**Baenziger PS, McMaster GS, Wilhelm WW, Weiss A, Hays CJ.** 2004. Putting genes into genetic coefficients. *Field Crops Research* **90**, 133–143.

**Bantern P, Patanothai A, Pannangpetch K, Jogloy S, Hoogenboom G.** 2004. Determination and evaluation of genetic coefficients of peanut lines for breeding applications. *European Journal of Agronomy* **21**, 297–310.

**Bassu S, Brisson N, Durand J-L, et al.** 2014. How do various maize crop models vary in their responses to climate change factors? *Global change biology* **20**, 2301–2320.

**Battisti DS, Naylor RL.** 2009. Historical Warnings of Future Food Insecurity with Unprecedented Seasonal Heat. *Science* **323**, 240–244.

**Berry PM, Sylvester-Bradley R, Berry S.** 2007. Ideotype design for lodging-resistant wheat. *Euphytica* **154**, 165–179.

**Beven K.** 2006. A manifesto for the equifinality thesis. *Journal of Hydrology* **320**, 18–36.

- Bird IF, Cornelius MJ, Keys AJ.** 1977. Effects of Temperature on Photosynthesis by Maize and Wheat. *Journal of Experimental Botany* **28**, 519–524.
- Boote KJ, Allen LH, Vara Prasad P V, Baker JT, Gesch RW, Snyder AM, Pan D, Thomas JMG.** 2005. Elevated Temperature and CO<sub>2</sub> Impacts on Pollination, Reproductive Growth, and Yield of Several Globally Important Crops. *Japanese Journal of Meteorology* **60**, 469–474.
- Breseghele F, de Morais OP, Pinheiro PV, et al.** 2011. Results of 25 Years of Upland Rice Breeding in Brazil. *Crop Science* **51**, 914.
- Cairns JE, Hellin J, Sonder K, Araus JL, MacRobert JF, Thierfelder C, Prasanna BM.** 2013. Adapting maize production to climate change in sub-Saharan Africa. *Food Security* **5**, 345–360.
- Challinor AJ, Osborne T, Shaffrey L, Weller H, Morse A, Wheeler T, Vidale PL.** 2009a. Methods and Resources for Climate Impacts Research. *Bulletin of the American Meteorological Society* **90**, 836–848.
- Challinor AJ, Parkes B, Ramirez-Villegas J.** 2014a. Land-use heterogeneity causes systematic bias in crop yield response to climate change. *Global Change Biology* **in press**.
- Challinor AJ, Simelton ES, Fraser EDG, Hemming D, Collins M.** 2010. Increased crop failure due to climate change: assessing adaptation options using models and socio-economic data for wheat in China. *Environmental Research Letters* **5**, 34012.
- Challinor AJ, Watson J, Lobell DB, Howden SM, Smith DR, Chhetri N.** 2014b. A meta-analysis of crop yield under climate change and adaptation. *Nature Climate Change* **4**, 287–291.
- Challinor AJ, Wheeler TR, Craufurd PQ, Ferro CAT, Stephenson DB.** 2007. Adaptation of crops to climate change through genotypic responses to mean and extreme temperatures. *Agriculture, Ecosystems & Environment* **119**, 190–204.
- Challinor AJ, Wheeler TR, Craufurd PQ, Slingo JM, Grimes DIF.** 2004. Design and optimisation of a large-area process-based model for annual crops. *Agricultural and Forest Meteorology* **124**, 99–120.
- Challinor A, Wheeler T, Hemming D, Upadhyaya H.** 2009b. Ensemble yield simulations: crop and climate uncertainties, sensitivity to temperature and genotypic adaptation to climate change. *Climate Research* **38**, 117–127.
- Cheng S-H, Cao L-Y, Zhuang J-Y, Chen S-G, Zhan X-D, Fan Y-Y, Zhu D-F, Min S-K.** 2007. Super Hybrid Rice Breeding in China: Achievements and Prospects. *Journal of Integrative Plant Biology* **49**, 805–810.

- Chenu K, Chapman SC, Tardieu F, McLean G, Welcker C, Hammer GL.** 2009. Simulating the yield impacts of organ-level quantitative trait loci associated with drought response in maize: a “gene-to-phenotype” modeling approach. *Genetics* **183**, 1507–23.
- Claessens L, Antle JM, Stoorvogel JJ, Valdivia RO, Thornton PK, Herrero M.** 2012. A method for evaluating climate change adaptation strategies for small-scale farmers using survey, experimental and modeled data. *Agricultural Systems* **111**, 85–95.
- Cooper M, Podlich DW, Smith OS.** 2005. Gene-to-phenotype models and complex trait genetics. *Australian Journal of Agricultural Research* **56**, 895.
- Crafts-Brandner SJ, Salvucci ME.** 2002. Sensitivity of photosynthesis in a C4 plant, maize, to heat stress. *Plant physiology* **129**, 1773–80.
- Craufurd PQ, Vadez V, Jagadish SVK, Prasad PVV, Zaman-Allah M.** 2013. Crop science experiments designed to inform crop modeling. *Agricultural and Forest Meteorology* **170**, 8–18.
- Deryng D, Sacks WJ, Barford CC, Ramankutty N.** 2011. Simulating the effects of climate and agricultural management practices on global crop yield. *Global Biogeochem. Cycles* **25**, GB2006.
- Dingkuhn M, Penning de Vries FWT, De Datta SK, van Laar HH.** 1991. Concepts for a new plant type for direct seeded flooded tropical rice. *Direct Flooded Rice in the Tropics*. Los Baños, Philippines: International Rice Research Institute, 17–38.
- Donald CM.** 1968. The breeding of crop ideotypes. *Euphytica* **17**, 385–403.
- El-Sharkawy M.** 2005. How can calibrated research-based models be improved for use as a tool in identifying genes controlling crop tolerance to environmental stresses in the era of genomics—from an experimentalist’s perspective. *Photosynthetica* **43**, 161–176.
- El-Sharkawy MA.** 2014. Global warming: causes and impacts on agroecosystems productivity and food security with emphasis on cassava comparative advantage in the tropics/subtropics. *Photosynthetica* **52**, 161–178.
- Ewert F, Porter JR.** 2000. Ozone effects on wheat in relation to CO<sub>2</sub>: modelling short-term and long-term responses of leaf photosynthesis and leaf duration. *Global Change Biology* **6**, 735–750.
- Fredeen AL, Raab TK, Rao IM, Terry N.** 1990. Effects of phosphorus nutrition on photosynthesis in *Glycine max* (L.) Merr. *Planta* **181**, 399–405.
- Gamma E, Helm R, Johnson R, Vlissides J.** 1994. *Design patterns: Elements of reusable object-oriented software*. Reading, UK: Addison-Wesley.

**Ghannoum O, Caemmerer S Von, Ziska LH, Conroy JP.** 2000. The growth response of C4 plants to rising atmospheric CO2 partial pressure: a reassessment. *Plant, Cell and Environment* **23**, 931–942.

**Gibson LR, Paulsen GM.** 1999. Yield Components of Wheat Grown under High Temperature Stress during Reproductive Growth. *Crop Science* **39**, 1841.

