UNIVERSITY of York

This is a repository copy of *Quantification of hyperpolarisation efficiency in SABRE and SABRE-Relay enhanced NMR spectroscopy*.

White Rose Research Online URL for this paper: <u>https://eprints.whiterose.ac.uk/136226/</u>

Version: Accepted Version

Article:

Richardson, Peter Michael orcid.org/0000-0002-6631-2459, John, Richard Owain, Parrott, Andrew et al. (5 more authors) (2018) Quantification of hyperpolarisation efficiency in SABRE and SABRE-Relay enhanced NMR spectroscopy. Physical Chemistry Chemical Physics. pp. 26362-26371. ISSN 1463-9084

https://doi.org/10.1039/C8CP05473H

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

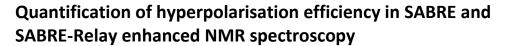
PCCP

ARTICLE

Received 00th January 20xx, Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/



Peter M. Richardson,^a Richard O. John,^a Andrew J. Parrott,^b Peter J. Rayner,^a Wissam Iali,^a Alison Nordon,^b Meghan E. Halse^{*a} and Simon B. Duckett^{*a}

Para-hydrogen (*p*-H₂) induced polarisation (PHIP) is an increasingly popular method for sensitivity enhancement in NMR spectroscopy. Its growing popularity is due in part to the introduction of the signal amplification by reversible exchange (SABRE) method that generates renewable hyperpolarisation in target analytes in seconds. A key benefit of PHIP and SABRE is that *p*-H₂ can be relatively easily and cheaply produced, with costs increasing with the desired level of *p*-H₂ purity. In this work, the efficiency of the SABRE polarisation transfer is explored by measuring the level of analyte hyperpolarisation as a function of the level of *p*-H₂ enrichment. A linear relationship was found between *p*-H₂ enrichment and analyte ¹H hyperpolarisation for a range of molecules, polarisation transfer catalysts, NMR detection fields and for both the SABRE and SABRE-Relay transfer mechanisms over the range 29 - 99 % *p*-H₂ purity. The gradient of these linear relationships were related to a simple theoretical model to define an overall efficiency parameter, *E*, that quantifies the net fraction of the NMR detection field and exceeds *E* = 20% for methyl-4,6-d₂-nicotinate when using a previously optimised catalyst system. For the SABRE-Relay transfer mechnism, efficiencies of up to *E* = 1% were found for ¹H polarisation of 1-propanol, when ammonia was used as the polarisation carrier.

Introduction

Nuclear magnetic resonance (NMR) spectroscopy is a widely used analytical tool for the identification, characterisation and quantification of molecules. However, many applications of NMR spectroscopy are limited by the relatively low sensitivity of the technique. The signal in an NMR experiment is derived from the population difference across nuclear spin states, the so-called polarisation. At thermal equilibrium, the nuclear polarisation is dictated by the Boltzmann distribution and is proportional to the applied static magnetic field.¹

The quest for increased polarisation and hence improved sensitivity has led to the development of large and expensive NMR spectrometers that use superconducting magnets to achieve very strong and homogeneous magnetic fields. An alternative route to increased NMR sensitivity is through hyperpolarisation. Hyperpolarisation refers to any method that generates a nuclear polarisation that is significantly larger than that dictated by the Boltzmann distribution at thermal equilibrium.²⁻⁵ A wide range of hyperpolarisation methods have been developed, each with different advantages and

challenges. The most widely used methods today include dynamic nuclear polarisation (DNP),^{6, 7} dissolution DNP (D-DNP),⁸⁻¹⁰ spin exchange optical pumping (SEOP),^{11, 12} brute force hyperpolarisation,¹³ and *para*-hydrogen (*p*-H₂) induced hyperpolarisation (PHIP).¹⁴⁻¹⁶

In this work we focus on PHIP methods, which use the singlet nuclear spin isomer of molecular hydrogen, *p*-H₂, as the source of polarisation. In PASADENA and ALTADENA, the original PHIP experiments of Bowers and Weitekamp,^{17, 18} hyperpolarisation is achieved by using *p*-H₂ in a hydrogenation reaction. PHIP has been widely used for the mechanistic study of hydrogenation reactions¹⁹⁻²⁴ and the generation of hyperpolarised MRI contrast agents for clinical diagnosis.²⁵⁻²⁷ More recently, the range of potential applications for PHIP has been increased with the introduction of the signal amplification by reversible exchange (SABRE) method.²⁸⁻³⁰ SABRE is a non-hydrogenative version of PHIP that catalytically transfers spin order from *p*-H₂ to a molecule of interest without chemical alteration of this target molecule.

There are many benefits of the SABRE approach when compared to other hyperpolarisation techniques. Firstly, the hyperpolarisation transfer step occurs outside of the NMR spectrometer in a weak polarisation transfer field (PTF) of the order of $0 - 10 \text{ mT.}^{30-33}$ Therefore the observed hyperpolarisation level is independent of the strength of the detection field. Secondly, polarisation is generated rapidly, over just a few seconds, thereby allowing for quick and simple experimental implementation. Thirdly, as no chemical change

^{a.}Centre for Hyperpolarisation in Magnetic Resonance, Department of Chemistry, University of York, UK.

b. WestCHEM, Department of Pure and Applied Chemistry and CPACT, University of Strathclyde, Glasgow, UK.

Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/x0xx00000x

occurs to the target analyte, the process is fully reversible. Hence, hyperpolarisation can be re-established easily by supplying fresh *p*-H₂.³⁴ Finally, *p*-H₂ can be generated relatively easily and can be stored for weeks to months.³⁵⁻³⁷ One of the key limitations of SABRE is the range of molecules that are amenable to hyperpolarisation due to the need for the target analyte to reversibly bind to the SABRE catalyst on a suitable timescale. This issue has been addressed recently by the introduction of a new mechanism for polarisation transfer called SABRE-Relay.^{38, 39} In principle this extends the SABRE method to include target molecules with any functional group that contains an exchangeable proton. Examples include, but are not limited to, alcohols, carboxylic acids, amines, amides, carbonates, and phosphates.³⁸

In this work we explore an aspect of SABRE that has implications both for the reproducibility of SABRE enhancement levels and the cost of the technology for a given application. Specifically, we study the relationship between the level of p-H₂ enrichment of the H₂ gas used in the SABRE reversible exchange reaction and the resultant level of hyperpolarisation on the target analyte. This relationship has cost implications because the level of p-H₂ enrichment is determined by the temperature at which the *ortho*-to-*para* conversion is achieved. ^{36, 40, 41} For example, cooling H₂ to 77 K via liquid N₂ is very inexpensive but only results in ~50% p-H₂ enrichment, whereas conversion temperatures below 30 K are required to reach >95% p-H₂ enrichment.³⁵ Instrumentation and maintenance costs in this temperature regime can be much higher.

Herein we experimentally determine the dependence of SABRE-derived hyperpolarisation on the p-H₂ enrichment level for different substrates, catalysts, NMR detection fields, and for both the standard SABRE and SABRE-Relay polarisation transfer mechanisms. By relating these empirical results to a simple model for p-H₂-derived polarisation, we extract an efficiency parameter that describes the fraction of p-H₂-derived polarisation that has been transferred to the target analyte. This measure can be used to evaluate the efficiency of a given experimental implementation of SABRE, independent of the p-H₂ enrichment level, and can also be used to determine the required level of p-H₂ enrichment for a given SABRE application.

