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1	A Phase 3, Randomized, double-blind, multicenter study to EValuate the safety and efficacy of
2	intravenous Iclaprim versus Vancomycin for the trEatment of acute bacterial skin and skin
3	structure infections suspected or confirmed to be due to Gram-positive pathogens: REVIVE-2
4	
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22 Running Head: Iclaprim for acute skin infections

23	Summary: Iclaprim achieved non-inferiority compared with vancomycin at its primary endpoint
24	of early clinical response (defined as a \geq 20% reduction in lesion size at 48-72 hours compared to
25	baseline) in a Phase 3 study for the treatment of acute bacterial skin and skin structure infections.
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31

Abstract

32

33	Iclaprim is a novel diaminopyrimidine antibiotic that may be an effective and safe treatment for
34	serious skin infections. The safety and efficacy of iclaprim were assessed in a global Phase 3,
35	double-blind, randomized, active-controlled trial. Six-hundred thirteen adults with acute bacterial
36	skin and skin structure infections (ABSSSI) suspected or confirmed to be due to Gram-positive
37	pathogens were randomized to iclaprim (80 mg) or vancomycin (15mg/kg), both administered
38	intravenously every 12 hours for 5 - 14 days. The primary endpoint was a \ge 20% reduction in
39	lesion size, compared with baseline, at 48 to 72 hours after the start of administration of study
40	drug in the intent-to-treat population. Among patients randomized to iclaprim, 78.3% (231 of
41	295) met this primary endpoint, compared with 76.7% (234 of 305) for those receiving
42	vancomycin (treatment difference: 1.58%, 95% CI: -5.10% to 8.26%). This met the pre-specified
43	10% non-inferiority margin. Iclaprim was well tolerated, with most adverse events categorized as
44	mild. In conclusion, iclaprim was non-inferior to vancomycin and was well tolerated in this
45	Phase 3 clinical trial for the treatment of acute bacterial skin and skin structure infections. Based
46	on these results, iclaprim may be an efficacious and safe treatment for skin infections suspected
47	or confirmed to be due to Gram-positive pathogens.
48	(This trial has ben registered at ClinicalTrials.gov under identifier NCT02607618.)

49

50 Keywords: iclaprim, vancomycin, acute bacterial skin and skin structure infections

Introduction

52	Acute bacterial skin and skin structure infections (ABSSSI) are common and potentially
53	serious infections that may require hospitalization, intravenous antibiotics, and/or surgical
54	intervention [1,2]. Most are caused by Gram-positive pathogens, including methicillin-resistant
55	Staphylococcus aureus (MRSA), methicillin-susceptible S. aureus (MSSA), and beta-hemolytic
56	streptococci [2]. Currently available treatment options have limitations. New therapeutic options
57	with improved efficacy, safety, and/or pharmacodynamics are needed for ABSSSI [3-4].
58	Iclaprim is a diaminopyrimidine, which inhibits bacterial dihydrofolate reductase, and is active
59	against drug-resistant pathogens [6-9]. Iclaprim demonstrates rapid in vitro bactericidal activity
60	in time-kill studies in human plasma [10]. In the previous Phase 3 clinical trial among patients
61	treated for ABSSSI (REVIVE-1), early clinical responses in the intent-to-treat (ITT) population
62	were 80.9% for iclaprim and 81.0% for vancomycin at the early time point [11]. We report the
63	second Phase 3 study (REVIVE-2) comparing the outcomes of patients treated with either
64	iclaprim or vancomycin for ABSSSI suspected or confirmed to be due to Gram-positive
65	pathogens.
66 67	
68	Methods
69	Study design and participants. REVIVE-2 was a double-blind, multicenter phase 3 non-
70	inferiority trial. Patients were randomized 1:1 to treatment with either iclaprim 80 mg IV q12h
71	(iclaprim) or vancomycin 15mg/kg IV q12h (vancomycin) (NCT02600611). This study design
72	followed both Food and Drug Administration (FDA) and European Medicines Agency (EMA)

guidance. Patients were enrolled between April 2016 and August 2017. The institutional review
board at each site approved the protocol, and all patients or their authorized representative
provided written informed consent.