**Goldenfeld N, Kadanoff LP.** 1999. Simple Lessons from Complexity. *Science* **284**, 87–89.

**Gourdji SM, Mathews KL, Reynolds M, Crossa J, Lobell DB.** 2013. An assessment of wheat yield sensitivity and breeding gains in hot environments. *Proceedings of the Royal Society B: Biological Sciences* **280**.

**Hew C-S, Krotkov G, Canvin DT.** 1969. Effects of Temperature on Photosynthesis and CO2 Evolution in Light and Darkness by Green Leaves. *Plant Physiology* **44**, 671–677.

**Hochman Z, van Rees H, Carberry PS, et al.** 2009. Re-inventing model-based decision support with Australian dryland farmers. 4. Yield Prophet □ helps farmers monitor and manage crops in a variable climate. *Crop and Pasture Science* **60**, 1057.

**Holzworth DP, Huth NI, DeVoil PG, et al.** 2014. APSIM – Evolution towards a new generation of agricultural systems simulation. *Environmental Modelling & Software*.

**Huntingford C, Hugo Lambert F, Gash JHC, Taylor CM, Challinor AJ.** 2005. Aspects of climate change prediction relevant to crop productivity. *Philosophical Transactions of the Royal Society B: Biological Sciences* **360**, 1999–2009.

**Iizumi T, Tanaka Y, Sakurai G, Ishigooka Y, Yokozawa M.** 2014. Dependency of parameter values of a crop model on the spatial scale of simulation. *Journal of Advances in Modeling Earth Systems* **in press**.

**Iizumi T, Yokozawa M, Nishimori M.** 2009. Parameter estimation and uncertainty analysis of a large-scale crop model for paddy rice: Application of a Bayesian approach. *Agricultural and Forest Meteorology* **149**, 333–348.

**Kennedy MC, O’Hagan A.** 2001. Bayesian calibration of computer models. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* **63**, 425–464.

**Khush GS.** 1995. Breaking the yield frontier of rice. *GeoJournal* **35**, 329–332.

**Khush GS.** 2001. Green revolution: the way forward. *Nature reviews. Genetics* **2**, 815–22.

**Knutti R, Sedláček J.** 2012. Robustness and uncertainties in the new CMIP5 climate model projections. *Nature Climate Change* **3**, 369–373.

**Kobza J, Edwards GE.** 1987. Influences of Leaf Temperature on Photosynthetic Carbon Metabolism in Wheat. *Plant Physiology* **83**, 69–74.

**Labate CA, Adcock MD, Leegood RC.** 1990. Effects of temperature on the regulation of photosynthetic carbon assimilation in leaves of maize and barley. *Planta* **181**, 547–54.

**Leakey ADB.** 2009. Rising atmospheric carbon dioxide concentration and the future of C4 crops for food and fuel. *Proceedings. Biological sciences / The Royal Society* **276**, 2333–43.

**Leakey ADB, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP, Ort DR.** 2009. Elevated CO<sub>2</sub> effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. *Journal of Experimental Botany* **60**, 2859–2876.

**Lobell DB.** 2014. Climate change adaptation in crop production: Beware of illusions. *Global Food Security*.

**Lobell DB, Burke MB, Tebaldi C, Mastrandrea MD, Falcon WP, Naylor RL.** 2008. Prioritizing Climate Change Adaptation Needs for Food Security in 2030. *Science* **319**, 607–610.

**Lobell DB, Hammer GL, McLean G, Messina C, Roberts MJ, Schlenker W.** 2013. The critical role of extreme heat for maize production in the United States. *Nature Climate Change* **3**, 497–501.

**Long SP, Ainsworth EA, Leakey ADB, Nösberger J, Ort DR.** 2006. Food for Thought: Lower-Than-Expected Crop Yield Stimulation with Rising CO<sub>2</sub> Concentrations. *Science* **312**, 1918–1921.

**Longstreth DJ, Nobel PS.** 1980. Nutrient Influences on Leaf Photosynthesis. *Plant Physiology* **65**, 541–543.

**Luquet D, Soulié JC, Rebolledo MC, Rouan L, Clément-Vidal A, Dingkuhn M.** 2012. Developmental Dynamics and Early Growth Vigour in Rice 2. Modelling Genetic Diversity Using Ecomeristem. *Journal of Agronomy and Crop Science* **198**, 385–398.

**McLaren CG, Metz T, van den Berg M, Bruskiewich RM, Magor NP, Shires D.** 2009. Chapter 4: Informatics in Agricultural Research for Development. *Advances in Agronomy* **102**, 135–157.

**Messina CD, Jones JW, Boote KJ, Vallejos CE.** 2006. A Gene-Based Model to Simulate Soybean Development and Yield Responses to Environment. *Crop Science* **46**, 456.

**Nagai T, Makino A.** 2009. Differences between rice and wheat in temperature responses of photosynthesis and plant growth. *Plant & cell physiology* **50**, 744–55.

- Ortiz R, Sayre KD, Govaerts B, Gupta R, Subbarao G V, Ban T, Hodson D, Dixon JM, Iván Ortiz-Monasterio J, Reynolds M.** 2008. Climate change: Can wheat beat the heat? *Agriculture, Ecosystems & Environment* **126**, 46–58.
- Parent B, Tardieu F.** 2012. Temperature responses of developmental processes have not been affected by breeding in different ecological areas for 17 crop species. *The New phytologist* **194**, 760–74.
- Peng S, Huang J, Sheehy JE, Laza RC, Visperas RM, Zhong X, Centeno GS, Khush GS, Cassman KG.** 2004. Rice yields decline with higher night temperature from global warming. *Proceedings of the National Academy of Sciences of the United States of America* **101**, 9971–9975.
- Peng S, Khush GS, Virk P, Tang Q, Zou Y.** 2008. Progress in ideotype breeding to increase rice yield potential. *Field Crops Research* **108**, 32–38.
- Porter JR, Semenov MA.** 2005. Crop responses to climatic variation. *Philosophical transactions of the Royal Society of London. Series B, Biological sciences* **360**, 2021–35.
- Porter JR, Xie L, Challinor AJ, Cochrane K, Howden M, Iqbal MM, Lobell DB, Travasso MI.** 2014. *Chapter 7. Food Security and Food Production Systems. Climate Change 2014: Impacts, Adaptation and Vulnerability. Working Group II Contribution to the IPCC 5th Assessment Report.* Geneva, Switzerland.
- Ramirez-Villegas J, Challinor A.** 2012. Assessing relevant climate data for agricultural applications. *Agricultural and Forest Meteorology* **161**, 26–45.
- Ramirez-Villegas J, Challinor AJ.** 2014. Towards a genotypic adaptation strategy for Indian groundnut cultivation using model ensembles. *Climatic Change* **Submitted**.
- Rivington M, Koo J.** 2011. *Report on the Meta-Analysis of Crop Modelling for Climate Change and Food Security Survey.* CGIAR Research Program on Climate Change, Agriculture and Food Security.
- Rosenzweig C, Jones JW, Hatfield JL, et al.** 2013. The Agricultural Model Intercomparison and Improvement Project (AgMIP): Protocols and pilot studies. *Agricultural and Forest Meteorology* **170**, 166–182.
- Ruane AC, Cecil LD, Horton RM, Gordón R, McCollum R, Brown D, Killough B, Goldberg R, Greeley AP, Rosenzweig C.** 2013. Climate change impact uncertainties for maize in Panama: Farm information, climate projections, and yield sensitivities. *Agricultural and Forest Meteorology* **170**, 132–145.
- Schmitt MR, Edwards GE.** 1981. Photosynthetic Capacity and Nitrogen Use Efficiency of Maize, Wheat, and Rice: A Comparison Between C 3 and C 4 Photosynthesis. *Journal of Experimental Botany* **32**, 459–466.