Theory

Polarisation of H₂ as a function of *para*-hydrogen enrichment level

The SABRE technique, illustrated schematically in Figure 1, is a catalytic process for transferring the nuclear spin order of $p-H_2$ to a target analyte.³⁰ The polarisation transfer process is mediated by a transition metal complex that reversibly binds $p-H_2$ and one or more molecules of the target analyte (R₂ in Figure 1). The active SABRE complex establishes a *J*-coupling network between the $p-H_2$ -derived ¹H nuclei and the NMR-active nuclei in the bound target molecule. With appropriate coupling constants and polarisation transfer fields (PTF, typically 0 – 10 mT), there is a spontaneous transfer of spin



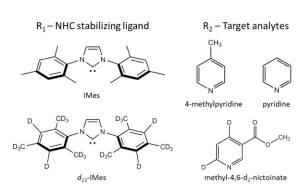


Figure 1. In the SABRE hyperpolarisation method, the active catalyst reversibly binds p-H₂ and one or more molecules of the target analyte (R₂). This establishes a *J* coupling network between the p-H₂-derived hydrides and the NMR-active nuclei in R₂ such that polarisation transfer can occur. Rapid chemical exchange of both p-H₂ and R₂ leads to the build-up of hyperpolarised R₂ in free solution. This process is reversible, meaning that hyperpolarisation can be regenerated upon supply of fresh p-H₂. The form of the SABRE catalyst can be changed to optimise SABRE efficiency for different analytes by varying R₁, a stabilising N-heterocyclic carbene.

order from p-H₂ to the target molecule.^{30, 31, 33, 42, 43} As p-H₂ and the target molecules undergo rapid chemical exchange between the bound and free forms, there is a build-up of hyperpolarised target molecules in free solution. Under a continuous supply of fresh p-H₂, the level of polarisation of the target molecules in solution will reach a steady-state that is dictated by the efficiency of the polarisation transfer process, the rate of ligand exchange, and NMR relaxation. This steadystate is typically reached on a timescale of tens of seconds.

The overall efficiency of SABRE can be quantified by the proportion of the available polarisation (derived from $p-H_2$) that is transferred and subsequently detected on the target analyte in free solution. Molecular hydrogen has two nuclear spin isomers: *p*-H₂, a nuclear singlet state, and *ortho*-hydrogen $(o-H_2)$, a nuclear triplet state. The energy level diagram for H_2 is given in Figure 2a, where α and β denote the spin-up and spin-down states of the protons in H₂, respectively, and the singlet and triplet states are defined as: $|S_0\rangle = \frac{1}{\sqrt{2}}(|\alpha\beta\rangle - |\beta\alpha\rangle)$, $|T_0\rangle = \frac{1}{\sqrt{2}}(|\alpha\beta\rangle + |\beta\alpha\rangle), |T_1\rangle = |\alpha\alpha\rangle, \text{ and } |T_{-1}\rangle = |\beta\beta\rangle. \text{ At room}$ temperature, all four energy levels are roughly equally populated and so H₂ contains approximately 25% p-H₂ and 75% o-H₂. Following a chemical reaction, where the protons from H₂ are transferred into chemically and/or magnetically different environments in the product molecule, the energy level diagram for the H₂-derived protons can be drawn as shown in Figure 2b, where the exact separation of these energy levels will depend on the strength of the applied magnetic field and the chemical shift and J-coupling network of the H₂-derived protons within the product molecule.

Consider a hydrogen addition reaction involving H_2 , where the fraction of molecules that are in the *para* and *ortho* forms are defined as n_p and n_o , respectively. If we assume that the population associated with the *ortho* form is distributed evenly

Unpolarised

Journal Name

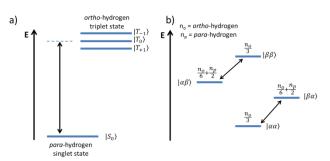


Figure 2. Energy level diagrams for the nuclear spin states of H₂ (a) before and (b) after undergoing a pair-wise hydrogenation reaction that leads to chemical or magnetic inequivalence between the two nuclei. Note: the two diagrams are not drawn to scale. The relationships between the states in (a) and (b) are as follows: $|S_0\rangle = \frac{1}{\sqrt{2}}(|\alpha\beta\rangle - |\beta\alpha\rangle)$, $|T_0\rangle = \frac{1}{\sqrt{2}}(|\alpha\beta\rangle + |\beta\alpha\rangle)$, $|T_{+1}\rangle = |\alpha\alpha\rangle$, and $|T_{-1}\rangle = |\beta\beta\rangle$.

amongst the three triplet states, we can assign a population of $n_o/3$ to each of the triplet states and a population of n_p to the singlet state. This would be the case at thermal equilibrium in a very weak magnetic field. Following a pair-wise hydrogen addition reaction, this population distribution will be mapped onto the energy levels in Figure 2b such that the populations of the superposition states, $(|\alpha\beta\rangle + |\beta\alpha\rangle)/\sqrt{2}$ and $(|\alpha\beta\rangle |\beta\alpha\rangle)/\sqrt{2}$, in H₂ are divided evenly between the $|\alpha\beta\rangle$ and $|\beta\alpha\rangle$ states of the protons in the product molecule.³⁵ As a result, the $|\alpha\alpha\rangle$ and $|\beta\beta\rangle$ states of the product molecule will have populations of $(n_o/3)$, while the |lphaeta
angle and |etalpha
angle states will have populations of $(n_o/6 + n_p/2)$. We define the latent polarisation of the system as the sum of the population difference between states that undergo single-quantum transitions, i.e. between $|\beta\alpha\rangle$ (or $|\alpha\beta\rangle$) and $|\beta\beta\rangle$ and $|\alpha\alpha\rangle$. The polarisation of the pair of protons from H_2 in the product molecule, P_{H_2} can therefore be defined by equation 1, where we have simplified the relationship using the substitution: $n_o = 1 - n_p$ and the population difference between each pair of energy levels is doubled to account for the two possible transitions.

$$P_{H_2} = 2\left(\frac{n_o}{6} + \frac{n_p}{2} - \frac{n_o}{3}\right) = \frac{4n_p}{3} - \frac{1}{3}$$
(1)

We note that at the moment of pair-wise p-H₂ addition, there is no detectable magnetisation because there is an even balance between the $\Delta m = +1$ and $\Delta m = -1$ transitions. However, in the case where the symmetry of H₂ is broken in the product molecule, equation 1 can be used to define the latent polarisation that will become observable as a result of coherent evolution under differences in chemical shift or *J*-coupling between the H₂-derived protons.^{31, 35}

Inspection of equation 1 reveals that when H₂ is at equilibrium at room temperature where the relative populations of the two isomers approach $n_p = 0.25$ and $n_o = 0.75$, the polarisation approaches zero, as expected. However, in the limit where p-H₂ enrichment, n_p , approaches 1, the polarisation also tends to 1, indicating the potential for large NMR signal enhancements. Therefore equation 1 provides a conversion between p-H₂ enrichment level, n_p , and the latent

ARTICLE

polarisation of the pair of protons in H_2 , which can be unlocked by a chemical reaction.

