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Incorporation of strontium in earthworm-secreted calcium carbonate granules  
produced in strontium-amended and strontium-bearing soil

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Running head: Sr in earthworm granules

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## Abstract

This paper investigates the incorporation of Sr into biomineralized calcium carbonate granules secreted by the earthworm *Lumbricus terrestris*. Experiments were conducted using an agricultural soil amended with  $\text{Sr}(\text{NO}_3)_2$  to give concentrations in the range 50 - 500  $\text{mg kg}^{-1}$  Sr and a naturally Sr-rich, Celestine-bearing soil containing up to 11 000  $\text{mg kg}^{-1}$  Sr. Granule production rates were in the range 0.26 – 2.3  $\text{mg}_{\text{CaCO}_3} \text{earthworm}^{-1} \text{day}^{-1}$ ; they showed no relationship with soil or soil solution Sr concentration but decreased with decreasing pH. Strong relationships exist ( $r^2 \geq 0.8$ ,  $p \leq 0.01$ ) between the Sr concentrations and Sr / Ca ratios of the granules and those of the soil, soil solution and earthworms. The highest bulk Sr concentration we recorded in the calcium carbonate granules was 5.1 wt% Sr whilst electron microprobe analysis recorded spot concentrations of up to 4.3 wt % Sr. X-ray diffraction and X-ray absorption spectroscopy indicate that the majority of the calcium carbonate is present as Sr-bearing calcite with trace amounts of Sr-bearing vaterite also being present. The granules produced in the Sr-amended soils concentrated Sr relative to Ca from the bulk soil and the earthworms. This suggests that earthworm secreted calcium carbonate may be significant in the cycling of  $^{90}\text{Sr}$  released into soils via nuclear accidents or leakage from nuclear waste storage facilities.

## 1. INTRODUCTION

The incorporation of Sr (and other elements) into calcite and other calcium carbonate phases has been, and continues to be, the subject of much research in the geochemical community (e.g. Bracco et al., 2012; Tertre et al., 2012; DePaolo, 2011; Tang et al., 2008; Lakshtanov and Stipp, 2007; Nehrke et al., 2007; Finch and Allison, 2007; Gabitov and Watson, 2006; Elzinga and Reeder, 2002; Fujita et al., 2004; Parkman et al., 1998; Pingitore et al., 1992; Stipp and Hochella, 1991; Tesoriero and Pankow, 1996). Some of the research is driven by the potential for calcite to immobilise contaminants, particularly  $^{90}\text{Sr}$  (e.g. Riley et

al., 1992; Achal et al. 2012; Perdrial et al., 2011; Spycher et al., 2009; Barkouki et al., 2011; Fujita et al., 2010; Tertre et al., 2012). Additionally, the potential use of Sr/Ca ratios in biominerals and speleothems for the reconstruction of past environments is also the subject of numerous studies (e.g. Finch and Allison, 2007; Wassenburg et al., 2012; Sinclair et al., 2012; Bluszcz et al., 2009; Fairchild et al., 2000; Dissard et al., 2010a, 2010b; Stoll et al., 2002). This paper is concerned with Sr partitioning into earthworm-secreted calcite; understanding this system may have important implications for site remediation and environmental reconstruction.

Earthworms are perhaps best known for their role in the breakdown of organic material and the mixing and aeration of soils (Edwards, 2004; Edwards and Bohlen, 1996). However, many species of earthworm also synthesise calcium carbonate (Canti and Pearce, 2003). The calcium carbonate is produced in the calciferous glands as micron-scale spherites which, in many species, go on to coalesce and form millimetre scale granules comprised predominantly of calcite, but also containing aragonite, vaterite and amorphous calcium carbonate (e.g. Canti and Pearce, 2003; Gago-Duport et al., 2008; Lee et al., 2008). The spherites and granules are secreted into the earthworm intestine and, from there, into the soil. Carbon isotope studies of *L. terrestris* granules indicate that the carbon in the granules comes from both carbon dioxide and consumed organic matter (Briones et al., 2008; Canti, 2009). Granule production rates by the earthworm *Lumbricus terrestris* have been linked to soil pH (Lambkin et al., 2011) but the function of the granules is still not clear with suggested functions including Ca, CO<sub>2</sub> and pH regulation in body tissues and fluids (Darwin, 1881, Robertson, 1936; Pearce, 1972).

It is well established that earthworms can accumulate metals when exposed to contaminated or amended soils (e.g. Nahmani et al., 2007). We have shown (Fraser et al., 2011) that, at least for Pb, metals can also accumulate in the calcium carbonate granules secreted by earthworms in those soils; Pb was both structurally incorporated within the calcite in the granules and also present as the Pb-carbonate cerrusite. The Pb-enriched granules contained an unexpectedly large amount of aragonite, consistent with studies

which highlight the influence of various trace elements on calcium carbonate polymorph transformations and stability (e.g., Sr, Mg, Zn,  $\text{SO}_4^{2-}$ ) (Bots et al., 2011; Finch and Allison, 2007; Morse et al., 1997; Reis et al., 2008; Rodriguez-Blanco et al., 2011a; Wang et al., 2012). Morgan et al. (2001, 2002) demonstrated that despite the chemical similarities between Sr and Ca and the accumulation of Sr by earthworms exposed to Sr-rich soils, earthworms are able to metabolically differentiate between Sr and Ca. Studies have shown that the distribution coefficients for Sr incorporation in biosynthesised calcite are up to an order of magnitude higher than values reported in natural and synthetic calcite (e.g. Fujita et al., 2004; Morgan et al., 2001). Strontium 90 is produced by nuclear fission and is a significant component of nuclear waste. It has been accidentally released into numerous environments due to leaks from storage facilities (e.g. Hanford, USA, Thompson et al., 2010; Oak Ridge, USA, Gu et al., 2005; Mayak, Russia, Standring et al., 2002 and Sellafield, UK, Gray et al., 1995) and nuclear accidents (e.g. Fukushima Daiichi). Evidence suggests that the partitioning of Sr into calcite (and other calcium carbonates) is related to environmental variables such as temperature and  $\text{CO}_2$  levels; consequently Sr/Ca ratios can be used as a tool for reconstruction of past environments (Fairchild et al., 2000; Dissard et al., 2010a, b).

The aims of the current study were therefore to determine: 1) the extent to which Sr would accumulate in the calcium carbonate produced by *L. terrestris* exposed to Sr-enriched soils, 2) whether distribution coefficients for this incorporation were consistent with those determined for inorganic systems, and 3) whether Sr incorporation impacted on granule mineralogy. Our results are placed in the context of the potential of earthworm calcium carbonate granules within the fields of contaminant immobilisation and environmental reconstruction.

## 2. METHODS

### 2.1. Earthworms and soils

Clitellate *Lumbricus terrestris* were obtained from Recycle Works Ltd. (Ribchester, PR3 3XJ, UK). They were kept for one week in a moist mixture of 1:2 by volume peat soil and Kettering Loam (Boughton Loam and Turf Management, Kettering, Northamptonshire, NN16 8UN, UK) prior to being used in the experiments.

Three arable soils were collected for the study, one, Hamble soil (HS) from near Theale, Berkshire OS 164 (SU-618-702) and two Yate soils (Yate Soil High, YSH, and Yate Soil Low, YSL) from the former celestine ( $\text{SrSO}_4$ ) mining area of Yate, Bristol OS 172 (ST-712-847) (Nickless et al., 1976).

## 2.2. Soil characterization

Prior to characterisation and experiment the soils were oven dried (at 40°C) and sieved to < 250 µm. Subsamples of the soil were dried at 105 °C to remove moisture completely and all results are expressed on a per mass of 105 °C dried soil basis. Selected soil properties are reported in Table 1. Soil water holding capacity (WHC) was determined gravimetrically following ISO 11465:1993 (ISO, 1993). Soil pH in deionised water was determined following BS ISO 10390:2005 (BSI 2005). Loss on ignition (LOI) was determined following BS EN 15935 (BSI 2009) and used as a proxy for organic matter content

Soil elemental composition was determined by aqua regia digest following BS ISO 12914 (BSI, 2010) using an Anton Parr Multiwave 3000 microwave followed by analysis using a Perkin Elmer Optima 7300 DV inductively coupled plasma – optical emission spectrometer ICP-OES). For quality control, an internal reference soil material (SS50) traceable to BCR-143R (Commission of the European Communities, Community Bureau of Reference) and blanks were digested in triplicate. Recoveries were 93% for Ca and 107% for Sr for SS50; repeated analysis of individual samples indicated a precision  $\leq 0.5\%$  and detection limits of less than 0.07 mg kg<sup>-1</sup>.

## 2.3. Earthworm incubation experiments

Incubation experiments were carried out on the Hamble and Yate soils in a temperature and ventilation controlled Memmert ICP 600 incubator set at 16 °C with minimal ventilation. For the Hamble soil, incubations were carried out on unamended soil (HS) and also Hamble soil to which solutions of anhydrous  $\text{Sr}(\text{NO}_3)_2$  (Sigma-Aldrich, CAS 10042-76-9) were added to give initial target Sr concentrations in soil of 50, 100, 150 and 500  $\text{mg Sr kg}^{-1}$  (denoted HS50, HS100, HS150 and HS500, respectively). Higher Sr concentrations (1000 and 4000  $\text{mg kg}^{-1}$ ) were also used but proved terminal to the earthworms and are not reported here. Amended soils were digested in aqua regia and analysed by ICP-OES to check concentrations. Incubation experiments were carried out in plastic containers enclosed in perforated plastic bags. Each container held 300 mg oven dried soil mixed with either deionised water or  $\text{Sr}(\text{NO}_3)_2$  solution to give a water content of c. 60% of the WHC. An individual *L. terrestris* was weighed and added into each soil sample. Five replicates were run for each concentration.

After 28 days earthworms were removed from the soil, depurated for 48 hours (Arnold and Hodson, 2007), digested by aqua regia and analysed for Sr and Ca by ICP-OES. Detection limits and precision were as reported for the soil digestions. Soil pore water was extracted overnight with 100 mm epoxy bodied MOM Rhizon samplers. pH was measured with a Jenway 3510 pH meter; precision was 0.36 %. Solutions were then acidified with 5% concentrated ( $\geq 69\%$ ) Sigma-Aldrich nitric acid ACS reagent grade, ISO $\geq 69\%$  (CAS 7697-37-2) prior to analysis by ICP-OES for Ca and Sr. An in house 500  $\mu\text{g kg}^{-1}$  standard gave recoveries of 90 – 110 %. Detection limits were  $\leq 0.017 \text{ mg kg}^{-1}$  and precision was < 2.2%. Soil sub-samples were taken from each incubation for pH measurement (BSI, 2005). The remaining soil was sieved to 500  $\mu\text{m}$  to recover the granules. Granules from each replicate treatment were dried and pooled for weighing to determine production rate expressed as  $\text{mg CaCO}_3/\text{g worm/day}$  prior to further characterization.