- Semenov MA, Stratonovitch P.** 2013. Designing high-yielding wheat ideotypes for a changing climate. *Food and Energy Security* **2**, 185–196.
- Semenov MA, Stratonovitch P, Alhabari F, Gooding MJ.** 2014. Adapting wheat in Europe for climate change. *Journal of Cereal Science* **59**, 245–256.
- Sinclair TR, Horie T.** 1989. Leaf Nitrogen, Photosynthesis, and Crop Radiation Use Efficiency: A Review. *Crop Sci.* **29**, 90–98.
- Sinclair TR, Seligman NG.** 1996. Crop Modeling: From Infancy to Maturity. *Agron. J.* **88**, 698–704.
- Singh P, Boote KJ, Kumar U, Srinivas K, Nigam SN, Jones JW.** 2012. Evaluation of Genetic Traits for Improving Productivity and Adaptation of Groundnut to Climate Change in India. *Journal of Agronomy and Crop Science* **198**, 399–413.
- Singh P, Nedumaran S, Ntare BR, Boote KJ, Singh NP, Srinivas K, Bantilan MCS.** 2013. Potential benefits of drought and heat tolerance in groundnut for adaptation to climate change in India and West Africa. *Mitigation and Adaptation Strategies for Global Change* **19**, 509–529.
- Singh P, Nedumaran S, Traore PCS, Boote KJ, Rattunde HFW, Prasad PVV, Singh NP, Srinivas K, Bantilan MCS.** 2014. Quantifying potential benefits of drought and heat tolerance in rainy season sorghum for adapting to climate change. *Agricultural and Forest Meteorology* **185**, 37–48.
- Stainforth DA, Aina T, Christensen C, et al.** 2005. Uncertainty in predictions of the climate response to rising levels of greenhouse gases. *Nature* **433**, 403–406.
- Stamp P, Visser R.** 2012. The twenty-first century, the century of plant breeding. *Euphytica*, 1–7.
- Tao F, Zhang Z.** 2013. Climate change, wheat productivity and water use in the North China Plain: A new super-ensemble-based probabilistic projection. *Agricultural and Forest Meteorology* **170**, 146–165.
- Trnka M, Rötter RP, Ruiz-Ramos M, Kersebaum KC, Olesen JE, Žalud Z, Semenov MA.** 2014. Adverse weather conditions for European wheat production will become more frequent with climate change. *Nature Climate Change* **4**, 637–643.
- Tubiello FN, Amthor JS, Boote KJ, Donatelli M, Easterling W, Fischer G, Gifford RM, Howden M, Reilly J, Rosenzweig C.** 2007. Crop response to elevated CO<sub>2</sub> and world food supply. *European Journal of Agronomy* **26**, 215–223.

**Vermeulen SJ, Challinor AJ, Thornton PK, et al.** 2013. Addressing uncertainty in adaptation planning for agriculture. *Proceedings of the National Academy of Sciences of the United States of America* **110**, 8357–62.

**White JW, Hoogenboom G, Kimball BA, Wall GW.** 2011. Methodologies for simulating impacts of climate change on crop production. *Field Crops Research* **124**, 357–368.

**White JW, Hunt LA, Boote KJ, Jones JW, Koo J, Kim S, Porter CH, Wilkens PW, Hoogenboom G.** 2013. Integrated description of agricultural field experiments and production: The ICASA Version 2.0 data standards. *Computers and Electronics in Agriculture* **96**, 1–12.

**Wilhelm EP, Mullen RE, Keeling PL, Singletary GW.** 1999. Heat Stress during Grain Filling in Maize: Effects on kernel growth and metabolism. *Crop Science* **39**, 1733.

**Wong SC, Cowan IR, Farquhar GD.** 1979. Stomatal conductance correlates with photosynthetic capacity. *Nature* **282**, 424–426.

**Yin X, Stamb P, Kropff MJ, Schapendonk AHCM.** 2003. Crop Modeling, QTL Mapping, and Their Complementary Role in Plant Breeding. *Agronomy Journal* **95**, 90–98.

**Yin X, Struik PC, Kropff MJ.** 2004. Role of crop physiology in predicting gene-to-phenotype relationships. *Trends in plant science* **9**, 426–32.

## Figure captions

**Figure 1** Percentage variance in historical crop yields explained by seasonal mean temperature and seasonal total precipitation across (A) crops and (B) regions. Variance explained is measured using the coefficient of determination ( $R^2$ ) as derived from the statistical models in Lobell *et al.* (2008). Both panels show the same data, but pooled differently. Variation for each crop in panel A reflects differences between regions and variation for each region in panel B reflects differences between crops. Thick red lines are the medians, boxes represent the interquartile range, whiskers extend to 5-95 % of the data, and red dots are outliers.

**Figure 2** Responses of (A) net photosynthesis to leaf temperature, (B) development rates to mean daily air temperature, and (C) crop yield to temperature during reproductive period. Data in panel A have been derived from the study of Nagai and Makino (2009) for wheat and rice, and from Bird *et al.* (1977), Schmitt *et al.* (1981), Crafts-Brandner and Salvucci (2002), and Labate *et al.* (1990). Solely for illustrative purposes, maize data were fitted to a spline curve with 5 degrees of freedom. Rice and wheat data were fitted to 3<sup>rd</sup> order polynomials as in Nagai and Makino (2009). Curves in panel B were plotted following Parent and Tardieu (2012). Development rates at each temperature in their models have all been normalised by development rates at 20 °C. Data from panel C were derived from Peng *et al.* (2004) for rice [hence  $x$ -axis for rice is minimum growing season temperature], from Gibson and Paulsen (1999) for wheat [hence  $x$ -axis is mean temperature during grain-filling], and from Wilhelm *et al.* (1999) for maize [hence  $x$ -axis is mean temperature post-anthesis]. For panel C all data were linearly scaled so that the maximum yield corresponded to a value of 1. Fits in panel C all follow a linear regression except for rice where the original 2<sup>nd</sup> degree polynomial of Peng *et al.* (2004) was used.

**Figure 3** Ways in which impact assessment is typically approached in projection-based frameworks. Red arrows indicate flow of information. The black hollow arrow in the bottom shows that as long as more information is derived from climate projections, uncertainties are likely to increase, as a result of what is known as “cascade of uncertainties”.

**Figure 4** Use and misuse of crop models, based on 178 model results published in climate change impacts studies between 1994 and 2014, and disaggregated by model type. (A) Fraction of results that perform simulations at the scale for which the model was designed; (B) fraction of results (at scales other than field) for each model type that use multiple parameter sets (i.e. account for parametric uncertainty); and (C) fraction of studies that state model evaluation procedures for their locations or areas of interest. Model types are as follows: CSM-FS: field-scale crop growth simulation model; CSM-RS: regional-scale crop growth simulation model; E/S: empirical and/or statistical. Note that field scale models are used above field scale in roughly 50 % of the cases.