76 Male and female patients ≥ 18 years of age with suspected or confirmed ABSSSI due to 77 Gram-positive pathogens were eligible for study participation. ABSSSI was defined as a 78 bacterial infection of the skin with a lesion size ≥ 75 cm². ABSSSIs were classified as major 79 cutaneous abscess, pure cellulitis/erysipelas, and/or wound infections (caused by external trauma 80 [e.g., needle sticks or insect bites]), and had the following characteristics: the presence of 81 purulent or seropurulent drainage before or after surgical intervention of a wound or at least 3 of 82 the following signs and symptoms: discharge, erythema (extending at least 2 cm beyond a wound 83 edge in one direction), swelling and/or induration, heat and/or localized warmth, and/or pain 84 and/or tenderness to palpation. Key inclusion and exclusion criteria are listed in Table 1. 85 Iclaprim was administered at 80mg (no hepatic impairment or Child-Pugh A) or 40 mg IV q12h (Child-Pugh B). Child-Pugh C patients were excluded. Vancomycin was administered 86 87 at 15 mg/kg IV and adjusted according to a nomogram with dosing every 12 hours (creatinine 88 clearance [CrCl] ≥50 mL/min), every 24 hours (CrCl ≥35-49 mL/min), every 48 hours (CrCl 89 \geq 25-34 mL/min), or according to vancomycin trough levels (CrCl <25 mL/min) or creatinine 90 clearance. The unblinded pharmacist prepared infusions for patients who were assigned to the 91 vancomycin arm, maintaining the same infusion volume as used for iclaprim. For each patient, 92 the unblinded pharmacist used the creatinine clearance or vancomycin trough levels (to which

the investigator was blinded) to adjust the vancomycin dosage to maintain a trough of 10 - 15 mg/L for patients with an organism with MIC ≤ 1 mg/L, or 15 - 20 mg/L for those with MIC >1

95 mg/L. Both iclaprim and vancomycin were infused over 120 minutes in 500mL normal saline.

96 Normal saline placebo infusions were used to maintain the blind when vancomycin was dosed at97 an interval greater than every 12 hours.

98 The protocol permitted concomitant antibiotic treatment with aztreonam or metronidazole 99 for patients in whom Gram staining of culturable material or cultures indicated Gram-negative 100 and anaerobic bacteria, respectively. Systemic antibiotics (other than aztreonam and 101 metronidazole) or topical antibiotics at the site of the ABSSSI under investigation were 102 prohibited.

Patients received their first dose of randomly allocated study medication within 24 hours after randomization. Study medications were administered for at least 5 days with continuation of treatment up to 14 days at the discretion of the investigator based on the assessment of resolution of signs and symptoms of the ABSSSI. This duration of treatment was in accordance with the Infectious Disease Society of America (IDSA) guidelines [12].

108 Clinical specimens for microbiologic evaluation, including pus from a wound or abscess, 109 and aspirate or skin biopsy from the leading edge of cellulitis, were obtained from patients prior 110 to randomization. At subsequent visits, additional specimens were obtained, for patients with 111 persistent clinical signs or symptoms. Specimens were evaluated by the local microbiology 112 laboratory, and isolates were subcultured and sent to a central microbiology laboratory for 113 confirmation of pathogen identity and MICs. S. aureus genotyping was not performed for this 114 study. In order to increase identification of patients with ABSSSI caused by Streptococcus 115 pyogenes (e.g., cellulitis), leading edge punch biopsies were encouraged for patients with pure 116 cellulitis and serological tests (ASO titers) for all patients were obtained at baseline and at test of 117 cure (2-3 weeks after baseline). A beta-hemolytic streptococci was considered present if the 118 patient had a ABSSSI and an elevated titer of ASO at baseline and/or at test of cure or a four fold

rise of antibody at test of cure from baseline. Two sets of blood samples for aerobic/anaerobic
cultures 10 minutes apart from different sites peripherally were obtained within 24 hours before
the first dose of study drug.