Two additional incubation experiments were carried out. In one, Hamble soil amended to 500  $\text{mg Sr kg}^{-1}$  was incubated but without any earthworm additions. No granules were recovered from this incubation supporting the earthworm-derived origin of the granules.

In the second, granules recovered from our unamended Hamble soil experiment were added to Hamble soil amended to 500 mg Sr kg<sup>-1</sup> using Sr(NO<sub>3</sub>)<sub>2</sub>. After 28 days the granules were characterised for chemical composition as described in Section 2.4.

## 2.4. Granule characterization

The majority of granules were dissolved in 5% nitric acid and analysed by ICP-OES. The certified reference material dolomite BCS No386 was also digested following this method and gave a recovery of 117% for Ca. Detection limits were  $\leq 0.028$  mg kg<sup>-1</sup> with precision of  $\leq 1.3$  %. Remaining granules were used in mineralogical and spatially resolved chemical investigations.

Thin sections of the granules were produced by embedding the granules in EpoFIX (Struers) resin and grinding to a thickness of 50-70  $\mu$ m, that is, 25-35  $\mu$ m either side of the granule centre. The granule slices were then mounted on Chance Glass Ltd. glass slides and mechanically polished using a 1  $\mu$ m particle size corundum slurry.

Bulk granule mineralogy was assessed by combining information gleaned from X-ray powder diffraction (XRD-NHM) on powdered granules and in-house, non-destructive X-ray microdiffraction ( $\mu$ XRD-NHM) on the polished granule sections that were then used for electron probe analysis and X-ray absorption spectroscopy. XRD-NHM data were collected in reflection geometry using a Nonius PDS 120 powder diffraction system consisting of an Inel curved, position sensitive detector (PSD) within a static beam-sample-detector geometry. This system allows the simultaneous measurement of the diffracted X-ray intensities at all angles of  $2\theta$  across 120° (Schofield et al., 2002). Cobalt K $\alpha_1$  radiation was selected from the primary beam by a germanium 111 crystal monochromator with the X-ray tube operating at 35 kV and 30 mA. Horizontal and vertical slits restricted the beam to a height of 0.24 mm and width of 4.0 mm. Individual granules were powdered in an agate pestle and mortar, mixed with acetone and thinly deposited on a circular sapphire substrate. NIST silicon powder SRM640 and silver behenate were used as external standards;



calibration and data collection were performed using in house software Diffgrab™. Data were collected for a minimum of 2000 s with samples spinning continuously in the plane of the sample surface and with the sample surface at an angle of 4.0° to the incident beam.  $\mu$ XRD-NHM data were collected using a Nonius PDS 120 powder diffraction system as described above. In this case a 100  $\mu$ m diameter beam was selected by a pinhole from a 300  $\mu$ m diameter primary beam of Cu K $\alpha$  radiation generated by a GeniX system with a Xenocs FOX2D CU 10\_30P mirror operating at 50 kV and 1 mA (Lambiv Dzemua et al., 2012). Measurements were made in reflection geometry. The surface of the granule polished section was brought into the focal point of the beam using a Zeiss Axio Cam MRc5 CCD optical system. The footprint of the beam on the sample was 750-500  $\times$  100  $\mu$ m. During data collection, the polished sections were spun continuously in the plane of the sample surface. NIST silicon powder SRM640 and silver behenate were used as external standards; calibration and data collection were performed using Diffgrab™ with data collection times of at least 3000 s.

Elemental distribution within the granules was mapped using both electron probe microanalysis (EPMA) and synchrotron based X-ray fluorescence (sXRF). EPMA element maps were generated using a Cameca SX100 electron microprobe operating at 15 kV and 100 nA with the beam set to a spot size of 1  $\mu$ m. Wavelength dispersive spectrometers (WDS) were used to detect elements Ca, Sr, Mn, Mg and Fe while the elements Na, Al, Si, P, S, Cl, K and Ti were detected using an energy dispersive spectrometer (EDS). Maps were 512  $\times$  512 pixels with step sizes of 4-5  $\mu$ m and dwell times of 180 – 200 ms.

Quantitative electron microprobe chemical analyses of the granules were performed using a WDS Cameca SX100 microprobe operating with 10 kV accelerating voltage, 100 nA beam current, and a spot size of  $\sim$ 20  $\mu$ m. All elements were analysed using WDS and the probe standards used were: calcite for Ca, celestine for Sr and S, MnTiO<sub>3</sub> for Mn, forsterite for Mg, fayalite for Fe, jadeite for Na, corundum for Al, KBr for K, and wollastonite for Si, ScPO<sub>4</sub> for P. The X-ray intensities were corrected using a standard PAP correction

procedure. Between 20 and 40 points were analysed on each granule along rim-core-rim line profiles. The atomistic detection limits for Ca, Mg, Sr and Mn were 0.05, 0.02, 0.03 and 0.04 wt %, respectively, and the wt % oxide standard deviations for CaO, MgO, SrO and MnO were 0.5, 0.02, 0.03 and 0.03, respectively.

The sXRF was performed on the microfocus beamline I18 at the Diamond Light Source (Mosselmans et al., 2009) where sXRF maps were collected using a 9-element Ge detector with the Si(111) cryogenically-cooled monochromator set to provide an incident X-ray energy of 16500 eV. The beam-on-sample size was  $5 \times 5 \mu\text{m}$ . and maps were collected with  $30 \mu\text{m}$  steps. XRF data were processed in PyMCA 4.4.1 (Solé et al., 2007). sXRF maps were used principally to determine suitable points for microfocus X-ray absorption spectroscopy.

#### **2.4.1. Micro X-ray Absorption Spectroscopy ( $\mu\text{XAS}$ )**

$\mu\text{XAS}$  was carried out on the thin sections of the granules using the microfocus beamline I18 at the Diamond Light Source.  $\mu\text{XAS}$  was carried out on one granule each extracted from HS150, HS500, YSL and YSH and two granules extracted from HS100. For most of the granules two Ca K-edge XANES and Sr K-edge EXAFS spectra were collected from the same point of interest at several different points. Recurrent spectra were compared to check for any sign of beam damage. For all the spectra obtained no changes were seen in these recurrent spectra. XAS data were processed in Athena (Ravel and Newville, 2005) and Pyspline (Tenderholt and Quinn, 2009) and fitted using DL\_Excurv (Tomic et al., 2004).

Sr K-edge spectra of relevant standards were recorded as follows: celestine (collected from the Yate soil),  $\text{SrCO}_3$  (Fisher Scientific), and  $\text{Sr}(\text{NO}_3)_2$  (Fisher Scientific) spectra were obtained in transmission mode using samples ground together with boron nitride and pressed into pellets. Fluorescence data were collected from Sr-containing aragonite (this sample is speleothem aragonite from Makapansgat Valley, South Africa and was provided by Dr. A. Finch, University of St Andrews), calcite with Sr adsorbed onto it,

earthworm-produced calcium carbonate granules with Sr adsorbed onto them and vaterite co-precipitated with Sr.

Calcite for the Sr adsorbed standard was synthesized following the method of Rodriguez-Blanco et al. (2011b). The powder produced had a BET surface area of  $0.99 \text{ m}^2 \text{ g}^{-1}$  whilst the granules had a BET surface area of  $0.83 \text{ m}^2 \text{ g}^{-1}$ . One gram of either the synthesised calcite or calcium carbonate granules recovered from our unamended Hamble soil experiment was equilibrated in 50 mL of a pH 7.5 solution of  $\text{NaHCO}_3$  and HCl at  $20^\circ \text{C}$  for 24 hours, the pH adjusted back to 7.5 and then Sr added as  $\text{Sr}(\text{NO}_3)_2$  (Sigma-Aldrich, CAS 10042-76-9) to give a solution concentration of 100 nM Sr per g calcite. After 24 hours the adsorbent was collected via centrifugation.

Vaterite co-precipitated with Sr was made following the method of Bots (2012). A solution of 100 mM  $\text{CaCl}_2$  was rigorously mixed with a solution of 50 mM  $\text{Na}_2\text{CO}_3$  and 1.25 M  $\text{Na}_2\text{SO}_4$  on a magnetic stirring plate. Sr was added to the  $\text{CaCl}_2$  solution as  $\text{Sr}(\text{NO}_3)_2$  to give a concentration of 100  $\mu\text{M}$  Sr. Vaterite precipitated instantly, was washed by filtration with deionised water to remove sulphate and dried in isopropanol. The precipitate was shown to be pure vaterite using XRD.

An attempt was made to synthesize standards of calcite co-precipitated with Sr following the method of Gruzensky (1967). Solutions of  $\text{CaCl}_2$  with  $\text{SrCl}_2$  were prepared to give final Sr concentrations in solution of 100  $\mu\text{M}$ , 1  $\mu\text{M}$  and 10 mM Sr. In each case however the resulting precipitate was a mixture. For solutions of 100  $\mu\text{M}$ , and 1  $\mu\text{M}$  Sr the calcite was precipitated with vaterite while the solution of 10 mM Sr produced calcite with Ca-bearing strontianite and aragonite.

Ca K-edge XANES data were collected in fluorescence mode from the speleothem aragonite and also synthetic vaterite stabilized with 4% sulphate (provided by Dr. P. Bots, University of Leeds). As the Ca XANES data were collected in fluorescence mode the spectra are distorted by self-absorption. Corrections can, in principle, be made for self absorption and a basic example of such a correction is shown for Ca K-edge XANES from a calcite single crystal in the Supplementary Information. In order to make an accurate self-

absorption correction, a knowledge of the density of the sample is required. As the granules of this study have a mineralogy that may vary within the volume of sample analysed at each point, this is not a feasible option. Consequently, no corrections have been applied to the Ca K-edge XANES from the granules in this study, however, as all the spectra were recorded in the same way comparisons between the spectra can be made.

## **2.5. Statistics**

Statistical analysis was carried out using Sigma Stat 3.0.1 by SPSS. All data were checked for normality using the Kolmogorov-Smirnov test before analysis and appropriate parametric or non-parametric statistics used. Soil Sr concentrations were compared using Kruskal-Wallis one way analysis of variance on ranks, changes in earthworm weight during the course of the experiment by one way analysis of variance. Pearson and Spearman's rank correlations were determined as appropriate for relationships between soil, soil solution, earthworm and granule chemistry and granule production.

## **3. RESULTS AND DISCUSSIONS**

### **3.1. Soil solution**

Sr concentrations in the amended Hamble soils varied about the target values potentially due to uncertainties in extraction efficiency and analysis. However, importantly for the present study soils with significantly different Sr concentrations were produced (Table 1,  $p \leq 0.01$ ). The low concentrations of Sr in the soil solution in the YSL and YSH soils relative to the Sr amended Hamble soils (Table 1) reflects the sources of Sr in the soils, celestine in the Yate soils (Nickless et al., 1976) and the more soluble  $\text{Sr}(\text{NO}_3)_2$  in the Sr-amended Hamble soil.