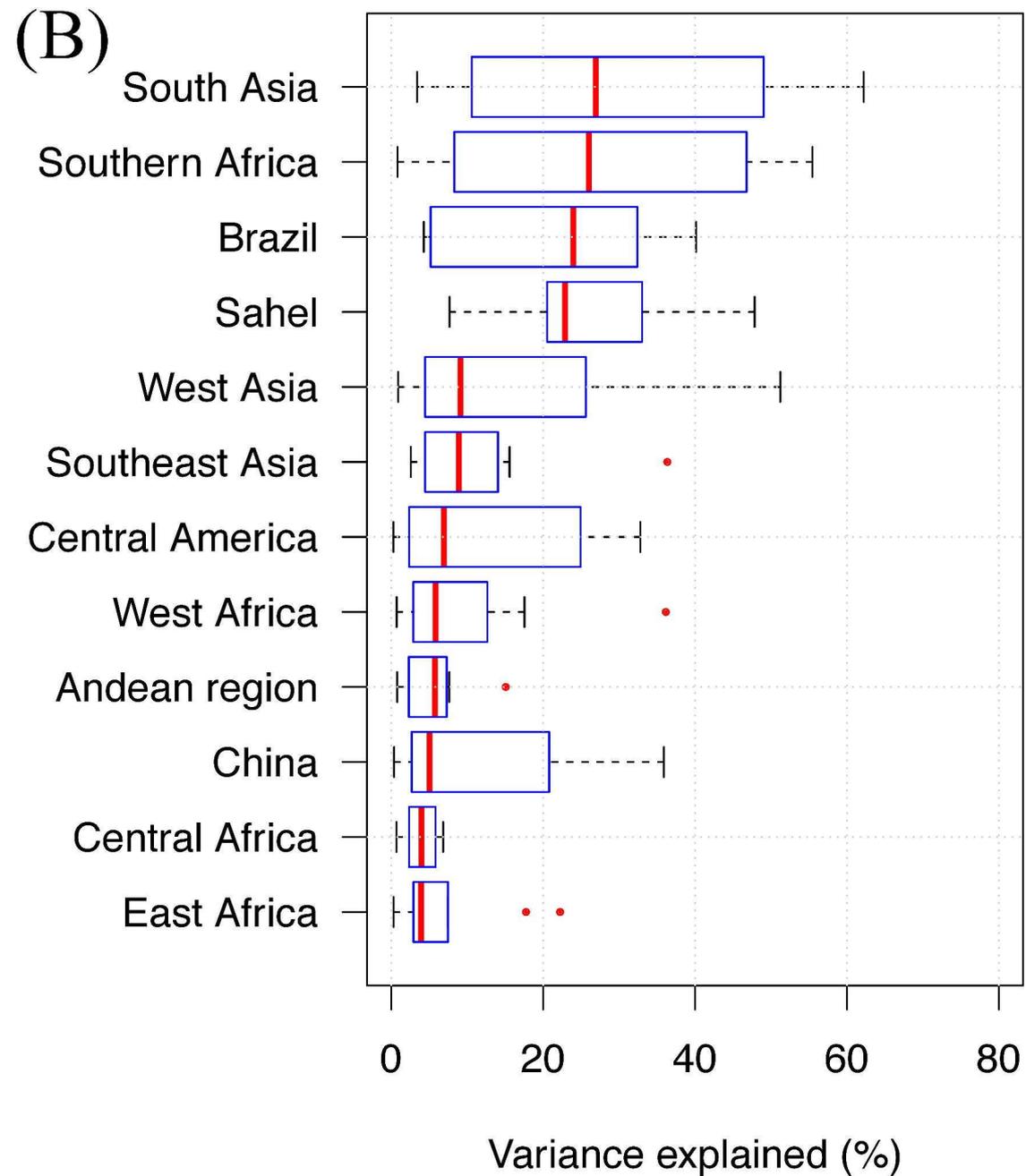
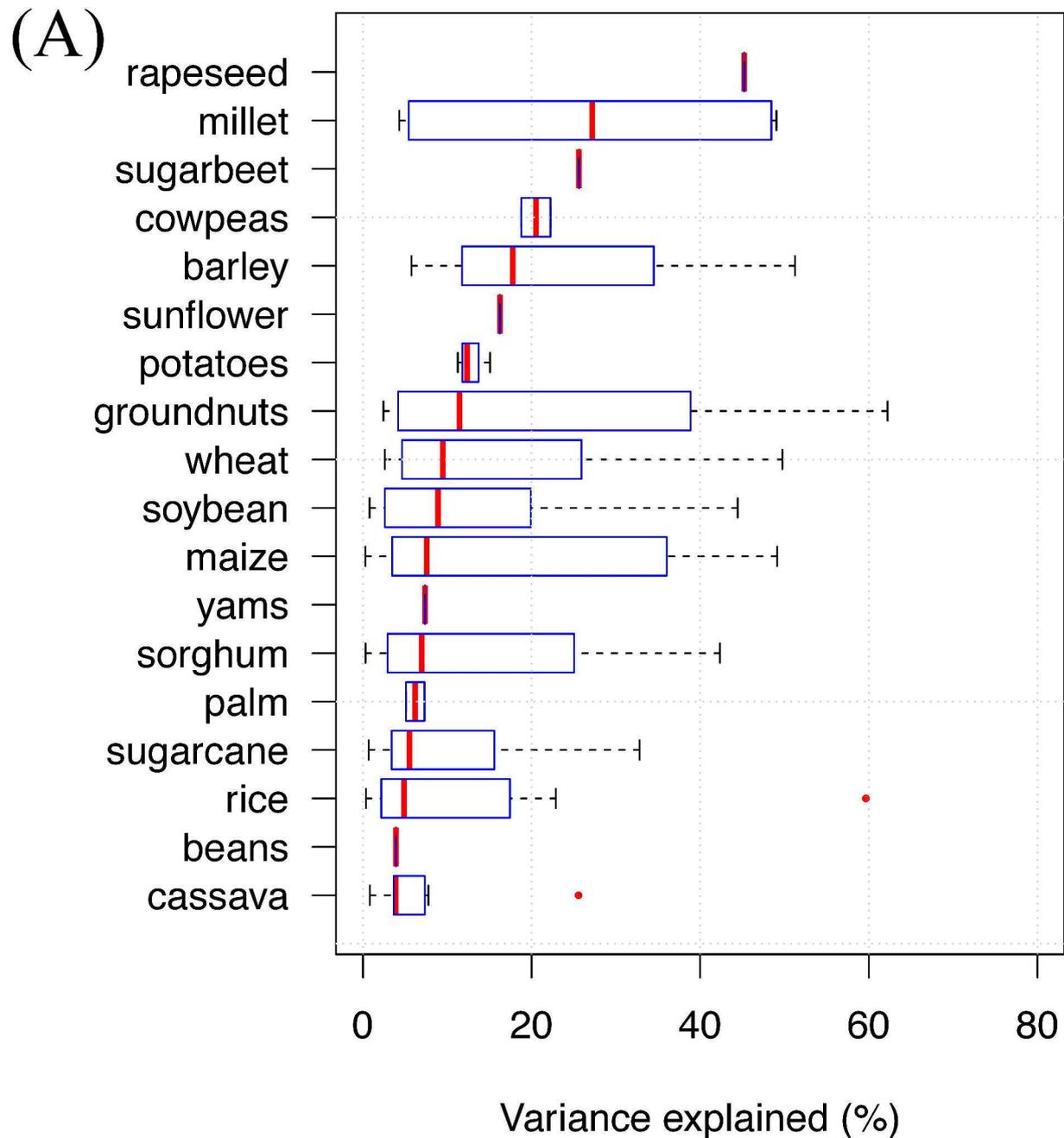
**Figure 5** The benefit of different adaptation practices expressed as percentage change, from the baseline, in yield with adaptation minus that without adaptation, adapted from Challinor *et al.* (2014b). Data in this figure consists of yield changes from 32 simulation studies for various crops as described in Challinor *et al.* (2014b). Bars are means for each category and