Quantifying para-hydrogen enrichment level

Transitions between the *ortho* and *para* states of H_2 are symmetry-forbidden. Therefore, conversion will only take place in the presence of a catalyst, for example a paramagnetic species such as iron (III) oxide or activated charcoal. To produce H_2 gas enriched in the *p*- H_2 state, it is cooled to the desired conversion temperature, *T*, in the presence of the catalyst. After conversion between the *ortho* and *para* states is achieved, the gas is heated up in the absence of the catalyst for use at room temperature. If carefully isolated from contact with paramagnetic species, the *p*- H_2 enriched gas can be stored at room temperature for long periods of time (i.e. weeks to months).^{36, 40, 41}

The level of p-H₂ enrichment achieved by this process will depend on the conversion temperature at which thermal equilibrium is established. The absolute populations of the p-H₂ (N_p) and o-H₂ (N_o) states at thermal equilibrium are governed by Boltzmann statistics. Due to the coupling of the even rotational states to the singlet nuclear spin state of p-H₂ and the coupling of the odd rotation states to the triplet nuclear spin states of o-H₂, N_p and N_o can be given by equations 2 and 3, where the rotational constant, θ_R , Is defined in equation 4, h is Planck's constant, k_B is Boltzmann's constant and I is the moment of inertia.³⁵

$$N_{p} = \sum_{J=even} (2J+1) \exp\left(-\frac{J(J+1)\theta_{R}}{T}\right)$$
(2)
$$N_{o} = 3 \sum_{J=odd} (2J+1) \exp\left(-\frac{J(J+1)\theta_{R}}{T}\right)$$
(3)
$$\theta_{R} = \frac{h^{2}}{8\pi^{2} I k_{R}}$$
(4)

In an NMR experiment, the observed signal from H₂ corresponds exclusively to o-H₂ because p-H₂ is a singlet and has no net angular momentum. Therefore, the amplitude of the observed signal for a fixed concentration of H₂ will depend on the relative level of o-H₂. If we define the observed NMR signal from o-H₂ as S_{ortho} and the maximum observable NMR signal, S_{max} , as that which would be observed in the case of 100% o-H₂ enrichment, we can relate the observed signal (S_{ortho}) to the populations of the two spin isomers, N_p and N_o as in equation 5.

$$S_{ortho} = S_{max} \frac{N_o}{N_o + N_p} \tag{5}$$

By inserting the expressions from equations 2 and 3 into equation 5, we can determine S_{max} and θ_R by quantifying the ¹H NMR signal response for H₂ gas that has been enriched at a range of conversion temperatures. Subsequently, the level of p-H₂ enrichment, n_p , can be calculated from either the NMR signal intensity for o-H₂ (S_{ortho}) or the conversion temperature (7) using equation 6 and the definitions for N_o and N_p in equations 2 and 3.

$$n_p = 1 - \frac{S_{ortho}}{S_{max}} = 1 - \frac{N_o}{N_o + N_p}$$
 (6)

Experimental

Para-hydrogen production

The generation of p-H₂ was achieved by using a closed helium refrigeration system which allows for temperatures down to 7 K. The system has been described previously but has had several modifications here.44 It consists of a compressor and low-temperature cooler, which provides a two-stage-closedcycle helium recirculation pump. In order to get good contact with the H₂ gas a copper block is used which is connected to the cooler. The copper block contains a tortuous void which introduces a significant residence time for H₂ in the presence of the interconversion catalyst (activated charcoal). The gas is passed through the copper block from an inlet at the bottom of the copper block to an outlet at the top. Since the cold head reaches a temperature of 7 K and at this temperature H_2 is a solid, the system has a feedback loop connected to a heater that is used to regulate the temperature of the copper block. A range of conversion temperatures was achieved by varying the heater power to stabilise at a given temperature. To hold the temperature at 140 K, the heater required around 60% of the maximum power, which was deemed to be the maximum safe operating level of the heater. Therefore, temperatures over the range of 140 K to 28 K were used. An additional point was acquired at 293 K by using thermally polarised H₂ gas taken directly from the H₂ cylinder. The feedback controller was able to hold the temperature of the cold head accurately to \pm 0.1 K, as measured by a thermocouple in the copper block. NMR tubes fitted with Young's valves can be attached to the apparatus via an adaptor. The system also includes a vacuum pump (Edwards RV pump), which can be combined with a H₂ bypass line to purge/flush the lines with H₂ before the addition of a sample. This vacuum pump also allows the removal of used H_2 from the headspace of the NMR tube before the subsequent addition of fresh $p-H_2$ gas. The hydrogen inlet was supplied with CP-grade hydrogen from an external cylinder via a regulator set to 3 bar (gauge) and the outlet pressure was monitored with a pressure gauge to ensure a fixed pressure of 4.0 ± 0.1 bar of gas was supplied at all times, measured with a digital pressure gauge (Baratron Pressure Transducer and Manometer MKS). When changing the temperature of the copper block, the system (including the void space within the copper block) was purged ten times before subsequent experiments were carried out to avoid any contamination from para-enriched H₂ generated at a different temperature.

Measuring p-H₂ concentration with NMR

In order to determine the p-H₂ concentration using liquid-state NMR, 0.6 mL of toluene- d_8 was placed inside an NMR tube fitted with a Young's valve (GPE scientific). The deuterated solvent was degassed using a 3-stage freeze-pump-thaw procedure with an acetone and dry-ice bath. 4 Bar (absolute) of H₂ gas was added to the headspace and the NMR tube was then shaken vigorously for 5 seconds to promote dissolution of the H₂ into the solvent. Each measurement was carried out using a fresh sample in a separate NMR tube. ¹H NMR measurements were carried out on a 500 MHz Bruker Avance III HD NMR spectrometer with a BBI probe using a single scan. This was repeated three times for each conversion temperature in order to take an average integral of the orthohydrogen signal. These average integrals were subsequently fitted to equation 5 to obtain fitting parameters that were used in the determination of the $p-H_2$ concentration (equation 6).

SABRE and SABRE-RELAY experiments

The SABRE enhanced ¹H NMR response was detected on either a 400 MHz Bruker Avance III using a BBI probe or a 43 MHz (1 T) Magritek Spinsolve Carbon benchtop NMR spectrometer. The SABRE samples contained 26 mM of the target analyte and 5.2 mM of the SABRE pre-catalyst in the form [IrCl(COD)(NHC)] (where COD = 1,5 cyclooctadiene) where the N-heterocyclic carbene (NHC) was either 1,3-bis(2,4,6-trimethylphenyl)imidazol-2-ylidine (IMes) or 1,3-bis(2,4,6-tris(methyl-d₃)-4,5-d₂phenyl)-imidazol-2-ylidine (d_{22} -IMes) (see Figure 1). The target analytes were: 4-methylpyridine, pyridine and methyl-4,6-d2nicotinate and the solvent was methanol- d_4 (Figure 1). Both of the catalysts and the methyl-4,6-d2-nicotinate were synthesised in-house,45 the analytes 4-methylpyridine and pyridine were purchased from Sigma-Aldrich. For each sample, a 7 mL bulk solution containing catalyst, analyte and solvent was prepared and 0.6 mL was subsequently distributed into ten different NMR tubes, each fitted with a Young's valve and degassed using a 3-stage freeze-pump-thaw-method in a bath of dry ice and acetone. The SABRE catalyst was activated by adding 4 bar $p-H_2$ to the headspace of the NMR tube and shaking vigorously for ten seconds. This was repeated six times and then the sample was left inside the NMR spectrometer for a further ten minutes to ensure full activation of each sample. Once activated, a single-scan thermal ¹H NMR spectrum was acquired as a reference for the SABRE enhancement factor calculation. For each subsequent SABRE experiment, the headspace of the NMR tube was evacuated and then charged with 4 bar p-H₂ at the desired enrichment level and shaken for 10 seconds in a handheld Halbach array with a static field of 63 G before being manually transferred to the NMR spectrometer for detection.⁴⁶ The sample transfer time was 3.5 ± 0.5 s. Each measurement was repeated 5 times.