### **3.2. Earthworm survival and chemistry**

No earthworms died over the duration of the experiment but they all lost weight. Average weight loss was  $6.4 \pm 11.8$  % of their body weight ( $n = 6 \pm \text{s.d.}$ ). There were no significant differences in weight loss between soils or treatments ( $p \geq 0.05$ ). Earthworm chemistry is summarised in Table 2. The range of Sr concentrations is similar to that reported for different species kept in soils collected from the Yate region by Morgan et al. (2001, 2002) but Ca concentrations are up to an order of magnitude higher. This appears to reflect the higher Ca concentrations in our soils compared to those in Morgan's study ( $1250 - 5540 \text{ mg kg}^{-1}$ ). Additionally Morgan et al. (2001) studied different species of earthworms, three of which, *Aporrectodea caliginosa*, *Aporrectodea longa* and *Allolobophora chlorotica*, have less well developed calciferous glands which produce far fewer granules (Canti and Pearce, 2003) which may impact on Ca accumulation. The concentration of Sr accumulated by earthworms, in this study, increased with the concentration of Sr in the soil ( $r^2 = 0.92$ ,  $p \leq 0.05$ ) and the soil solution ( $r^2 = 0.88$ ,  $p \leq 0.05$ ), but no correlation was found for Ca supporting the conclusion of Morgan et al. (2001) that Ca uptake is regulated whilst Sr uptake is not.

Similarly to Morgan et al. (2001) concentration factors calculated for earthworm body loads using the bulk soil concentrations for Sr and Ca were  $0.89 \pm 0.42$  and  $1.14 \pm 0.45$  (mean  $\pm$  s.d.,  $n = 32$ ), respectively. Distribution coefficients (D) for Sr and Ca were calculated as the ratio of the Sr/Ca in the earthworm and Sr/Ca in the soil or soil solution and are given in Table 3. On average values for earthworm distribution coefficients are  $0.81 \pm 0.31$  and  $0.51 \pm 0.26$  (mean  $\pm$  s.d.,  $n = 32$ ) for partitioning between the earthworm and soil and the earthworm and soil solution, respectively, indicating that if uptake and accumulation is from the bulk soil there is no discrimination between Sr and Ca ( $D \cong 1$ ) whereas if uptake is from the metal in solution Ca is preferentially taken up ( $D < 1$ ). For the earthworm – soil distribution coefficients the unamended Hamble soil and the YSL soil have relatively low values presumably reflecting the low Sr content of the unamended Hamble soil and the non-bioavailable nature of the Sr in the YSL soil, respectively.

The earthworm – soil solution partition coefficient for YSH is higher than the other values, possibly reflecting the relatively high Sr / Ca ratio of the YSH soil solution. Unlike the findings of Morgan et al. (2001) there is no indication that Sr bioaccumulation decreases relative to Ca at higher soil Ca concentrations. This may reflect the narrower range of Sr and Ca concentrations in our study and the fact that the low Sr concentration soils were amended with highly soluble (and therefore more bioavailable)  $\text{Sr}(\text{NO}_3)_2$  whilst the high Sr concentration soils contained the less soluble (and therefore less bioavailable) celestine.

### 3.3. Granule production, bulk mineralogy and bulk chemistry

Granule production rates (Table 2) were similar to those reported by Fraser et al. (2011) in Pb amended artificial soils and by Lambkin et al. (2011) in agricultural soils . There were no significant correlations between the Sr concentration of the soil or soil solution and granule production rate. The lower production rate recorded for soil YSL is consistent with the reduction in granule production with decreasing pH reported by Lambkin et al. (2011).

Analysis of bulk and micro XRD data shows that calcite is the main component of the granules with vaterite often present as well (Table 4). Trace amounts of quartz were identified in all the granules but no Sr-carbonate or Sr-sulphate phases such as strontianite, carbocernaite or celestine were identified. The quartz is potentially incorporated within granules during their transportation from the calciferous glands into oesophageal pouches where the granules are stored before being excreted and where granules growth may still occur (Lee et al., 2008).

The bulk concentrations of Sr and Ca in the granules as measured by ICP-OES are reported in Table 2, where it can be seen that substantial levels of Sr are incorporated into the calcium carbonate granules. The concentrations of Sr reported in the granules are high but similar values have been reported in the literature. Concentrations of Sr in inorganically

produced calcite reported in the literature include 8505 mg kg<sup>-1</sup> (Pingitore et al., 1992), 1477 mg kg<sup>-1</sup> (Tang et al., 2008) and 1300 – 3500 mg kg<sup>-1</sup> (Gabitov and Watson, 2006). Concentrations of Sr in biogenic calcite are often higher, with reported concentrations including 27 000 mg kg<sup>-1</sup> in the common groundwater, gram-positive bacteria, *Bacillus pasteurii*, (Warren et al., 2001, with vaterite also present) and up to 5000 mg kg<sup>-1</sup> in decapods (Veizer, 1983). EPMA of the granules confirmed the high Sr concentrations within the granules (Table 4). For granules produced in the Hamble soils the granules with the highest SrO levels (up to 5 wt% SrO or 4.3 wt % Sr) are those produced in the soils amended to the highest concentration of Sr. The high concentrations of Sr may in part reflect the crystallisation history of the predominantly calcite granules given that initially the calcium carbonate is amorphous (e.g. Gago-Duport et al. 2008) and that amorphous calcium carbonate can be preserved in the granules (e.g. Lee et al. 2008). Calcite that forms via amorphous calcium carbonate can have elevated Mg concentrations (e.g. Radha et al., 2012; Raz et al, 2000; Wang et al. 2012) and it is possible that similar effects occur for Sr though we are not aware of any published studies on this subject. It should also be noted that the bulk Mg concentrations in the granules are low ( $\leq 31$  mg kg<sup>-1</sup>) and even narrow bands of high Mg concentration (Table 4) are below the levels found in many biogenic calcites or thought to represent maxima for calcite formed directly from solution rather than from an amorphous precursor (e.g. Berner, 1975, Fernández-Díaz et al., 1996; Loste et al., 2003)

In order to assess the potential for Sr adsorbing to granule surfaces post secretion/excretion granules produced in unamended, Sr-free Hamble soil were placed in Hamble soil amended to 500 mg Sr kg<sup>-1</sup> for 28 days. Subsequent ICP-OES analyses revealed that Sr levels associated with the granules increased from 345 mg kg<sup>-1</sup> in the control granules (Table 2) to 1370 mg kg<sup>-1</sup>. In contrast Sr levels in granules produced by earthworms in Hamble soil amended to 500 mg Sr kg<sup>-1</sup> were 34200 mg kg<sup>-1</sup> (Table 2). This

suggests that if Sr adsorption from the soil solution to the granule surface occurs then it only accounts for a small fraction of the total Sr associated with the granules.

As described in detail by Fraser et al. (2011), in X-ray diffraction the calcite 104 peak position is a good indicator of relative changes in the size of the calcite unit cell. Calcite 104 peak positions taken from bulk XRD data are reported in Table 4. In order to ensure that these XRD measurements are compared with appropriate Sr levels in granule calcite, the wt% SrO of each granule was estimated from the average wt% SrO value from EPMA point analyses within a rim-core line profile of large calcite crystals making up the granules (Table 4) rather than using the Sr concentrations measured by ICP-OES (Table 2) which are average values for a range of granules and which will therefore include Sr present in vaterite; however the trends observed are the same regardless of which Sr data are used. Figure 1 shows the calcite 104 peak position as a function of the estimated average wt% SrO from EPMA analyses for the granules produced by earthworms in Hamble soils (and not those produced in Yate soils for which there is no “control” specimen, i.e. granules produced in an equivalent soil but with the absence of Sr). It can be seen that the calcite 104 peak shifts to lower  $2\theta$  values as the Sr concentration in the granule calcite increases. A shift of the calcite 104 peak to lower  $2\theta$  values is indicative of an increasing unit cell size suggesting that Sr, which is larger than Ca, is structurally incorporated by the calcite.

No aragonite was identified within any of the granules of this study, although the study of Fraser et al. (2011) found granules comprising calcite and aragonite with no vaterite being present. The current study used a natural soil amended with Sr and a natural Sr-rich soil, whereas in the study of Fraser et al. (2011) the biogenic calcium carbonate granules were produced by *L. terrestris* in artificial soil amended with Pb. This suggests that the mineralogy of the calcium carbonate granules may be influenced by soil, consistent with experiments on the inorganic calcium carbonate system (e.g. Bots et al., 2011; Finch and Allison, 2007; Rodriguez-Blanco et al., 2011a). The production of mixtures of different



phases when we attempted to co-precipitate calcite with Sr using different concentrations of Sr in solution to produce standards also supports this suggestion.

The Sr concentrations and the Sr/Ca ratio of the granules are strongly related ( $r^2 \geq 0.8$ ,  $p \leq 0.01$ ) to those of the soil, the soil solution and the earthworms. Distribution coefficients (Table 3) suggest differences in the partitioning of Sr and Ca in the granules between the Sr amended and naturally Sr-rich soils. The partitioning of Sr and Ca from the soil and soil solution is mediated by the earthworm metabolic processes. As such the distribution coefficients reported here cannot be fairly compared with distribution coefficients for Sr and Ca partitioning in inorganic calcite precipitated from solution. However, the values obtained are similar to that of 0.49 obtained for biogenic calcite produced by the bacterium *Bacillus pasteurii*, (Fujita et al., 2004) and greater than many obtained for inorganic calcite (e.g.  $0.021 \pm 0.003$  by Tang et al., (2008) and up to 0.140 by Tesoriero and Pankow, (1996)) suggesting that earthworms, like bacteria (Warren et al., 2001), can more efficiently partition Sr into calcite than inorganic processes. Indeed the distribution coefficients for partitioning between the granules and bulk soils suggest that, for the Sr-amended soils, the granules may preferentially concentrate Sr with the granule – earthworm partition coefficients for these soils also suggesting the granules concentrate Sr relative to Ca compared to the earthworm tissues. This finding is consistent with that of Morgan (1981) who found that Sr injected directly into the coelomic cavity of *L. terrestris* in the form of  $\text{SrCl}_2$  solution was detected in the calciferous glands and spherites within the glands but not in the chloragogenous tissue, an organ associated with the accumulation and metabolism of Ca.