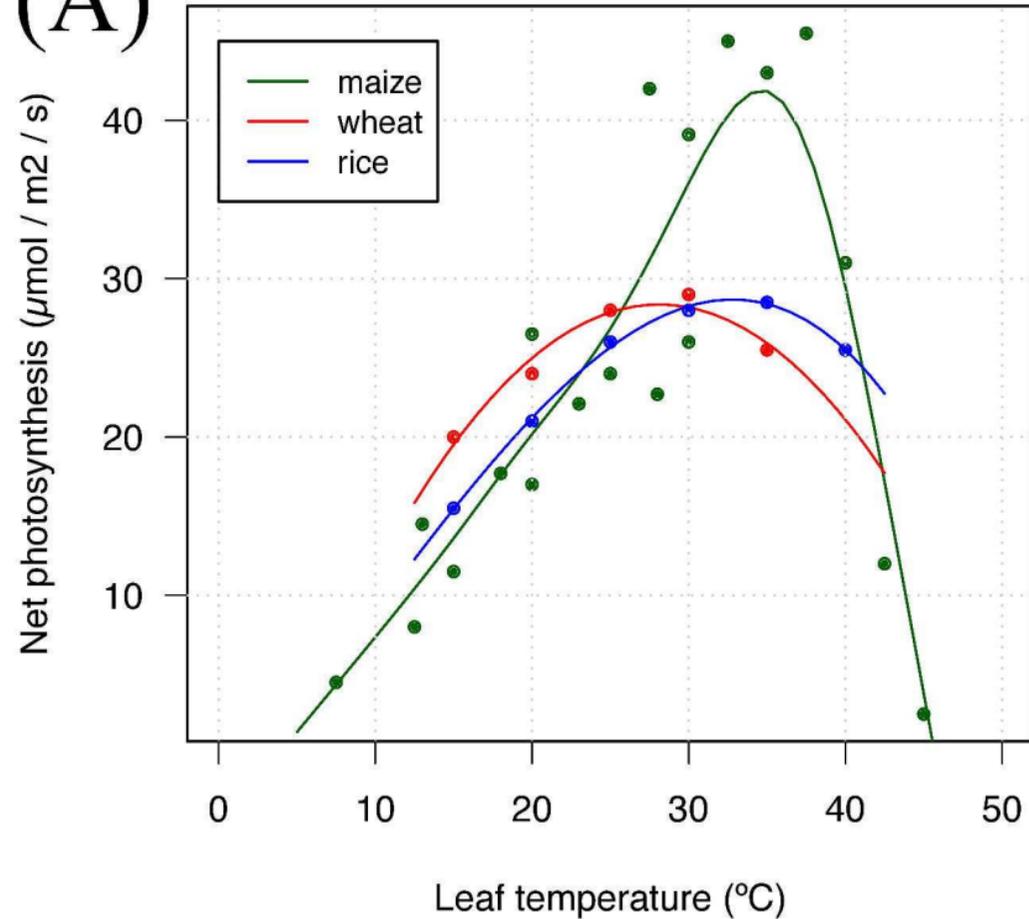
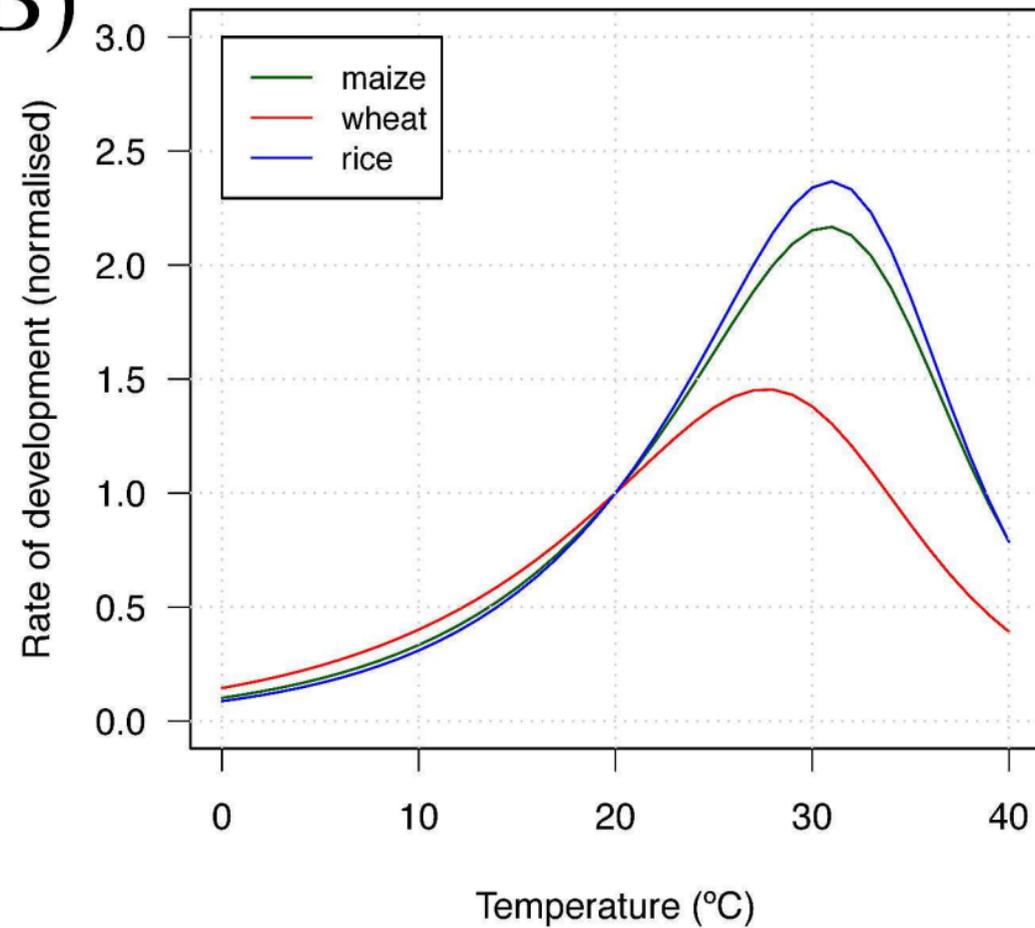
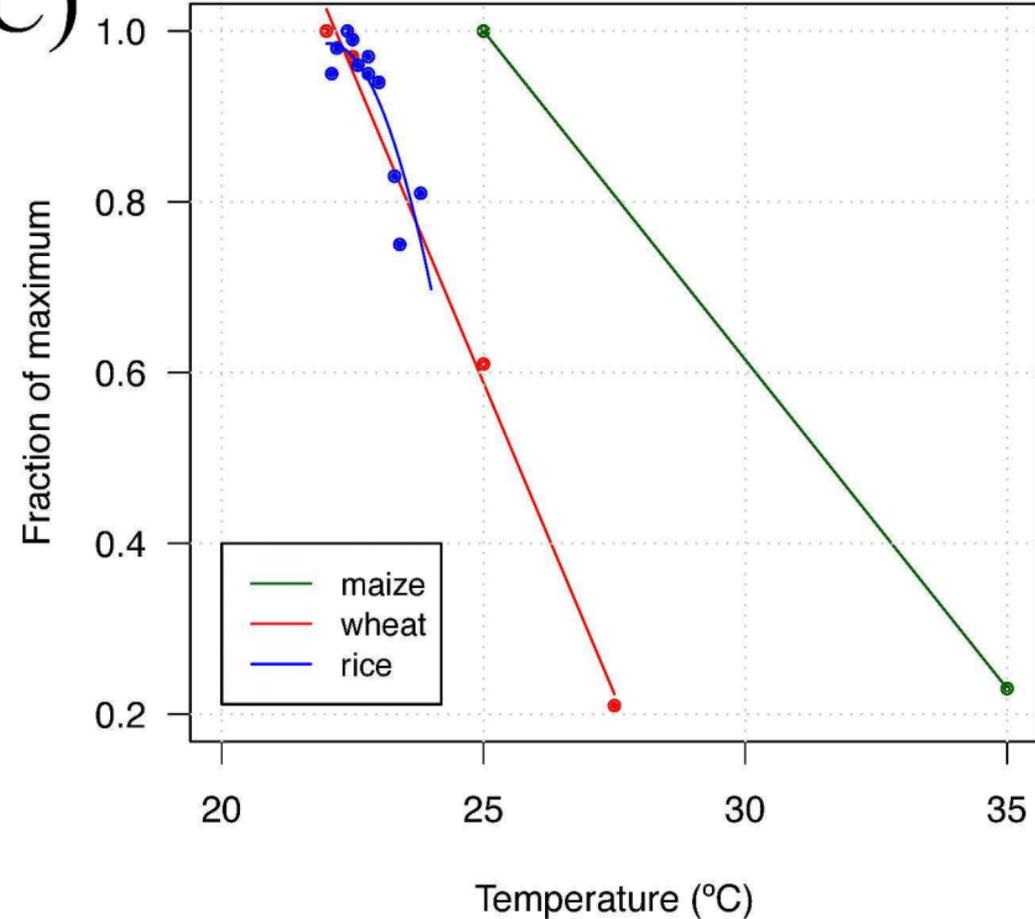
red lines indicate standard error. Note that the vast majority of data in the second category come from a single study (Deryng *et al.*, 2011).