Journal Name

For the SABRE-Relay method, samples were made by first preparing a 7 mL bulk solution containing 5.2 mM of the precatalyst [IrCl(COD)(IMes)] in dichloromethane- d_2 (DCM- d_2) in a 10 mm diameter NMR tube fitted with a Young's valve. The sample was degassed using a 3-stage freeze-thaw-pump process using liquid nitrogen. Ammonia gas was introduced to the head-space of the NMR tube and dipped in liquid nitrogen quickly to promote condensation, and the tube was sealed and subsequently shaken vigorously for 10 seconds to promote dissolution of the ammonia. The amount of ammonia in solution was quantified using liquid state ${}^{1}H$ NMR to be 42 ± 2 mM (see supporting information for full details). 26 mM (13.6 μ L) of 1-propanol was added to the NMR tube and the solution was de-gassed using a 3-stage freeze-pump-thaw method with liquid nitrogen. The 7 mL bulk solution was distributed to ten different NMR tubes for analysis (each 0.6 mL). The SABRE catalyst was activated by adding 4 bar of H₂ to the headspace of the NMR tube, which was then shaken for ten seconds and left overnight.

In order to calculate the enhancement factor and subsequently the polarisation, a single scan non-hyperpolarised ¹H NMR spectrum was acquired with the same settings as the subsequent SABRE-enhanced detection. The enhancement factors, ε , were then determined by taking a ratio of the thermal and hyperpolarised integrals as in equation 7.

$$\varepsilon = \frac{hyperpolarised integral}{thermal reference integral}$$
(7)

Polarisation, *P*, was calculated by scaling the enhancement factor to the thermal polarisation level in the detection field using equation 8, where γ is the gyromagnetic ratio, B_0 is the detection field, *T* is the sample temperature, \hbar is the reduced Planck's constant, and k_B is Boltzmann's constant.

$$P = \epsilon \frac{\gamma B_0 \hbar}{2k_B T} \tag{8}$$

Results and Discussion

Para-hydrogen enrichment calibration

Quantification of p-H₂ enrichment has been demonstrated previously using a range of experimental methods including thermal conductance,⁴⁷ Raman spectroscopy,^{48, 49} and ¹H NMR spectroscopy, where the NMR response of o-H₂ is measured directly in the gas phase.⁴⁰ Here we took a liquid-state NMR approach in which the *para*-enriched H₂ is dissolved in a fixed volume of toluene- d_8 and the ¹H NMR response for o-H₂ in solution is quantified. A similar approach has been employed previously to study the effect of p-H₂ conversion rates in diamagnetic solvents.⁴¹ The main advantage of liquid-state NMR is the increased sensitivity over the gas-phase approach, particularly when quantifying high levels of p-H₂ enrichment where the residual signals from o-H₂ are very weak.

Figure 3a presents the ¹H NMR response for p-H₂-enriched H₂ gas dissolved in toluene- d_8 as a function of conversion temperature. The solid line is a fit to equation 5, where the

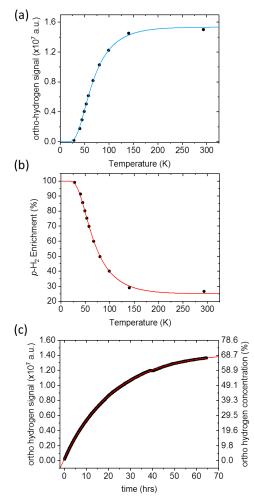


Figure 3. (a) ¹H NMR signal response level of *p*-H₂-enriched H₂ gas dissolved in toluene (black circles), for conversion temperatures ranging from 28 K to 293 K. A fit to equation 5 (blue line), gives a value of $\theta_R = (87.8 \pm 0.7)$ K and $S_{max} = (2.04 \pm 0.02) \times 10^7$, (b) the corresponding *p*-H₂ enrichment calculated from equation 6, where the red line was calculated using the fitted value of $\theta_R \cdot$ (c) ¹H NMR *o*-H₂ signal intensity monitored over 64 hours. At *t* = 0, hydrogen gas enriched to >99% *p*-H₂ at a conversion temperature of 28 K was dissolved in toluene-*d*₈ in an NMR tube and placed inside the NMR spectrometer. A fit to an exponential function to equilibrium (red line) reveals a time constant for the conversion to *o*-H₂ in this case of 22.62 ± 0.02 hours.

rotational constant was found to be $\theta_R = 87.8 \pm 0.7$ K. This is in excellent agreement with the value calculated previously from equation 4 (87.57 K ³⁵), as well as other experimentally determined values from the literature: $\theta_R = 87.6$ K,⁵⁰ $\theta_R = 84.837$ K ³⁶ and $\theta_R = 85.3$ K.⁵¹ Using the fitted value for the rotational constant, a plot of *p*-H₂ enrichment as a function of conversion temperature can be produced from equation 6, as demonstrated in Figure 3b.

The liquid-state NMR approach that we used to quantify the $o-H_2$ ¹H NMR response assumes that an equal concentration of H_2 is dissolved at each step of the experiment and that the conversion between $p-H_2$ and $o-H_2$ in solution is much slower than the timescale of our measurements. The validity of these assumptions is supported by the quality of the fits to the theoretical predictions in Figure 3a and 3b. This also suggests that the generator is very efficient at cooling and has sufficient catalyst surface area to reach equilibrium before the gas has left the cooling block. Further confirmation was obtained by

measuring the liquid-state ¹H NMR response for H₂ gas, enriched at 28 K and dissolved in toluene- d_8 , as a function of time inside the NMR spectrometer. The time-dependent ¹H NMR response in Figure 3c indicates a characteristic *para*-to*ortho* exponential conversion time constant in solution of the order of 23 hours, which is comparable to conversion times observed previously in the gas phase⁴⁰ and liquid phase.⁴¹ This is orders of magnitude slower than the tens of seconds required for each experimental measurement presented in Figure 3a and 3b. The fit to the data in Figure 3c shows that the signal tends to a value of (1.446 ± 0.001) x 10⁷ which is comparable to the intensity for H₂ at room temperature of (1.501 ± 0.001) x 10⁷, which implies that the concentration of H₂ in solution is not significantly changing throughout the experiment.