The granule-soil and granule-soil solution partition coefficients are similar to those for the earthworm-soil and earthworm–soil solution partition coefficients for the amended soils but are substantially lower for the YSL and YSH soils. The granules have a relatively lower Sr/Ca ratio than the earthworms in the YSL and YSH soils compared to the Sr amended soils. This suggests that the accumulated Sr from the YSL and YSH soils is somehow transported or metabolised differently to that accumulated from the amended soils or has a

different availability due to the soil chemistry. This would be expected given the different forms of Sr present in the soils and is supported by the granule–earthworm partition coefficients which show more partitioning of Sr relative to Ca in the granules compared to the earthworms for the Sr amended soils. Despite the strong correlation between granule and earthworm Sr, the granule-earthworm partition coefficients suggest that after Sr is accumulated in the earthworm the partitioning of Sr into granules via the calciferous gland is not a straight forward process dependent solely on Sr concentrations. For example the distribution coefficient decreases with increasing earthworm Sr/Ca ( $r^2 = 0.69$ ,  $p \leq 0.05$ ) perhaps suggesting that at higher Sr/Ca ratios there is preferential excretion of Sr preventing its partitioning into the granules or that the transport path of Sr to the granules is saturated (Chwodhury et al., 2000; Dodd, 1967). Alternatively this may reflect a precipitation rate effect. In inorganic systems higher rates of calcite precipitation result in greater partitioning of Sr into calcite (e.g. Nehrke et al., 2007; Tang et al., 2008; Tesoriero and Pankow, 1996; Gabitov and Watson, 2006). Granule production rates are lower in the YSL and YSH soils than in the HS soils perhaps suggesting a lower precipitation rate of calcite in the earthworm calciferous glands and a consequent reduction in the partitioning of Sr into the calcite. This also raises the possibility that the apparent preferential partitioning of Sr into biogenic calcite compared to inorganic calcite discussed above may be due to precipitation kinetics.

The potential difference in partitioning of Sr to the granules between the amended and non-amended soils suggests that further investigation is warranted. As such we carried out spatially resolved studies to determine the compositional and crystallographic distribution of Sr in the granules.

### **3.4. Sr distribution within granules**

The internal structure of granules has been well described by Lee et al. (2008), and in this study comprises two distinct types, with each type being found in granules extracted from all the different treatments used in this study. The first type comprise a densely compact aggregate of >30  $\mu\text{m}$  carbonate crystals whose orientations appear to be consistent (often radial) within individual layers (e.g. Fig. 2a). These granules are made up of only a few distinct layers. The second type are silicate-inclusion rich and comprise several concentric rings that are poorly linked together and are interspersed with large void spaces. The carbonate crystals within these types of granules are generally smaller than those of the more dense granules (e.g. Fig. 2b).

EPMA element maps of the granules from Hamble soils all show concentric zoning of Sr, Mg and Mn. Similar zoning has been observed previously in granules produced by *L. terrestris* in Hamble soil using cathodoluminescence and SEM imaging (Lee et. al., 2008). There is no relationship between type of zoning and the different granule morphologies described above. In general the concentric rings for the Sr were broad in nature. However, there appears to be no consistency to the zoning within the granules either across the study or within individual experiments / Sr amendments (Fig. 3). Some granules from the Hamble soils showed Sr-rich cores and rims, some granules showed Sr-rich cores and Sr-poor rims, and some granules showed Sr-poor cores with Sr-rich rims.

In contrast to the Sr, the concentric rings for Mg were generally very narrow and distinct (Fig. 4) reflecting relatively large changes in Mg concentration (Table 4). This wide range in Mg concentrations suggests that, unlike various other organisms (e.g. Bentov and Erez, 2006; Lorens and Bender, 1977; Wang et al., 2012) there is no biotic control on the Mg content of the earthworm-produced calcite. There appears to be no consistency between the granules regarding the number, frequency or radial position of the Mg rich rings. The zoning of the Mn within the granules was generally more diffuse than that for Sr and Mg, and the concentric nature of the zoning showed broad bands rather than the ring structure displayed

by Mg. The Mn zoning patterns within the granules had far more consistency than for Sr and Mg with granules from the same experiment displaying the same basic distribution of Mn-rich and Mn-poor bands.

Statistical correlations between Ca, Sr, Mg and Mn were determined for both the EPMA element maps and the quantitative point analyses across the rim-core-rim line profiles. No significant correlations exist between any pair of elements.

Barker and Cox (2011) showed that laboratory synthesized inorganic calcite co-precipitated with rare earth elements shows the same style of zoning as that observed in the granules strongly suggesting that inorganic processes are responsible for the granule zoning. The zoning is likely due to episodic elemental enrichment of the fluid surrounding the granules and the subsequent incorporation of the enriched elements in the growing granules. These fluctuations in concentration may be due to the composition of soil and soil solution that the earthworm encounters at any given moment in time. Alternatively, or additionally, they may be due to the incorporation of the trace elements into the granules being more rapid than their replenishment at the granule growth front by diffusion through the fluid in the calciferous gland; over time the concentration of the granule-incorporated element would again increase in the fluid leading to its renewed incorporation into the granule (Shore and Fowler, 1996). The apparent inconsistency in the zonation pattern for granules within the same experiment may be due to changes over time in the chemistry of the fluid from which the granules precipitate. In addition the elemental zoning may relate to changes in the mechanism or pathway of the calcium carbonate formation reaction with elemental enrichment being associated with the behaviour of potential precursor phases such as amorphous calcium carbonate and/or vaterite.

The granules produced by *L. terrestris* in the Yate soil showed a different pattern of elemental zoning. Sr was only present in a 5 – 60  $\mu\text{m}$  wide zone around the rim of the YSL granules and only present in ~200  $\mu\text{m}$  diameter patches of microcrystalline calcium

carbonate in YSH granules. The zonation pattern for Mg within the YSL and YSH granules was the same as that for the granules from Sr amended Hamble soil, while Mn showed no zoning pattern in YSH and a small enriched rim in the YSL granules that correlated with the zoning shown by Sr. These differences may reflect differences in the speciation of these elements, and hence their availability for uptake, between the amended and naturally Sr-rich soils. However, none-with-standing the large range of Mg values shown in the zoning mitigating against a biological control over the Mg content of the calcite, the similarity in the Mg zoning between the HS and YSL and YSH granules may somehow reflect the biological origins of the granules.

### **3.5. Sr and Ca $\mu$ XAS of the granules**

#### **3.5.1. Ca K-edge XANES**

The Ca XANES spectra of the calcium carbonate standards and representative spectra obtained from granules recovered from the experiments are shown in Figure 5. For the HS granules we only present spectra obtained from a granule extracted from HS100 as these are typical of the spectra obtained from the granules extracted from the other HS soils, which themselves are presented in the Supplementary Information. Also given in the Supplementary Information are spectra from other points analysed on granules extracted from YSL and YSH.

The Ca XANES spectrum of the calcite standard has a pre-edge feature with two peaks at 4.039 and 4.042 keV (marked “1” and “2” in Fig. 5) compared to the other standards which show only one peak in the pre-edge. This pre-edge feature has previously been highlighted by Lam et al. (2007) and Gebauer et al. (2010) and can be used to differentiate calcite from vaterite or aragonite. The vaterite Ca-XANES spectrum in Fig. 5 is

similar to vaterite spectra previously reported in the literature (Bots, 2012; Gebauer et al., 2010; Hayakawa et al., 2008; Lam et al., 2007). This spectrum has a single broad peak as a pre-edge feature at 4.040 keV and two broad post edge peaks at 4.048 keV (marked “3” in Fig. 5) and 4.056 keV (marked “4” in Fig. 5) and these two features can be used to differentiate between vaterite and aragonite.

Typical Ca XANES spectra obtained from the granules are shown in Fig. 5. The majority of the spectra (43 out of 45) obtained from granules recovered from the amended Hamble soils from which granules were analysed (e.g. HS100 point A and B), all 6 spectra obtained from granules recovered from YSL (e.g. YSL Point A) and 6 of the 10 spectra obtained from granules recovered from YSH (e.g. YSH Point A) are similar to the calcite Ca K-edge XANES spectrum (Fig. 5). The structural motif of the diagnostic pre- and post-edge features at c. 4.048 eV and 4.060 eV are the same in all these spectra, and thus confirm that these granules are mostly calcitic in nature. Small differences in the post-edge region of these spectra exist, specifically in the shape of the main peak and the broadness of the second oscillation, which are similar to those described in our earlier study of Pb in earthworm granules (Fraser et al., 2011). As described earlier, the granules predominantly comprise large single crystals of calcite with respect to the size of the microbeam and thus each spectrum is likely to be associated with one individual calcite crystal. Consequently, the small differences observed in these spectra are probably due to differences in the orientation of the calcite crystals with respect to the polarised nature of the X-ray beam (see Supplementary Information for more detail). Some of the differences observed between these spectra, however, may be the result of electronic or crystal-structure changes induced by structural incorporation of Sr or Mn into the calcite lattice. The other two Ca XANES spectra from granules recovered from Hamble soil (i.e. 2 of the 45 spectra) are indicative of mainly vaterite and are represented on Fig 5 by the spectrum labelled HS100 Point C. The remaining 4 spectra obtained from granules recovered from YSH (e.g. YSH Point B in Fig. 5) are also indicative of vaterite. The difference in the pre-edge region between the two

polymorphs is clearly shown in the difference in the derivative spectra near 4.039 keV shown in the inset of Fig 5.

Hence the Ca K-edge XANES analysis only indicates the presence of two calcium carbonate phases. It should be noted that the attenuation length for X-rays just after the Ca K-edge in calcite is around 8 microns, thus each XANES spectrum is from c. 200  $\mu\text{m}^3$  of the sample. As XAS is an averaging technique, phases that are present at less than about 10 volume% will be difficult to identify. We see calcite XANES for nearly all the points sampled in granules obtained from both the Hamble and YSL soils with only an occasional spectrum of vaterite from a Hamble soil granule. Thus, in agreement with the XRD results (Table 4), we conclude calcite is the dominant phase for the HS and YSL granules with vaterite a minority phase in the HS granules. Ca K-edge XANES for granules extracted from YSH indicated the presence of both calcite (YSH Point A, Fig. 5) and vaterite (YSH Point B, Fig. 5), with vaterite XANES being recorded more frequently than in the granules from the other soils. Again the Ca K-edge XANES are in broad agreement with the XRD results (Table 4) whereby the mineralogy of the YSH granules is mostly calcite with additional vaterite.

### 3.5.2. Sr K-edge XANES and EXAFS

Sr K-edge XANES spectra recorded from the Sr standards and typical Sr K-edge XANES from the granules are shown in Fig. 6. All the Sr K-edge XANES spectra collected from different granules are presented in the Supplementary Information. The majority of the Sr K-edge XANES spectra resemble the spectra of our standards “Sr adsorbed onto calcite” and “Sr adsorbed onto granule” (Fig. 6), though some small differences are evident in the intensity of the shoulder (or first oscillation) on the high energy side of the white line at c. 16.122 keV. As described for such differences observed between the Ca K-edge XANES spectra, these slight variations are probably related to the orientation of the sample with respect to the polarization of the X-ray beam. The Sr K-edge XANES spectra in Fig. 6 for the

granules recovered from the soils (except YSH Point B) and the 2 standards for Sr adsorbed onto calcite and granules are very similar to those previously reported in the literature for Sr structurally incorporated into the calcite lattice (Pingatore et al., 1992; Parkman et al., 1998; Fujita et al., 2004; Finch and Allison, 2007). This suggests that during the preparation of our standards “Sr adsorbed onto calcite” and “Sr adsorbed onto granules” Sr has become structurally incorporated into the calcite lattice, either through diffusion or via a dissolution re-precipitation mechanism (Lakshtanov and Stipp, 2007; Stipp and Hochella, 1991; Tang et al., 2008; Tesoriero and Pankow, 1996). Furthermore, it suggests that in the majority of our granules the Sr is structurally bound within the calcite lattice.