**Figure 6** Different plant types of rice. Left: tall conventional plant type. Centre: improved high-yielding and high-tillering plant type typical of the green revolution. Right: low-tillering ideotype (new plant type) with larger sink capacity (larger panicles and grains) and sturdier stems. Taken from Khush (2001).

**Figure 7** Simulated future potential benefits from genotypic adaptation (including ideotype design) as derived from available modelling studies for four different crops in different sites. Studies are as follows: Semenov and Stratonovitch (2013) and Challinor *et al.* (2010) for wheat; Singh *et al.* (2014) for sorghum; Singh *et al.* (2012, 2013) and Challinor *et al.* (2007, 2009b) for groundnut; and Lobell *et al.* (2013) for maize. The benefit of genotypic adaptation has been calculated as the difference between yield changes under adaptation and that under no adaptation, except in the case of Challinor *et al.* (2010) for which the relative change in crop failure rate between adaptation and no-adaptation results was used. Thick red lines are the medians, boxes represent the interquartile range, whiskers extend to 5-95 % of the data, and red dots are outliers.

**Figure 8** Proposed framework for incorporating genetic information into simulation studies of genotypic adaptation. Figure derived from the practical example of Chenu *et al.* (2009). The dashed line that links the genetic portion of the diagram with the environment indicates that analyses are needed to identify traits whose QTLs are constant across environments.



**(A)****(B)****(C)**

# CLIMATE

# CROP

Climate projections from GCMs

Calibration, bias correction

Delta method    Weather typing    Nudging

Downscaling

RCM    Delta method(s)    Weather typing    Nudging  
Weather generator    Statistical    Neural networks