Signal amplification by reversible exchange (SABRE)

A series of SABRE hyperpolarisation measurements were recorded using H₂ gas containing known levels of p-H₂ enrichment, calculated from the conversion temperature using the calibration curve in Figure 3b. Figure 4 presents the relationship between the level of p-H₂ enrichment, n_p , and the observed polarisation levels for the individual resonances of the target molecule, P_S . It can be readily observed that the polarisation of the target molecule is linearly dependent on the percentage of $p-H_2$ for all resonances. This effect is robust across different target molecules (see Figures 4a, 4b and 4c) and for different SABRE catalysts (see Figure 4c and 4d) and is consistent with the analytical model of SABRE hyperpolarisation developed by Barskiy et al..⁵²

The slope of each linear correlation is a measure of the efficiency of the transfer of the latent polarisation of the *para*enriched H₂ to the target molecule. In order to quantify this, we define a SABRE efficiency parameter, *E*, as the ratio of the observed SABRE polarisation P_S to the polarisation of the *para*enhanced H₂ for a given level of *p*-H₂ enrichment, P_{H_2} .

$$E = \frac{P_S}{P_{H_2}} = \frac{3P_S}{4n_p - 1}$$
(9)

Rearranging we obtain a linear relationship between P_s and n_p with a slope of 4E/3.

$$P_{S} = \frac{4E}{3}n_{p} - \frac{E}{3}$$
(10)

Therefore, the efficiency parameter can be extracted directly from the slope of each linear correlation in Figure 4. We note that a similar approach has been used previously to quantify the efficiency of polarisation transfer when p-H₂ is used in a hydrogenation reaction (i.e. hydrogenative PHIP).⁵³

The SABRE efficiency parameters for the different substrate and catalyst systems in Figure 4 are presented in Table 1. These can be used to directly compare the efficacy of SABRE hyperpolarisation for the different systems. First, we note that the efficiency of SABRE is not uniform within each molecule. This well-known feature of SABRE must be taken into account when developing a quantitative model for SABRE.⁵⁴ However, the non-uniform distribution of polarisation is reproducible

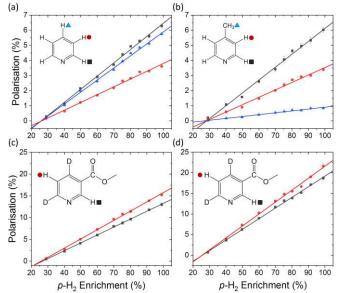


Figure 4. SABRE polarisation level as a function of p-H₂ enrichment for (a) R₂ = pyridine and R₁ = IMes, (b) R₂ = 4-methylpyridine and R₁ = IMes, (c) R₂ = methyl-4,6-d₂-nicotinate and R₁ = IMes, and (d) R₂ = methyl-4,6-d₂-nicotinate and R₁ = d₂₂-IMes. All spectra were acquired in methanol-d₄ using a 400 MHz Bruker NMR spectrometer at 298 K. The data points and error bars represent the average and ± one standard deviation of five repeat measurements, respectively. The point at 99% *p*-H₂ enrichment in (d) was calculated from SABRE-enhanced and thermally polarised ¹H NMR spectra that were acquired with a de-tuned probe to avoid radiation damping effects.

under fixed experimental conditions and so is unlikely to be a fundamental barrier to quantitative models based on SABRE. $^{\rm 54,}$ $^{\rm 55}$

Figure 4c and 4d illustrate the benefits of selective deuteration of the target molecule. This is beneficial because it concentrates the available polarisation on fewer sites and it increases the relaxation times of the remaining protons.⁴⁵ Finally, the use of a partially deuterated SABRE catalyst further improves the efficiency of the polarisation transfer.⁵⁶ It is postulated that this is due to the combined effect of increasing the NMR relaxation times of the target molecules bound to the catalyst and limiting the distribution of *p*-H₂-derived polarisation to the non-exchanging ligands. Thus there is a more efficient transfer to the target molecules bound *trans* to the hydrides in the active complex (see Figure 1).³⁰

In all of the examples presented in Figure 4, a consistent experimental procedure was used. However, this method can also be used to compare the efficacy of different experimental implementations. For example, this approach could also be used to explore the efficiency of automated SABRE approaches, where p-H₂ is bubbled through the solution and transferred to the NMR spectrometer for detection either manually³² or under flow.^{57, 58}

NMR detection at 1 T

With the introduction of low-cost, homogeneous permanent magnet benchtop spectrometers, high quality sub-ppb spectra can be acquired, thus increasing the utility of benchtop NMR for a broad range of applications.⁵⁹ It has been shown that the inherent sensitivity limit of low-field NMR can be overcome using various hyperpolarisation methods at fields from around 1 T down to ultra-low field.⁶⁰⁻⁶²

Analyte (R ₂)	Catalyst NHC (R ₁)	SABRE-Relay carrier	NMR detection field T (MHz)	SABRE efficiency (<i>E</i>) %
pyridine	IMes	N/A	9.4 (400)	2,6-H (<i>ortho</i>) 6.5 ± 0.1
				3,5-H (<i>meta</i>) 3.9 ± 0.1
				4-H (para) 6.0 ± 0.1
4-methylpyridine	IMes	N/A	9.4 (400)	2,6-H (<i>ortho</i>) 6.4 ± 0.2
				3,5-H (<i>meta</i>) 3.7 ± 0.1
				$4-CH_3$ (<i>methyl</i>) 0.93 ± 0.04
methyl-4,6-d ₂ -nicotinate	IMes	N/A	9.4 (400)	2-H (ortho) 13.6 ± 0.1
				5-H (<i>meta</i>) 15.9 ± 0.3
methyl-4,6-d ₂ -nicotinate	d ₂₂ -IMes	N/A	9.4 (400)	2-H (ortho) 19.4 ± 0.3
				5-H (<i>meta</i>) 21.7 ± 0.5
methyl-4,6-d ₂ -nicotinate	IMes	N/A	1.0 (43)	2-H (ortho) 12.3 ± 0.2
				5-H (<i>meta</i>) 15.9 ± 0.2
1-propanol	IMes	NH₃	9.4 (400)	1-OH 0.57 ± 0.03
				$1-CH_2$ 0.95 ± 0.05
				2-CH ₂ 0.57 ± 0.03
				3-CH ₃ 0.73 ± 0.03

Table 1. SABRE efficiency values for the indicated resonances of the specified analytes as determined by equation 9 and the lines of best fit in Figures 4, 5, and 6.

SABRE hyperpolarisation is generated outside of the NMR spectrometer in a relatively weak polarisation transfer field (typically 0 - 10 mT). Therefore, the polarisation level is expected to be independent of the magnetic field of the NMR spectrometer used for signal detection. Consequently, if all other experimental conditions (e.g. PTF, polarisation transfer time, and sample transfer time) are kept constant, we expect to measure the same SABRE efficiency response at different detection fields. To test this, the dependence of the SABRE polarisation of methyl-4,6-d₂-nicotinate on the $p-H_2$ enrichment was measured using a 1 T (43 MHz) benchtop NMR spectrometer for signal detection. The results in Figure 5 illustrate that, as in the high-field case, a linear trend is found between the observed SABRE polarisation and the p-H₂ enrichment. Furthermore, the calculated efficiency parameters for the two resonances of methyl-4,6-d₂-nicotinate (12.3% and 15.9%) are in good agreement with those measured using a 400 MHz spectrometer for detection (13.6% and 15.9%).