Some of the Sr K-edge XANES spectra obtained from granules extracted from the YSH soil (i.e. 5 of the 11 spectra) appear to show significant differences to the majority of the Sr K-edge XANES spectra; these spectra are represented in Fig. 6 by spectrum YSH Point B. They are similar both in appearance and also in terms of the energy shift of the white line with respect to the other spectra of Fig. 6 to the Sr K-edge XANES spectrum of the inorganically synthesised Sr co-precipitated vaterite standard. Thus we assign them to Sr structurally incorporated into vaterite, an interpretation supported by the Ca K-edge XANES and XRD results.

Of all the Sr K-edge EXAFS spectra from the granules, 34 out of the 35 spectra obtained from 5 different granules produced in the different Sr-amended Hamble soil (e.g. HS100 Points A and B, Fig. 6), all 5 spectra obtained from granules produced in YSL (e.g. YSL Point A, Fig. 6) and 6 of the 11 spectra obtained from granules produced in YSH (e.g. YSH Point A, Fig. 6) look very similar to each other and also to the EXAFS of our standards “Sr adsorbed on calcite” and “Sr adsorbed on granules”. These spectra were collected from the same points for which the Sr K-edge XANES spectra were collected, and which suggested that the Sr was structurally bound within the calcite. Pingatore et al. (1992) suggested a model for Sr within the calcite lattice in which the first 3 shells comprise six O



atoms, six C atoms and six Ca atoms, respectively. This model, which reflects well the crystal structure reported by Effenberger et al. (1981), also represents the best fit to the Sr K-edge EXAFS spectra for these points whereby Sr is surrounded by six O atoms at a distance of 2.52 Å, followed by a six C atoms and six Ca atoms at a distance of c. 3.33 Å and c. 4.09 Å, respectively (see Table 5). This is the model for incorporation into calcite used by Elzinga and Reeder (2002) in their studies of other elements. It differs from the models used by Parkman et al. (1992) and Finch and Allison (2007), in that they use a shell occupancy of 3 for C in the second coordination shell. However the Sr-C and Sr-Ca distances in our model are similar to those reported in both of those studies.

The  $\text{Sr}^{2+}$  ion is substantially bigger than the  $\text{Ca}^{2+}$  ion with a six-coordinate ionic radius of 1.21 Å compared with 1.00 Å for Ca (Shannon, 1976) and thus it is expected that the calcite lattice would be locally expanded around a substituting Sr. We find a Sr-O bond distance of c. 2.51 Å compared to 2.36 Å for Ca-O in calcite (Effenberger et al., 1981). The Sr-EXAFS data from granules obtained in HS soils indicates that Sr replaces Ca in the calcite lattice with a local structural distortion of +7.2% for Ca-O bond lengths and +3.4% for Ca-C distances, respectively. For comparison, Finch and Allison (2007), quantified a 6.5% local dilation in the calcite structure as a consequence of the Sr substitution.

The Sr substitution into calcite model fits the Sr data at HS100 Point C (Fig. 7); at the same point the Ca K-edge XANES indicated the presence of vaterite (Fig. 5). This apparent discrepancy is due to differences in the sampling volume for Ca and Sr XAS. The attenuation length of X-rays at the Sr K-edge (16.1 keV) in calcium carbonate is c. 350 µm, while just after the Ca K-edge (4.05 keV) it is only 8 µm. Thus the Sr EXAFS will be from the whole granule slice thickness while the Ca XANES represents only the top 8 µm or so. Consequently, while the Ca K-edge XANES data is from vaterite which dominates the outer 8 µm of the sample, the Sr K-edge XAS data is from both this vaterite and also from underlying calcite. As the calcite is the dominant phase in the sampled volume the Sr K-edge

EXAFS data can be well fitted by the Sr in calcite model. Indeed, the spectrum presented in Fig. 7 has some features that appear to be part way between those of Sr in calcite and those of YSH point B (see below).

Although the Sr in calcite model fitted well 6 of the 11 spectra collected from granules produced in YSH, the other 5 spectra resemble YSH Point B in Fig. 7 and these spectra are better fit by another model; (Sr with 8.5 O atoms at 2.55 Å, 5.2 C atoms at 2.95 Å and then 3.5 Ca atoms at 4.13 Å; Table 5). This model is the same as that for our standard vaterite co-precipitated with Sr (Table 5). The precise nature of the vaterite crystal structure is still under debate (Demichelis et al., 2012; Kamhi, 1963; Meyer, 1969; Meyer, 1960), but nevertheless it appears that the radial distribution of atoms about the Ca in vaterite is more complex than that of calcite. The "standard" crystallographic model for vaterite (e.g. Kamhi, 1963) has Ca-O, Ca-C, Ca-C and Ca-Ca interatomic distances of 2.28 Å, 2.96 Å, 3.32 and 4.24 Å, respectively. Ca-XAS results of Becker et al. (2003) on biogenic vaterite showed a vaterite model with distances of 2.37 Å for Ca-O, 3.09 Å for Ca-O or Ca-C and 4.24 Å for Ca-Ca. Due to the disordered nature of the vaterite crystal structure with respect to that of calcite (e.g. Demichelis et al. 2012), authors of Ca-EXAFS studies have been cautious interpreting or fitting shells and quantifying coordination numbers between 2.9 Å and 3.8 Å (Becker et al., 2003; Demichelis et al., 2012; Lam et al., 2007), but it is generally agreed that Ca-O and Ca-Ca distances of 2.37 Å and 4.24 Å, respectively are characteristic of vaterite. Taking into account the size difference between Sr and Ca our Sr K-edge EXAFS results on Sr incorporation into vaterite are in broad agreement with the existing Ca-XAS data on biogenic and inorganically synthesised vaterite (Becker et al., 2003; Lam et al., 2007), strongly suggesting that Sr can substitute for Ca in the vaterite structure. Furthermore, this study provides evidence of this occurring in inorganically synthesized vaterite as well as vaterite biogenically produced by the earthworm *Lumbricus terrestris*.

#### 4. CONCLUSIONS

Granules of calcium carbonate secreted by the earthworm *Lumbricus terrestris* in Sr-rich soils, both those amended with Sr in the form of  $\text{Sr}(\text{NO}_3)_2$  and those that are naturally Sr-rich due to mineralisation, are predominantly calcite with minor amounts of vaterite. In contrast to our previous experiments in which granules were produced in Pb-rich soils (Fraser et al., 2011) no aragonite was found; nor did we detect vaterite in our Pb-rich granules. These findings suggest that the chemistry of the soil or soil solution that the earthworms are exposed to influences granule mineralogy. However, we detected no systematic differences in granule mineralogy across the soils investigated in this study despite their significantly different Sr contents.

The mode of incorporation of Sr in the granules appears to differ from that of Pb in our previous study. Pb was concentrated around granule edges and was predominantly adsorbed to the granule surface prior to secretion of the granules with smaller amounts present either as Pb in calcite or cerussite. Sr is incorporated throughout the granules giving rise to oscillatory zoning with no Sr-carbonate phase being detected. This reflects the increased incorporation of Sr into the calcite lattice by comparison to that observed for Pb (Fraser et al., 2011).

The Sr content of the granules was at the high end of concentrations in calcite previously reported in the literature; distribution coefficients for the partitioning of the Sr into the granules were relatively high compared to those reported for inorganic systems. This may reflect a kinetic effect such as those reported in inorganic systems (e.g. Nehrke et al., 2007; Tang et al., 2008; Tesoriero and Pankow, 1996; Gabitov and Watson, 2006) with the distribution coefficients being lowest for soils in which granule production was also the lowest. The distribution coefficients indicate that, despite their chemical similarities, earthworms are able to metabolically differentiate between Ca and Sr, both in terms of

uptake from the soil and incorporation into the calcium carbonate granules which they produce.

Whilst granules are unlikely to concentrate Sr from the soil solution relative to Ca, or concentrate Sr from the bulk soil relative to Ca in naturally Sr-rich soils, our results show that if a soil were to experience a significant increase in its Sr concentration, for example by the accidental release of a  $^{90}\text{Sr}$ -rich fluid, calcium carbonate granules could accumulate Sr relative to Ca from both the bulk soil and from earthworms. Thus earthworm secreted calcium carbonate granules may have a role to play in the movement of  $^{90}\text{Sr}$  in terrestrial ecosystems.

Granules have been shown to survive thousands of years in the soil (e.g. Canti, 2007), The use of granules as a record of palaeotemperatures is currently being explored (Versteegh et al., 2012). Our findings, that Sr substitutes into the granules with minimal modification of the calcite structure is encouraging for its use as a palaeoproxy. However the use of soil-based mineral palaeoproxies is fraught with difficulty due to the heterogeneity of soils. Much further work on the impact of soil chemistry and temperature on the partitioning of Sr into earthworm secreted calcium carbonate would be required before the full potential of granule trace element chemistry as a palaeoproxy can be assessed.

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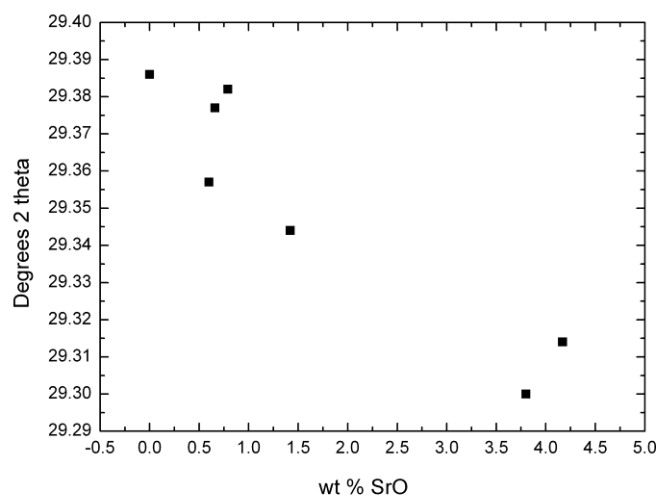
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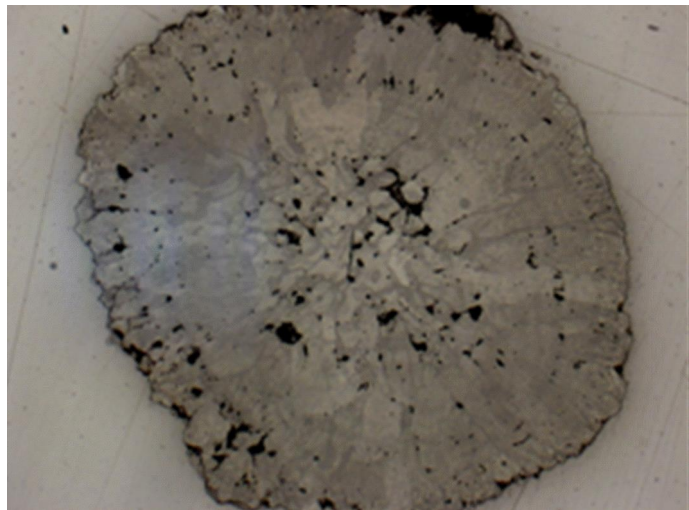
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## Figure Captions



**Figure 1.** Plot of the calcite 104 peak position ( $\lambda = \text{Cu K}\alpha_1$ ) as a function of average wt% SrO as estimated from point EPMA analyses across a core-rim line profile.