Statistical models

Time series    Panel    Cross-section

Niche-based models

AEZ    EcoCrop    SDMs

Large-area process-based models

GLAM    MCWLA    PRYSBI  
Adjusted field-scale model

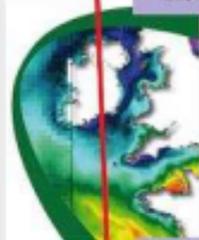
Field-scale process-based models

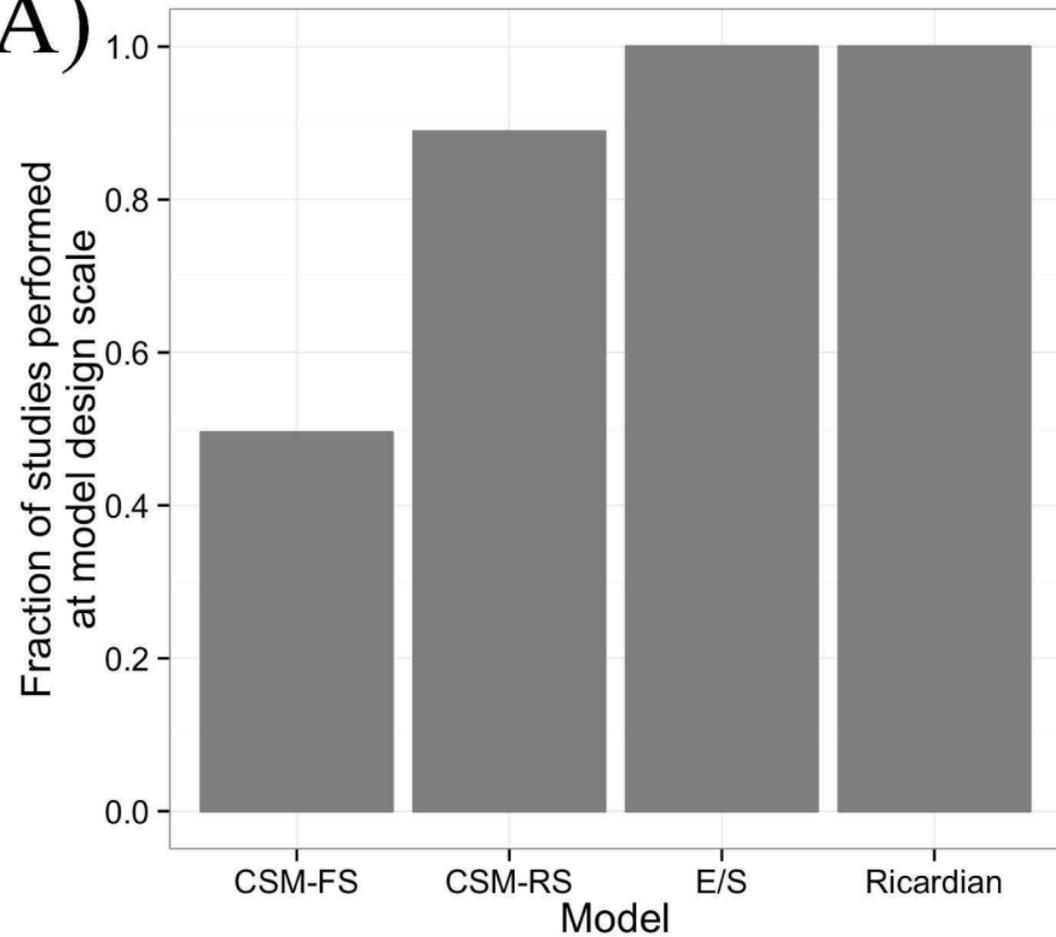
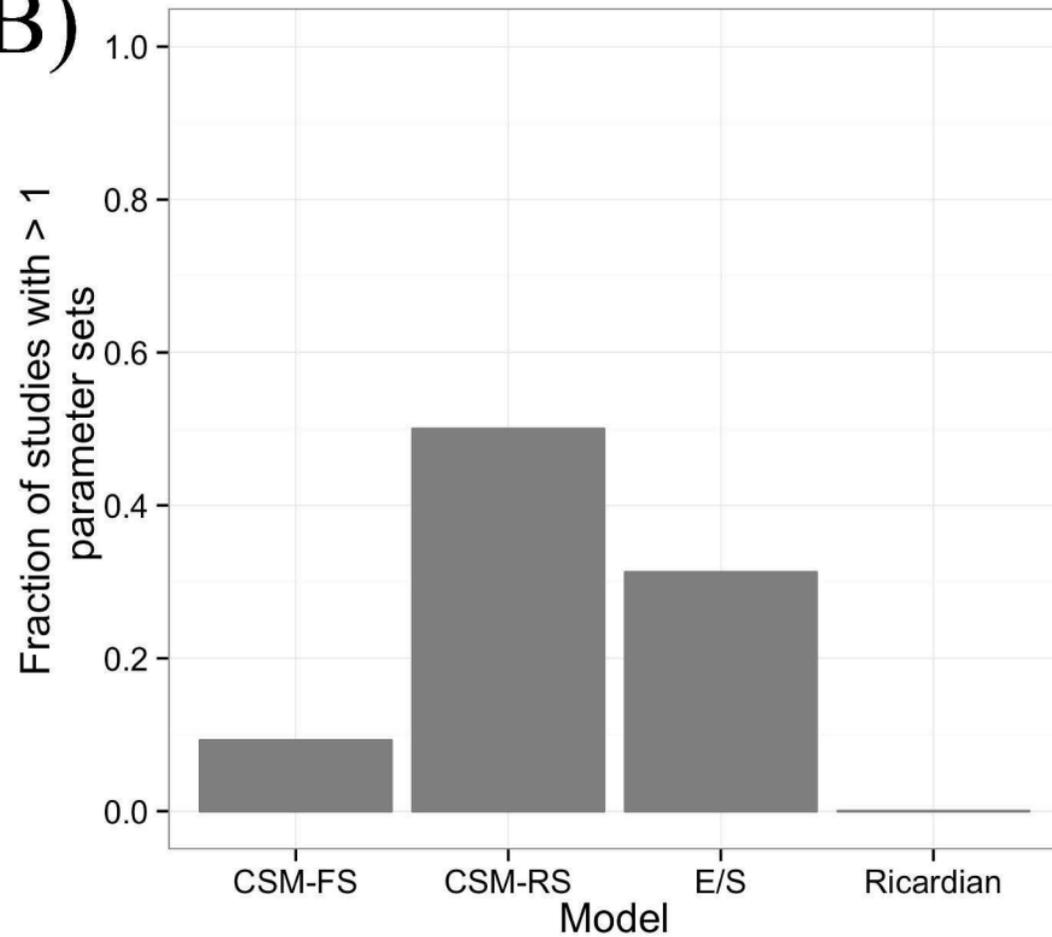
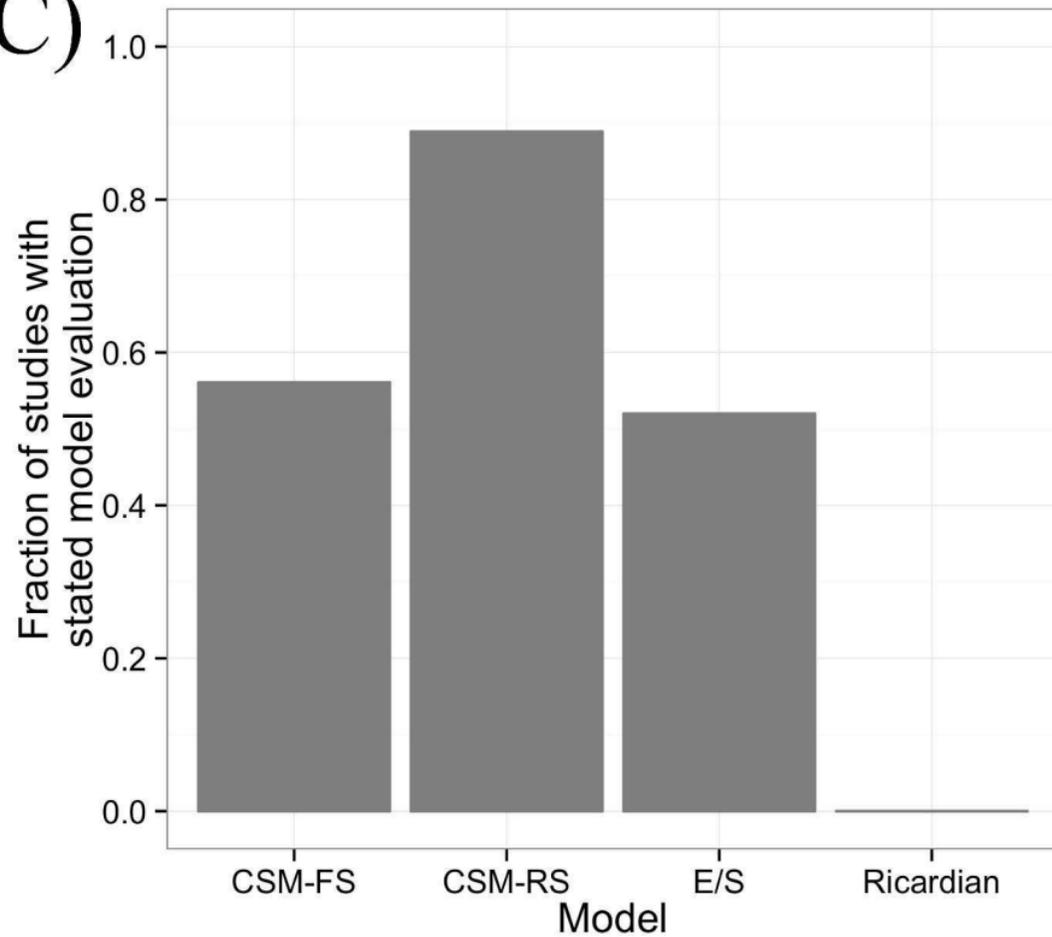
100+ models