It is important to note here an issue with the quantification of SABRE hyperpolarisation that we have frequently encountered for highly polarised samples. Under highly efficient SABRE conditions and when using high levels of $p-H_2$ enrichment, peak broadening and anti-phase peak character is often observed in SABRE-enhanced ¹H NMR spectra. We believe this to be a result of radiation damping. For the experiments in Figure 4d, carried out on a 400 MHz NMR spectrometer, the spectra acquired at the highest level of $p-H_2$ enrichment gave broad peaks with anti-phase character. However, narrow pure emission peaks were recovered by de-tuning the probe. This was only necessary for the final point in Figure 4d corresponding to 99% p-H₂. The resultant polarisation level, calculated using a reference measurement acquired with the de-tuned probe, followed the linear trend observed at lower p-H₂ enrichment levels (Figure 4d). In the benchtop NMR case (Figure 5), the same solution could not be applied because the

spectrometer cannot be de-tuned. Using a reduced tip-angle of 30°, a SABRE hyperpolarised spectrum was acquired which resulted in a narrow, pure emissive peak for the H-5 proton (red points in Figure 5) with a corresponding polarisation level (calculated using an appropriately acquired reference spectrum) consistent with the linear trend. However, the H-2 resonance (black points in Figure 5) retained anti-phase character. This gives an under-estimation of the polarisation level for the H-2 resonance (black squares) at the highest $p-H_2$ enrichment level in Figure 5, and so this point has been excluded from the linear fit. More details and example spectra are presented in the supporting information document.

Given the linear relationship between analyte polarisation and

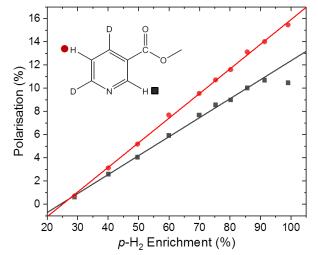


Figure 5. Methyl-4,6- d_2 -nicotinate hyperpolarisation level under SABRE as a function of p-H₂ enrichment. Spectra were acquired on a 1 T benchtop NMR spectrometer (Magritek Spinsolve Carbon) and used the [IrCl(COD)(IMes)] pre-catalyst in methanold₄.The final point at 99 % p-H₂ enrichment was acquired using a 30° pulse for both the SABRE experiment and corresponding thermal reference. The H2 (ortho) position proton resonance still retained some antiphase character and gives a slightly reduced overall polarisation level as a consequence; it is omitted from the linear fit.

 $p-H_2$ enrichment level illustrated herein, we find that a more general solution to this issue for quantitative optimisation of highly efficient SABRE experiments is to use a lower level of $p-H_2$ enrichment. If the level of $p-H_2$ enrichment is known, a straightforward calculation of the efficiency parameter (equation 9) allows for extrapolation to the maximum achievable polarisation level at 100% $p-H_2$ enrichment.

SABRE-Relay

Thus far we have considered the effect of $p-H_2$ concentration on the SABRE technique. However, it is also of interest to investigate the more recent SABRE-Relay method.³⁸ In SABRE-Relay a carrier molecule is hyperpolarised through the standard SABRE mechanism. The polarisation of this carrier is then transferred to the target analyte through a subsequent reversible exchange reaction, such as proton exchange.³⁸ Thus the target substrate molecule becomes hyperpolarised without interacting directly with the SABRE catalyst. In the example presented in Figure 6, NH₃ is used as the carrier and the target molecule is 1-propanol. The solvent, DCM- d_2 , is chosen as it has no exchangeable protons. However, if the solvent is not completely dry, any residual water present can participate in the proton exchange process and also become hyperpolarised.

Consider the SABRE-Relay hyperpolarised ¹H NMR spectrum in Figure 6a. The single absorption peak is assigned to hyperpolarised o-H₂, while the emissive peaks are species in solution that have been hyperpolarised by SABRE (ammonia) or SABRE-Relay (1-propanol and water). We observe two resonances for NH_3 that correspond to carrier molecules bound to the catalyst (2.19 ppm) and in free solution (0.51 In addition, we observe two well-resolved .(mag hyperpolarised peaks of 1-propanol: 1-CH₂ (black square, 3.61 ppm) and 3-CH₃ (blue triangle, 0.97 ppm), and one overlapping peak containing contributions from the 1-OH and 2-CH₂ (red circle, 1.31-1.91 ppm). This set of overlapping peaks also contains a contribution from hyperpolarised water at 1.51 ppm. Using the same approach as for SABRE introduced above, we can evaluate the effect of $p-H_2$ enrichment on the observed polarisation level of these three resonances, with the understanding that the third resonance contains the combined effects of 1-OH and 2-CH₂ of 1-propanol as well as the residual water in the solvent. The results of this study, shown in Figure 6b, indicate that, as in the SABRE case, the relationship between p-H₂ enrichment and SABRE-Relay polarisation is linear. The corresponding SABRE-Relay efficiencies, calculated using equation 9 (see Table 1) are lower than for the analytes polarised directly by SABRE but are nonetheless significant (maximum of 0.94% polarisation). As with the SABRE case, we find that the highest polarisation efficiency is observed for the proton resonance closest to the source of the hyperpolarisation (the exchanging 1-OH proton). Interestingly, the methyl group appears to have a higher efficiency than the 2-CH₂, which is closer to the source of hyperpolarisation. However, this could simply be an artefact of the peak overlap, which acts to reduce the apparent hyperpolarisation of 2-CH₂ due to the lower polarisation efficiencies of the water and the

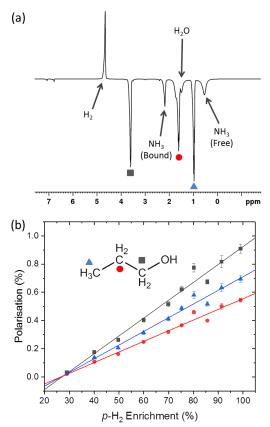


Figure 6. (a) An example SABRE-Relay hyperpolarised ¹H NMR spectrum of 1-propanol. Data was acquired in a single scan on a 400 MHz Bruker NMR spectrometer at 298 K. (b) 1-propanol SABRE-Relay hyperpolarisation level as a function of p-H₂ enrichment when ammonia is the polarisation carrier and [IrCl(COD)(IMes)] the SABRE pre-catalyst in DCM- d_2 .

rapidly exchanging 1-OH proton. SABRE-Relay efficiency is dependent on the concentration of the carrier (ammonia in this case). Measurements of the concentration of ammonia in the SABRE-Relay samples using high-field ¹H NMR showed some variability in the concentration of ammonia across the ten samples (standard deviation of 3.5%). This variability could account for the deviation from the linear relationship observed at 80% *p*-H₂ enrichment in Figure 6. Further details are provided in the supporting information document.

Conclusions

In this work we have explored the polarisation transfer efficiency in SABRE and SABRE-Relay enhanced NMR spectroscopy for a range of experimental conditions. This was achieved by monitoring the observed hyperpolarisation of a target analyte as a function of the level of *para*-enrichment of the H₂ gas used. A linear relationship was found over the range of enrichment levels spanning 29 to 99 % for different target analytes, polarisation transfer catalysts, NMR detection fields, and for both the SABRE and SABRE-Relay polarisation transfer mechanisms. The reproducibility and universality of this linear relationship suggests that the distribution of polarisation within a SABRE hyperpolarised system is in a steady-state and can provide a highly reproducible NMR response for future quantitative applications.

5.