978 **Figure 2.** Optical images of granules produced by *L. terrestris* in Hamble soils amended with  
979 various Sr concentrations showing the two general morphological types that the granules



980 displayed. (a) is a granule from HS500  
981 (b) is a granule from HS100. Granules are c. 2 mm in diameter.

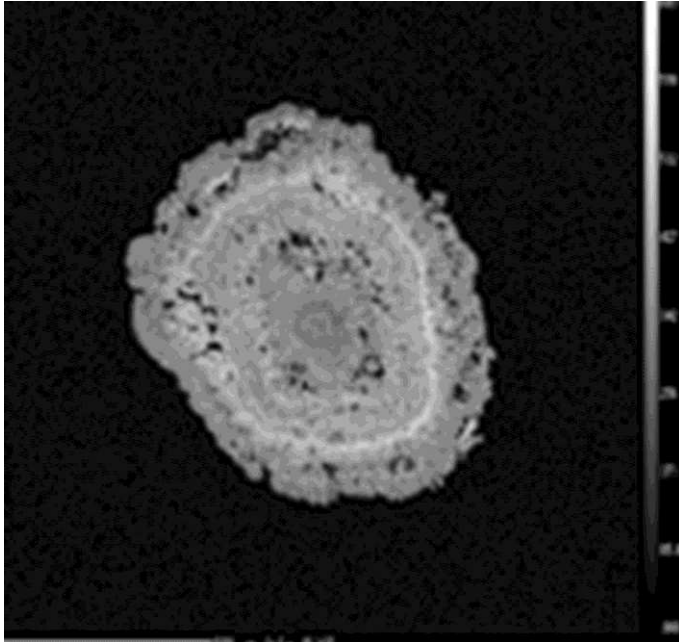


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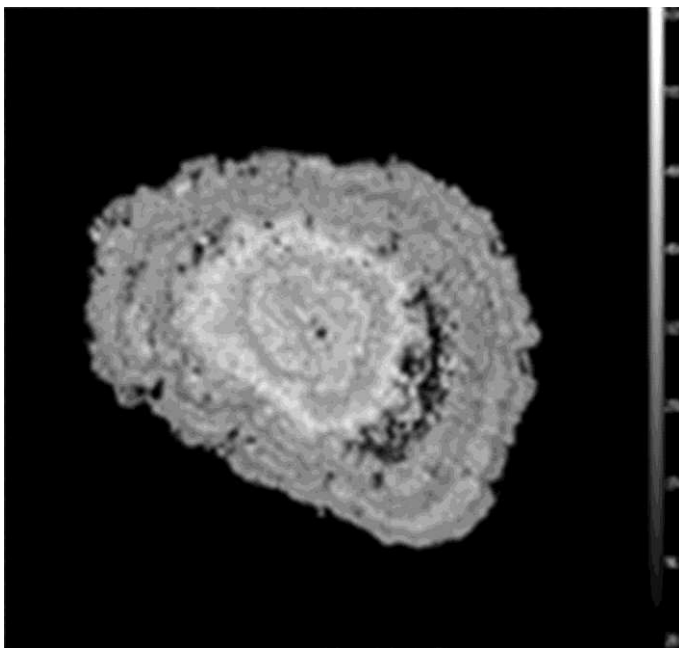


**Figure 3.** Sr distribution maps from EPMA of granules produced by *L. terrestris* in Hamble soils amended with various Sr concentrations.

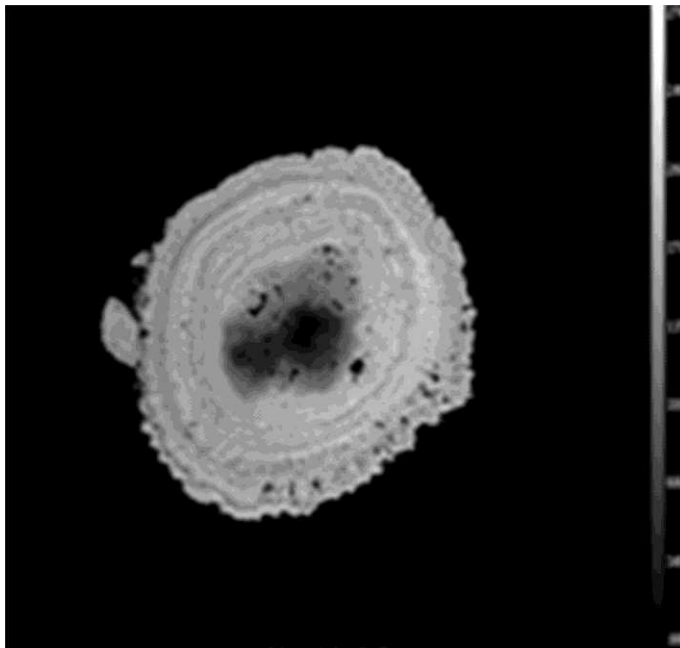
(a) a granule recovered from the depurate of earthworms kept in HS100



(b) a granule recovered from the depurate of earthworms kept in HS100



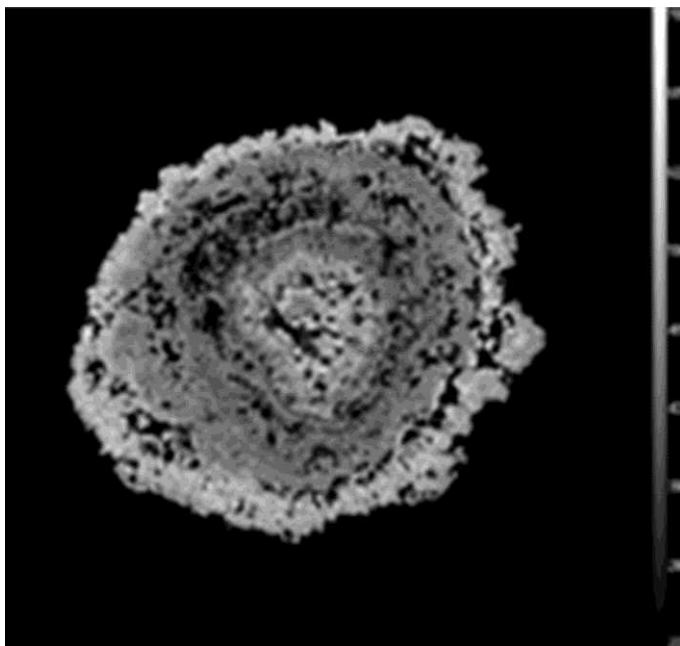
995 (c) a granule extracted from HS500 and



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998 (d) a granule extracted from HS100.

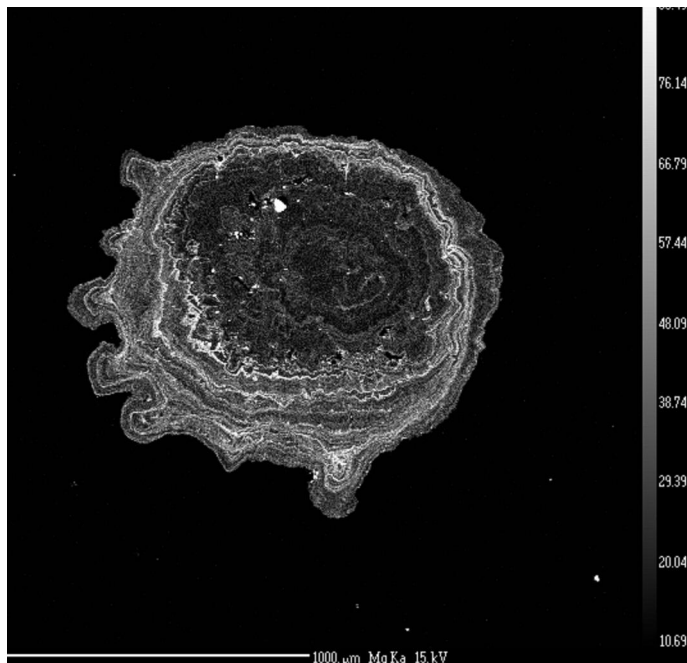


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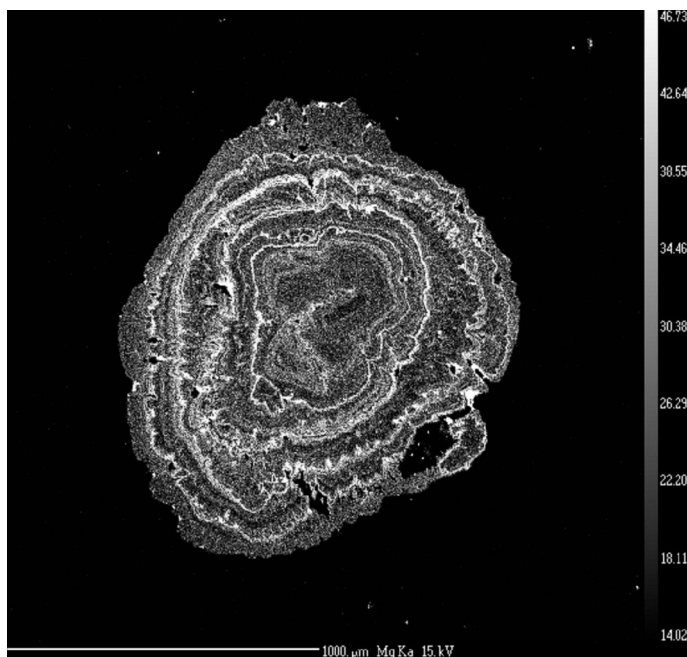
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**Figure 4.** Mg and Mn distribution maps from EPMA of granules produced by *L. terrestris* in Hamble soils amended with various Sr concentrations.