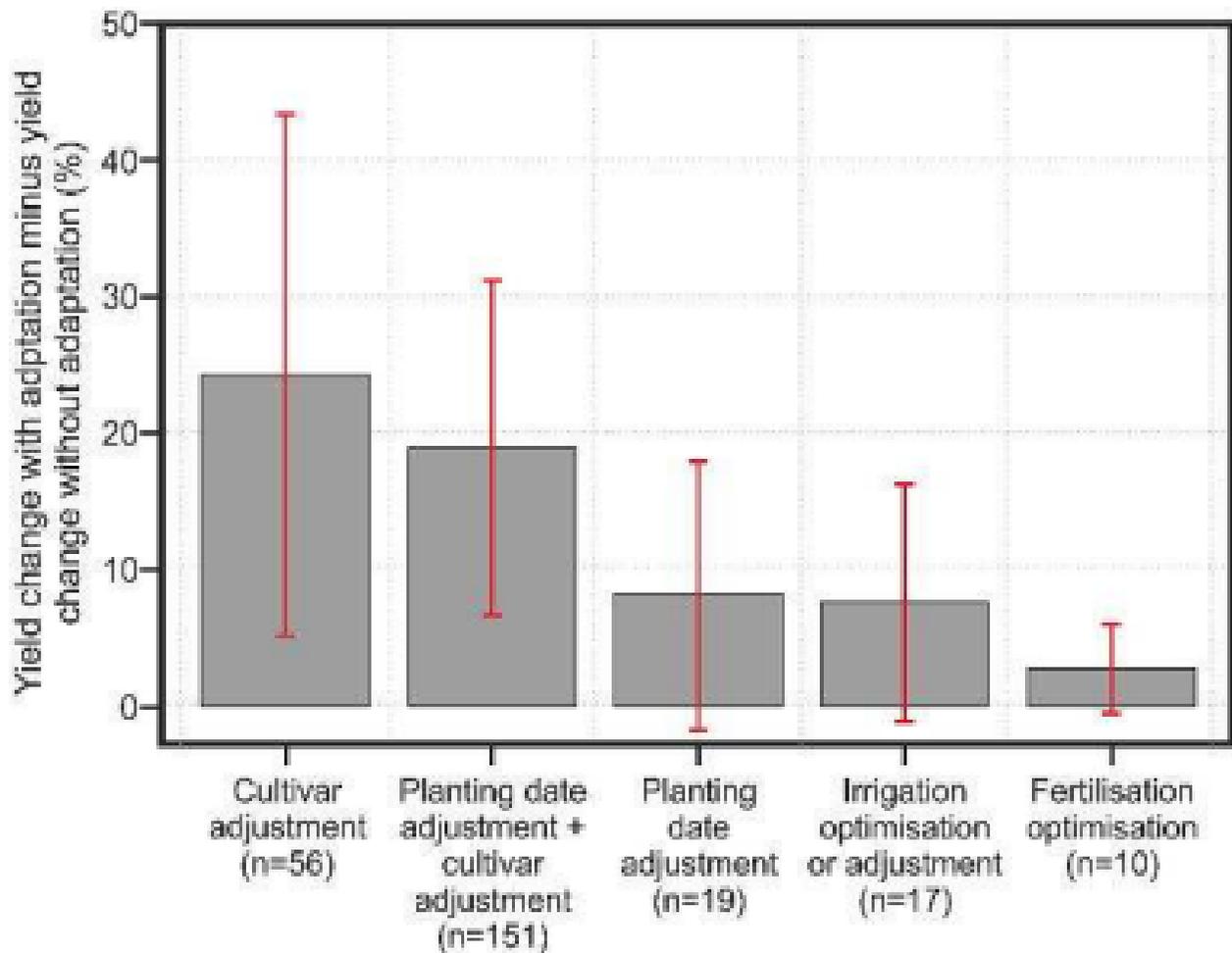


Increased uncertainty

Effective adaptation strategies



**(A)****(B)****(C)**





Benefit from genotypic adaptation (%)

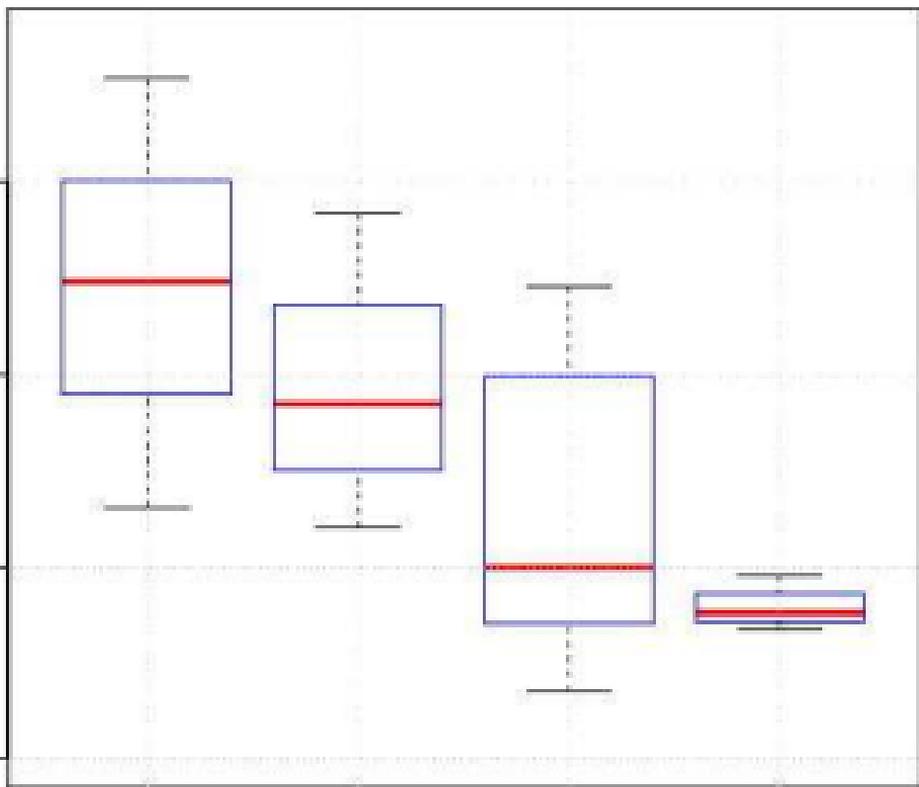
60  
40  
20  
0

wheat

sorghum

groundnut

maize



## Genetics

Parental  
QTL data

Genetic  
model

Population  
parameter values  
(i.e. parameterised  
recombinants)

## Ensemble crop simulation

Process-based  
crop model(s)

+

Coupled  
parameterisation(s)  
(e.g. leaf  
elongation)

Yield and  
components

## Environment

Soil

Weather

Atmospheric  
constituents

Management