6.

8.

9.

Journal Name

The gradient of each linear correlation was related to a standard theoretical model of $p-H_2$ to define an efficiency parameter, E, that quantifies the fraction of the available $p-H_2$ polarisation that is transferred to the target analyte under the given experimental conditions. It was shown that the SABRE efficiency is independent of the NMR detection field, as expected, and is comparable for analytes with similar chemical properties such as pyridine and 4-methylpyridine. High SABRE efficiencies of up to 21.7% were observed for a selectively deuterated substrate, methyl-4,6-d₂-nicotinate, in conjunction with a highly efficient SABRE catalyst $[Ir(H)_2(methyl-4,6-d_2$ nicotinate)₃(d_{22} -IMes)].⁴⁵ This result supports previous conclusions regarding the benefits of selective deuteration for improved ¹H SABRE hyperpolarisation through the combined effects of extending ¹H hyperpolarisation lifetimes and the concentration of the available hyperpolarisation across fewer ¹H resonances.⁴⁵

The hyperpolarisation of 1-propanol via SABRE-Relay exhibited a polarisation transfer efficiency of up to 0.95%. While significant, this transfer efficiency is an order of magnitude lower than that for direct SABRE hyperpolarisation. Given that SABRE-Relay is a relatively new development, this suggests that there is significant scope for further optimisation. For example, the polarisation transfer conditions have been optimised for the first stage of the SABRE-Relay process (transfer to the carrier) and not the second stage: the relay of polarisation to the target analyte.³⁸

The efficiency measurements were made possible by using a p- H_2 generator that can access interconversion temperatures ranging from 28 K to 140 K, yielding p-H₂ enrichment levels between around 99 % and 28 %. The quantitative relationship between the conversion temperature and the level of $p-H_2$ enrichment was verified experimentally using liquid-state ¹H NMR and the standard theoretical description of p-H₂ and o- H_2 . We have shown that the $p-H_2$ concentration can be accurately and efficiently measured using NMR spectroscopy in a single scan if the hydrogen gas is first dissolved into solution. This provides a sensitivity advantage over the more established method of gas-phase NMR detection, where 256 scans were required.⁴⁰ The amount of dissolved H₂ gas in solution was found to be stable over a 64 hour measurement window, with conversion from the $p-H_2$ back to equilibrium with a relaxation time of ~23 hours. This suggests that the levels of p-H₂ enrichment were constant over the timescales of our SABRE measurements (order of minutes).

Although a clear sensitivity advantage is gained by using the highest available level of p-H₂ enrichment, low-temperature generators like the one detailed herein come with significant costs for both the initial purchase and maintenance. Using the efficiency parameter defined herein, the sensitivity implications of the use of the lower levels of p-H₂ enrichment associated with lower-cost generators can be quantified, and so an appropriate cost-to-sensitivity trade off can be determined for a given application of SABRE-enhanced NMR or MRI.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to thank Dr Victoria Annis for synthesis of the [IrCl(COD)(IMes)] catalyst, and the EPSRC (EP/M020983/1 and EP/R51181X/1) and Wellcome Trust (092506 and 098335) for funding.

Notes and references

- M. H. Levitt, D. Suter and R. R. Ernst, J. Chem. Phys., 1986, 84, 4243-4255.
- 2. L. T. Kuhn, *Hyperpolarization Methods in NMR* Spectroscopy, Springer-Verlag Berlin Heidelberg, 2013.
- J. H. Lee, Y. Okuno and S. Cavagnero, J. Magn. Reson., 2014, 241, 18-31.
- 4. P. Nikolaou, B. M. Goodson and E. Y. Chekmenev, *Chem. Eur. J.*, 2015, **21**, 3156-3166.
 - M. E. Halse, Trends Anal. Chem., 2016, 83, 76-83.
 - C. P. Slichter, Rep. Prog. Phys., 2014, 77, 072501.
- 7. A. Abragam and M. Goldman, *Rep. Prog. Phys.*, 1978, **41**, 395-467.
 - J. H. Ardenkjaer-Larsen, B. Fridlund, A. Gram, G. Hansson, L. Hansson, M. H. Lerche, R. Servin, M. Thaning and K. Golman, *Proc. Natl. Acad. Sci. U.S.A.*, 2003, **100**, 10158-10163.
 - J. N. Dumez, Magn. Reson. Chem., 2017, 55, 38-46.
- G. Zhang and C. Hilty, Magn. Reson. Chem., 2018, DOI: 10.1002/mrc.4735.
- 11. T. G. Walker and W. Happer, *Rev. Mod. Phys.*, 1997, **69**, 629-642.
- 12. B. M. Goodson, J. Magn. Reson., 2002, **155**, 157-216.
- 13. M. L. Hirsch, N. Kalechofsky, A. Belzer, M. Rosay and J. G. Kempf, *J. Am. Chem. Soc.*, 2015, **137**, 8428-8434.
- J. Hovener, A. N. Pravdivtsev, B. Kidd, C. R. Bowers, S. Gloggler, K. V. Kovtunov, M. Plaumann, R. Katz-Brull, K. Buckenmaier, A. Jerschow, F. Reineri, T. Theis, R. V. Shchepin, S. Wagner, N. M. M. Zacharias, P. Bhattacharya and E. Y. Chekmenev, Angew Chem Int Ed Engl, 2018, DOI: 10.1002/anie.201711842.
- 15. J. Natterer and J. Bargon, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1997, **31**, 293-315.
- 16. S. B. Duckett and R. E. Mewis, *Acc. Chem. Res.*, 2012, **45**, 1247–1257.
- C. R. Bowers and D. P. Weitekamp, J. Am. Chem. Soc., 1987, 109, 5541-5542.
- M. G. Pravica and D. P. Weitekamp, *Chem. Phys. Lett.*, 1988, **145**, 255-258.
- R. U. Kirss and R. Eisenberg, J. Organomet. Chem., 1989, 359, C22-C26.
- 20. R. Eisenberg, Acc. Chem. Res., 1991, 24, 110-116.
- 21. P. Hübler, R. Giernoth, G. Kümmerle and J. Bargon, *J. Am. Chem. Soc.*, 1999, **121**, 5311-5318.
- 22. L. T. Kuhn and J. Bargon, in *In situ NMR Methods in Catalysis*, Springer, 2006, pp. 25-68.
- J. López-Serrano, S. B. Duckett, S. Aiken, K. Q. Almeida Leñero, E. Drent, J. P. Dunne, D. Konya and A. C. Whitwood, J. Am. Chem. Soc., 2007, 129, 6513-6527.