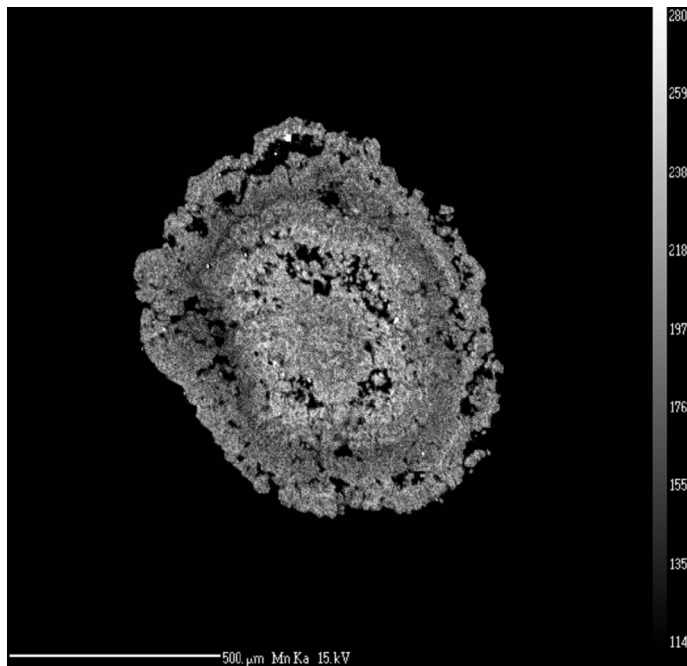
(a) Mg map from a granule from HS150



(b) Mg map of a granule from HS100



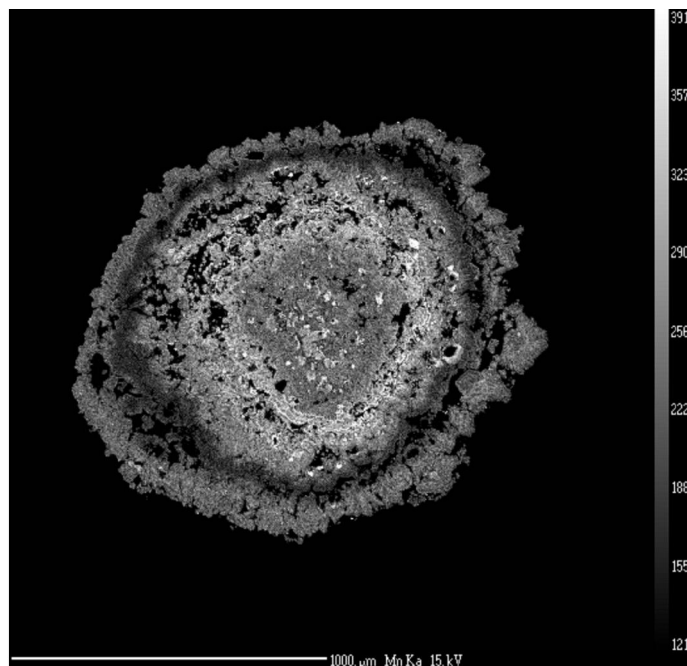
1010 (c) Mn map of a granule recovered from the depurate of an earthworm kept in HS100 and



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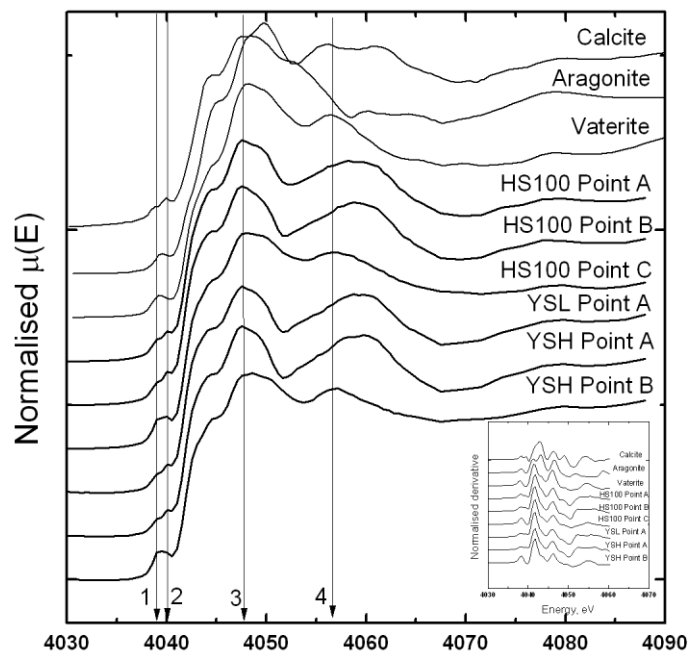
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1013 (d) Mn map of a granule from HS100.

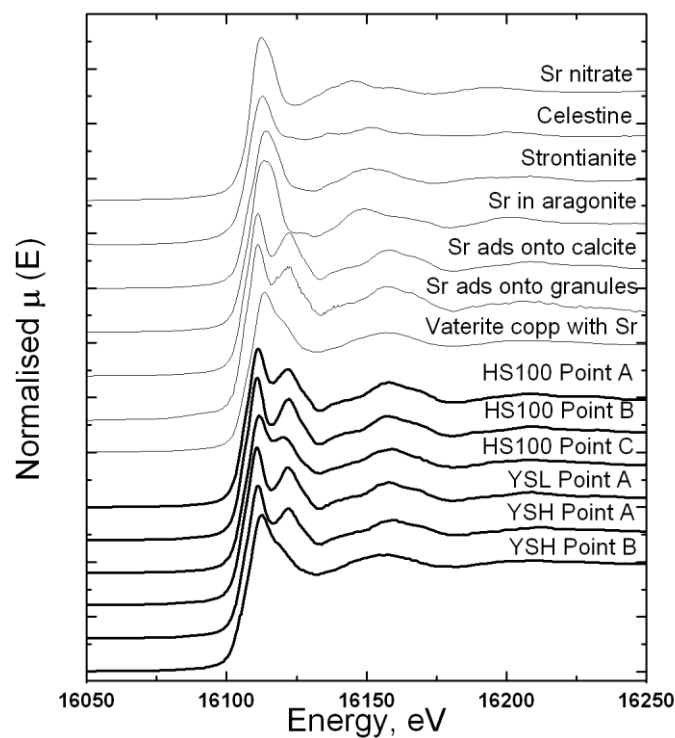


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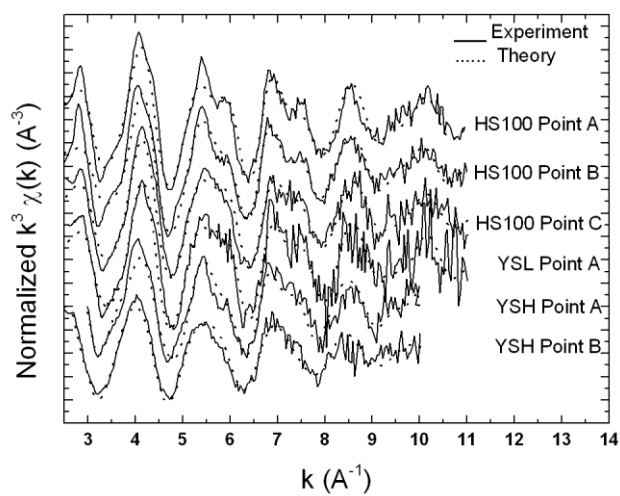
**Figure 5.** Ca K-edge XANES spectra for the carbonate standards: calcite (data from Fraser et al., 2011), Sr-bearing aragonite (obtained from Dr A. Finch, University of St Andrews) and synthetic vaterite (obtained from Dr P. Bots, University of Leeds) and for selected points from individual granules extracted from HS100, YSL and YSH. The first derivative of the pre-edge and edge are shown in the inset. “1”, “2”, “3” and “4” highlight diagnostic features of the spectra (see text for details).



**Figure 6.** Sr K -edge XANES spectra for the Sr standards used in this study and for three points on a granule from HS100 (HS100 Points A to C), a point on a granule from YSL (YSL Point A) and two points on a granule from YSH (YSH Point A and B). Spectra were collected from the same points as those in Fig. 5. For the standards “Sr ads onto calcite” and “Sr ads onto granule” are for Sr adsorbed onto calcite and granules, respectively. “Vaterite copp with Sr” is for the synthetic vaterite co-precipitated with Sr.



1031 **Figure 7.**  $k^3$ -weighted EXAFS spectra with EXAFS model (dotted line) for the same data as  
 1032 Fig. 6.



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Table 1. Soil properties. Values are mean  $\pm$  s.d. with number of replicates given in brackets.

| Parameter /Soil                               | HS                        | HS50                       | HS100                      | HS150                      | HS500                      | YSL                        | YSH                         |
|-----------------------------------------------|---------------------------|----------------------------|----------------------------|----------------------------|----------------------------|----------------------------|-----------------------------|
| pH                                            | 8.3<br>(n = 1)            | 8.18 $\pm$ 0.10<br>(n = 5) | 8.24 $\pm$ 0.02<br>(n = 5) | 8.24 $\pm$ 0.0<br>(n = 5)  | 8.05 $\pm$ 0.06<br>(n = 5) | 5.62 $\pm$ 0.16<br>(n = 5) | 8.06 $\pm$ 0.22<br>(n = 5)  |
| WHC / % (n = 3)                               | 39.9 $\pm$ 0.5            | ND <sup>a</sup>            | ND <sup>a</sup>            | ND <sup>a</sup>            | ND <sup>a</sup>            | 74.9 $\pm$ 0.2             | 51.4 $\pm$ 9.4              |
| Organic matter content (LOI) / %<br>(n = 1)   | 3.0                       | ND <sup>a</sup>            | ND <sup>a</sup>            | ND <sup>a</sup>            | ND <sup>a</sup>            | 9.2                        | 3.2                         |
| Target Sr concentration / mg kg <sup>-1</sup> | -                         | 50                         | 100                        | 150                        | 500                        | -                          | -                           |
| Actual Soil Sr / mg kg <sup>-1</sup>          | 32 $\pm$ 2<br>(n = 3)     | 72 $\pm$ 6<br>(n = 5)      | 145 $\pm$ 42<br>(n=5)      | 180 $\pm$ 30<br>(n=5)      | 600 $\pm$ 50<br>(n=5)      | 950 $\pm$ 50<br>(n=5)      | 11000 $\pm$ 900<br>(n=5)    |
| Soil Ca / mg kg <sup>-1</sup>                 | 8550 $\pm$ 990<br>(n = 3) | 8 600 $\pm$ 400<br>(n = 5) | 8 900 $\pm$ 500<br>(n = 5) | 8 100 $\pm$ 400<br>(n = 5) | 9 800 $\pm$ 300 (n = 5)    | 3 540 $\pm$ 240<br>(n = 5) | 24 100 $\pm$ 800<br>(n = 5) |
| Soil solution Sr / mg L <sup>-1</sup>         | ND <sup>a</sup>           | 5.58 $\pm$ 1.35<br>(n = 5) | 10.1 $\pm$ 2.79<br>(n = 5) | 19.7 $\pm$ 1.96<br>(n = 5) | 131 $\pm$ 25.28<br>(n = 5) | 21.9 $\pm$ 10.9<br>(n = 5) | 80.8 $\pm$ 11.6<br>(n = 5)  |
| Soil solution Ca / mg L <sup>-1</sup>         | ND <sup>a</sup>           | 288 $\pm$ 81               | 298 $\pm$ 55               | 359 $\pm$ 117              | 790 $\pm$ 49               | 95 $\pm$ 44                | 228 $\pm$ 38                |