- K. V. Kovtunov, V. V. Zhivonitko, I. V. Skovpin, D. A. Barskiy
 and I. V. Koptyug, in *Hyperpolarization Methods in NMR* Spectroscopy, Springer, 2012, pp. 123-180.
- 25. M. Goldman, H. Johannesson, O. Axelsson and M. Karlsson, *Magn. Reson. Imaging*, 2005, **23**, 153-157.
- F. Reineri, A. Viale, G. Giovenzana, D. Santelia, W. Dastru, R. Gobetto and S. Aime, *J. Am. Chem. Soc.*, 2008, **130**, 15047-15053.
- P. Bhattacharya, E. Y. Chekmenev, W. F. Reynolds, S. Wagner, N. Zacharias, H. R. Chan, R. Bunger and B. D. Ross, NMR Biomed., 2011, 24, 1023-1028.
- 28. R. W. Adams, S. B. Duckett, R. A. Green, D. C. Williamson and G. G. Green, *J. Chem. Phys.*, 2009, **131**, 194505.
- 29. P. J. Rayner and S. Duckett, *Angew Chem Int Ed Engl*, 2018, DOI: 10.1002/anie.201710406.
- R. W. Adams, J. A. Aguilar, K. D. Atkinson, M. J. Cowley, P.
 I. P. Elliott, S. B. Duckett, G. G. R. Green, I. G. Khazal, J.
 López-Serrano and D. C. Williamson, *Science*, 2009, **323**, 1708-1711.
- T. Theis, M. L. Truong, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, J. Am. Chem. Soc., 2015, 137, 1404-1407.
- M. L. Truong, T. Theis, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, J. Phys. Chem. C, 2015, 119, 8786-8797.
- M. J. Cowley, R. W. Adams, K. D. Atkinson, M. C. R. Cockett, S. B. Duckett, G. G. R. Green, J. A. B. Lohman, R. Kerssebaum, D. Kilgour and R. E. Mewis, *J. Am. Chem.* Soc., 2011, 133, 6134-6137.
- J. B. Hovener, N. Schwaderlapp, T. Lickert, S. B. Duckett, R.
 E. Mewis, L. A. R. Highton, S. M. Kenny, G. G. R. Green, D.
 Leibfritz, J. G. Korvink, J. Hennig and D. von Elverfeldt, *Nature Communications*, 2013, 4, 2946.
- R. A. Green, R. W. Adams, S. B. Duckett, R. E. Mewis, D. C. Williamson and G. G. Green, *Prog. Nucl. Magn. Reson.* Spectrosc., 2012, 67, 1-48.
- 36. A. Farkas, *Orthohydrogen, Parahydrogen and Heavy Hydrogen*, Cambridge University Press, London, 1935.
- 37. S. Wagner, *Magn. Reson. Mat. Phys. Biol. Med.*, 2014, **27**, 195-199.
- 38. W. Iali, P. J. Rayner and S. B. Duckett, *Sci. Adv.*, 2018, **4**, eaao6250.
- 39. S. S. Roy, K. M. Appleby, E. J. Fear and S. B. Duckett, *J. Phys. Chem. Lett.*, 2018, **9**, 1112-1117.
- 40. B. Feng, A. M. Coffey, R. D. Colon, E. Y. Chekmenev and K. W. Waddell, *J. Magn. Reson.*, 2012, **214**, 258-262.
- 41. C. Terenzi, S. Bouguet-Bonnet and D. Canet, *J. Chem. Phys.*, 2017, **146**, 154203.
- 42. E. B. Ducker, L. T. Kuhn, K. Munnemann and C. Griesinger, J. Magn. Reson., 2012, **214**, 159-165.
- A. N. Pravdivtsev, K. L. Ivanov, A. V. Yurkovskaya, P. A. Petrov, H. H. Limbach, R. Kaptein and H. M. Vieth, J. Magn. Reson., 2015, 261, 73-82.
- D. Blazina, S. B. Duckett, T. K. Halstead, C. M. Kozak, R. J. Taylor, M. S. Anwar, J. A. Jones and H. A. Carteret, *Magn. Reson. Chem.*, 2005, 43, 200-208.
- P. J. Rayner, M. J. Burns, A. M. Olaru, P. Norcott, M. Fekete, G. G. R. Green, L. A. R. Highton, R. E. Mewis and S. B. Duckett, *Proc. Natl. Acad. Sci. U.S.A.*, 2017, **114**, E3188-E3194.

- P. M. Richardson, S. Jackson, A. J. Parrott, A. Nordon, S. B. Duckett and M. E. Halse, *Magn. Reson. Chem.*, 2018, **56**, 641-650.
- 47. B. A. Tom, S. Bhasker, Y. Miyamoto, T. Momose and B. J. McCall, *Rev. Sci. Instrum.*, 2009, **80**, 016108.
- 48. K. Sundararajan, K. Sankaran, N. Ramanathan and R. Gopi, J. Mol. Struct., 2016, **1117**, 181-191.
- A. J. Parrott, P. Dallin, J. Andrews, P. M. Richardson, O. Semenova, M. E. Halse, S. B. Duckett and A. Nordon, *Appl. Spectrosc.*, 2018, doi.org/10.1177/0003702818798644.
- 50. P. Atkins and J. de Paula, *Physical Chemistry*, Oxford; New York:Oxford University Press, 8th edn., 2006.
- 51. D. A. McQuarrie, *Statistical Mechanics*, University Science Books, Sausalito, 2000.
- D. A. Barskiy, A. N. Pravdivtsev, K. L. Ivanov, K. V. Kovtunov and I. V. Koptyug, *Phys. Chem. Chem. Phys.*, 2016, 18, 89-93.
- M. Emondts, J. F. P. Colell, B. Blumich and P. P. M. Schleker, *Phys. Chem. Chem. Phys.*, 2017, **19**, 21933-21937.
- N. Eshuis, B. J. van Weerdenburg, M. C. Feiters, F. P. Rutjes, S. S. Wijmenga and M. Tessari, Angewandte Chemie International Edition, 2015, 54, 1481-1484.
- N. K. J. Hermkens, R. L. E. G. Aspers, M. C. Feiters, F. P. J. T. Rutjes and M. Tessari, *Magn. Reson. Chem.*, 2018, 56, 633-640.
- 56. M. Fekete, O. W. Bayfield, S. B. Duckett, S. Hart, R. E. Mewis, N. Pridmore, P. J. Rayner and A. Whitwood, *Inorganic Chemistry*, 2013, **52**, 13453-13461.
- L. S. Lloyd, R. W. Adams, M. Bernstein, S. Coombes, S. B. Duckett, G. G. Green, R. J. Lewis, R. E. Mewis and C. J. Sleigh, *J. Am. Chem. Soc.*, 2012, **134**, 12904-12907.
- R. E. Mewis, K. D. Atkinson, M. J. Cowley, S. B. Duckett, G. G. Green, R. A. Green, L. A. Highton, D. Kilgour, L. S. Lloyd, J. A. Lohman and D. C. Williamson, *Magn. Reson. Chem.*, 2014, 52, 358-369.
- 59. B. Blümich and K. Singh, *Angew. Chem., Int. Ed. Engl.,* 2017, **56**, 2-17.
- T. Theis, M. P. Ledbetter, G. Kervern, J. W. Blanchard, P. J. Ganssle, M. C. Butler, H. D. Shin, D. Budker and A. Pines, J. Am. Chem. Soc., 2012, 134, 3987-3990.
- R. Borowiak, N. Schwaderlapp, F. Huethe, T. Lickert, E. Fischer, S. Bar, J. Hennig, D. von Elverfeldt and J. B. Hovener, *Magn. Reson. Mat. Phys. Biol. Med.*, 2013, 26, 491-499.
- K. Buckenmaier, M. Rudolph, C. Back, T. Misztal, U. Bommerich, P. Fehling, D. Koelle, R. Kleiner, H. A. Mayer, K. Scheffler, J. Bernarding and M. Plaumann, *Sci. Rep.*, 2017, 7, 13431.