<sup>a</sup>ND = Not determined, property assumed to be unaffected by addition of Sr salt to Hamble soil



Sr in earthworm granules

Table 2. Earthworm and granule data. Values either represent mean values  $\pm$  s.d. (n = 5) or, for granules, single values derived from the combining of all 5 replicates to give sufficient mass for accurate measurement and analysis.

| Parameter /Soil                                                                             | HS                        | HS50             | HS100            | HS150              | HS500            | YSL            | YSH              |
|---------------------------------------------------------------------------------------------|---------------------------|------------------|------------------|--------------------|------------------|----------------|------------------|
| Earthworm Sr / mg kg <sup>-1</sup>                                                          | 13.6 $\pm$ 6.10           | 69.1 $\pm$ 17.1  | 179 $\pm$ 83.5   | 213 $\pm$ 36.5     | 708 $\pm$ 112    | 603 $\pm$ 92.9 | 4100 $\pm$ 1070  |
| Earthworm Ca / mg kg <sup>-1</sup>                                                          | 10600 $\pm$<br>1500       | 10500 $\pm$ 1980 | 11900 $\pm$ 4300 | 9550 $\pm$<br>1960 | 11300 $\pm$ 1600 | 6100 $\pm$ 508 | 11300 $\pm$ 1500 |
| Granule Sr / mg kg <sup>-1</sup>                                                            | 345 $\pm$ 23 <sup>a</sup> | 4000             | 7930             | 12000              | 34200            | 14700          | 51400            |
| Granule Ca mg kg <sup>-1</sup>                                                              | ND                        | 425000           | 449000           | 427000             | 447000           | 338000         | 402000           |
| Production rate / mg CaCO <sub>3</sub> g <sub>worm</sub> <sup>-1</sup><br>day <sup>-1</sup> | 0.28                      | 0.47             | 0.38             | 0.32               | 0.48             | 0.05           | 0.19             |
| Production rate / mg CaCO <sub>3</sub><br>earthworm <sup>-1</sup> day <sup>-1</sup>         | 1.52                      | 0.89             | 1.50             | 1.67               | 2.30             | 0.26           | 0.74             |

<sup>a</sup>values taken from Hamble soil data, Lee et al. (2008)

Table 3. Distribution coefficients for Sr and Ca partitioning. Values for soil and soil solution distribution coefficients are mean values  $\pm$  s.d. (n = 5). For granules, single values based on the mean soil, soil solution and earthworm concentrations of Sr and Ca and the Sr and Ca concentrations in the pooled granules from all 5 replicates are given.

| Components / Soil         | HS              | HS50            | HS100           | HS150           | HS500           | YSL             | YSH             |
|---------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Earthworm / soil          | $0.58 \pm 0.22$ | $0.79 \pm 0.10$ | $1.01 \pm 0.43$ | $1.05 \pm 0.29$ | $1.04 \pm 0.19$ | $0.37 \pm 0.05$ | $0.78 \pm 0.09$ |
| Earthworm / soil solution | ND              | $0.36 \pm 0.06$ | $0.42 \pm 0.19$ | $0.41 \pm 0.06$ | $0.38 \pm 0.06$ | $0.44 \pm 0.05$ | $1.01 \pm 0.16$ |
| Granule / soil            | ND              | 1.12            | 1.09            | 1.27            | 1.25            | 0.16            | 0.28            |
| Granule / soil solution   | ND              | 0.50            | 0.50            | 0.51            | 0.46            | 0.19            | 0.36            |
| Granule / earthworm       | ND              | 1.43            | 1.17            | 1.26            | 1.22            | 0.44            | 0.35            |

Table 4. The calcium carbonate mineralogy as identified by XRD (XRD-NHM and  $\mu$ XRD-NHM) and the trace element chemistry from EPMA of granules. The calcite 104 peak 2 $\theta$  position is provided based upon the wavelength of Cu K $\alpha_1$  radiation. The average wt% SrO, MgO and MnO levels are taken from EPMA analyses within a rim-to-core line profile.

| Sample name        | Carbonate phases identified | Calcite 104 peak 2 $\theta$ (XRD-NHM) | Range of SrO concentrations | Average wt % SrO  | Range of MgO concentrations | Average wt % MgO  | Range of MnO concentrations | Average wt % MnO  |
|--------------------|-----------------------------|---------------------------------------|-----------------------------|-------------------|-----------------------------|-------------------|-----------------------------|-------------------|
| HS control         | calcite, vaterite           | 29.386(2)                             | 0                           | 0.04 <sup>a</sup> | -                           | 0.06 <sup>a</sup> | -                           | 0.02 <sup>a</sup> |
| HS50               | calcite, vaterite           | 29.357(2)                             | 0.48-0.83                   | 0.60              | 0.04-0.017                  | 0.08              | 0.20-0.39                   | 0.30              |
| HS100              | calcite, vaterite           | 29.377(2)                             | 0.10-1.58                   | 0.66              | 0.06-0.23                   | 0.12              | 0.05-0.55                   | 0.22              |
| HS100              | Calcite                     | 29.382(2)                             | 0.56-1.14                   | 0.79              | 0.02-0.17                   | 0.07              | 0.19-0.41                   | 0.30              |
| FRESH <sup>b</sup> |                             |                                       |                             |                   |                             |                   |                             |                   |
| HS150              | Calcite                     | 29.344(2)                             | 0.97-1.99                   | 1.42              | 0.02-0.33                   | 0.10              | 0.02-0.45                   | 0.23              |
| HS150              | Calcite                     | 29.314(2)                             | 3.49-5.04                   | 4.17              | 0.01-0.10                   | 0.05              | 0.01-0.07                   | 0.04              |
| FRESH <sup>2</sup> |                             |                                       |                             |                   |                             |                   |                             |                   |
| HS500              | Calcite                     | 29.301(2)                             | 0.11-4.91                   | 3.85              | 0.02-0.19                   | 0.07              | 0.01-0.07                   | 0.04              |
| YSL                | calcite, vaterite           | 29.352(2)                             | 0.08-0.29                   | 0.12              | 0.08-0.19                   | 0.14              | 0.03-0.10                   | 0.06              |
| YSH                | calcite, vaterite           | 29.352(2)                             | 0.07-0.19                   | 0.13              | 0.12-0.28                   | 0.19              | 0.01-0.08                   | 0.04              |

<sup>a</sup> values taken from Lee et al. (2008)

<sup>b</sup> FRESH refers to granules collected from earthworm depurate at the end of the experiment rather than those recovered from the bulk soil by sieving

Table 5. Table summarising XAS fits for the selected points on a granules produced by *L. terrestris* in HS100, YSL and YSH together with relevant standards

| Point/Std                        | Notes                                             | Scattering Atom | Coordination Number | Interatomic distance (Å) | Debye-Waller factor (Å <sup>2</sup> ) | Fit Index |
|----------------------------------|---------------------------------------------------|-----------------|---------------------|--------------------------|---------------------------------------|-----------|
| Sr in HS100 Point A              | High Sr in the centre of the granule              | O               | 6                   | 2.51(±0.02)              | 0.013(±0.002)                         | 28.7      |
|                                  |                                                   | C               | 6                   | 3.38(±0.06)              | 0.040(±0.017)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.09(±0.02)              | 0.024(±0.005)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.04(±0.06)              | 0.032(±0.017)                         |           |
| Sr in HS100 Point B              | Low Sr on the edge of the granule                 | O               | 6                   | 2.52(±0.01)              | 0.014(±0.002)                         | 34.9      |
|                                  |                                                   | C               | 6                   | 3.35(±0.03)              | 0.030(±0.011)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.10(±0.04)              | 0.029(±0.008)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.04(±0.07)              | 0.027(±0.009)                         |           |
| Sr in HS100 Point C              | Medium Sr near a rim                              | O               | 6                   | 2.51(±0.01)              | 0.012(±0.003)                         | 31.5      |
|                                  |                                                   | C               | 6                   | 3.36(±0.06)              | 0.049(±0.027)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.10(±0.04)              | 0.036(±0.065)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.02(±0.07)              | 0.044(±0.022)                         |           |
| Sr in YSL Point A                | Medium Sr near the edge of the granule            | O               | 6                   | 2.50(±0.01)              | 0.013(±0.004)                         | 39.5      |
|                                  |                                                   | C               | 6                   | 3.32(±0.08)              | 0.035(±0.025)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.10(±0.06)              | 0.024(±0.007)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.02(±0.14)              | 0.040(±0.028)                         |           |
| Sr in YSH Point A                | High Sr on a hotspot top left side of the granule | O               | 6                   | 2.53(±0.02)              | 0.017(±0.004)                         | 33.7      |
|                                  |                                                   | C               | 6                   | 3.29(±0.11)              | 0.030(±0.017)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.08(±0.06)              | 0.037(±0.050)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.01(±0.09)              | 0.034(±0.036)                         |           |
| Sr adsorbed onto granule         |                                                   | O               | 6                   | 2.52(±0.02)              | 0.008(±0.006)                         | 33.0      |
|                                  |                                                   | C               | 6                   | 3.40(±0.19)              | 0.048(±0.082)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.10(±0.07)              | 0.028(±0.017)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.02(±0.06)              | 0.019(±0.027)                         |           |
| Sr in YSH Point B                | Medium Sr                                         | O               | 8.2                 | 2.55(±0.01)              | 0.025(±0.005)                         | 29.3      |
|                                  |                                                   | C               | 5.2                 | 2.95(±0.10)              | 0.035(±0.025)                         |           |
|                                  |                                                   | Ca              | 3.5                 | 4.13(±0.06)              | 0.040(±0.019)                         |           |
| Sr adsorbed onto calcite         |                                                   | O               | 6                   | 2.51(±0.01)              | 0.010(±0.004)                         | 31.7      |
|                                  |                                                   | C               | 6                   | 3.33(±0.11)              | 0.008(±0.017)                         |           |
|                                  |                                                   | Ca              | 6                   | 4.07(±0.03)              | 0.022(±0.010)                         |           |
|                                  |                                                   | Ca              | 6                   | 5.00(±0.04)              | 0.008(±0.007)                         |           |
| Vaterite co-precipitated with Sr |                                                   | O               | 8                   | 2.55(±0.01)              | 0.022(±0.003)                         | 23.8      |
|                                  |                                                   | C               | 5.7                 | 2.94(±0.15)              | 0.035(±0.022)                         |           |
|                                  |                                                   | Ca              | 4.4                 | 4.17(±0.04)              | 0.043(±0.014)                         |           |