122

123 **Endpoints.** The primary efficacy endpoint of the study was the proportion of patients 124 who achieved an early clinical response (ECR), defined as a $\geq 20\%$ reduction in lesion size 125 compared with baseline at the early time point (ETP) 48-72 hours after the start of administration 126 of the study drug in the intent-to-treat (ITT) population. The secondary endpoints of the study 127 included: (1) clinical cure rate at test of cure (TOC) 7 - 14 days after the last dose of study drug, 128 as measured by both the traditional and a modified composite TOC assessments in the ITT population (see below); and (2) safety and tolerability of iclaprim compared with vancomycin. 129 130 Clinical cure at the TOC visit, conducted 7 - 14 days post-EOT, was evaluated using two 131 prespecified definitions. First, clinical cure at the TOC visit was defined as complete resolution 132 of all signs and symptoms of ABSSSI such that no further antibiotic treatment or surgical 133 procedure were needed. This definition of clinical cure is used for pivotal Phase 3 studies of 134 ABSSSI [5]. Secondly, a modified clinical cure at TOC was also evaluated as a \geq 90% reduction 135 in lesion size compared with baseline, no increase in lesion size since ETP, and no requirement 136 for additional antibiotics (except aztreonam or metronidazole for polymicrobial infections) or 137 unplanned significant surgical procedures after ETP. This modified clinical cure was intended to 138 allow for an additional measure (i.e., 90% reduction in lesion size) similar to the early clinical 139 response (ECR, i.e., 20% reduction in lesion size).

Patients were evaluated at a baseline assessment, then evaluated daily through ETP, and
then every 48 - 72 hours through EOT. Treatment duration was 5 - 14 days based on investigator

142 assessment. Patients were then evaluated at the TOC assessment conducted 7 - 14 days post-

EOT, followed by a late follow-up phone call conducted 28 to 32 days after the first dose (Figure1).

Safety was assessed by Common Terminology Criteria for reported treatment emergent
adverse events (TEAEs), serious adverse events (SAEs), hematology, clinical chemistry, liver
function tests, coagulation, urinalysis, vital signs, physical examinations, and electrocardiograms
(ECGs).

149 **Statistical analysis.** The statistical analyses evaluated the efficacy and safety of iclaprim 150 compared with vancomycin. Statistical tests for efficacy analyses were two-sided, and at the 151 level of significance alpha = 0.05. Confidence intervals (CIs) were calculated as a two-wided 152 95% confidence interval. Continuous data were summarized by treatment group using the 153 number of patients in the analysis population (N), mean, standard deviation (SD), median, and 154 range, and categorical data were summarized by treatment group using N and percentage. 155 Demographics and baseline characteristics were summarized using descriptive statistics. The 156 primary efficacy analysis was performed in the ITT population. Secondary analyses were 157 performed in the ITT predefined populations that included diabetes, mild, moderate and severe 158 renal impairment. By-patient and by-pathogen bacteriological outcomes at EOT and TOC were 159 presented as frequency distributions of outcomes by treatment group for patients with a 160 confirmed Gram-positive pathogen at baseline. The safety population was defined as all 161 randomized patients who received at least one dose of study medication. The incidence of 162 TEAEs was summarized at the overall patient level, Medical Dictionary for Regulatory 163 Activities (MedDRA) version 20.0 system organ class level, and preferred term level. Separate 164 tabulations were provided by severity and relationship to study medication and for SAEs.

165 Laboratory data, vital signs and ECGs were evaluated by presentation of summary statistics of166 raw data and changes from baseline.

167	Six hundred patients (approximately 300 per treatment group) randomized (1:1) were
168	targeted for this study. Using Farrington and Manning's method for non-inferiority (NI) testing
169	with a 1 sided alpha of 0.025, assuming a 75% ECR rate in each group and a 10% non-inferiority
170	bound delta, a sample size of 295 ITT patients per treatment group was required for 80% power.
171	
172	Results
173	Demographics
174	The study randomized 613 patients, and 600 fulfilled criteria for the ITT population, the
175	prespecified efficacy population for FDA, from 40 study sites in 10 countries. Figure 2 shows the
176	disposition of patients. The patients lost to followup in each treatment group were similar to
177	other patients randomized in the severity of their ABSSSIs (5 for iclaprim and 6 for
178	vancomycin). There were 13 patients randomized in error; these were identified prior to
179	unblinding and were not included in the ITT analysis. Of these, 6 had lesions that did not meet
180	study entry criteria (lesion size <75 cm ²) and 7 were unable or unwilling to follow study
181	procedures. The baseline and demographic characteristics of patients treated with either iclaprim
182	or vancomycin were comparable (Tables 2 and 3). The proportion of patients with fever at
183	baseline in the iclaprim and vancomycin cohorts were 27.1% and 26.2%, respectively. The
184	baseline mean (S.D.) lesion sizes of patients in the iclaprim and vancomycin cohorts were 372.3
185	(305.8) cm ² and 357.0 (271.1) cm ² , respectively. Treatment groups were similar for baseline
186	ABSSSI categories, laboratory parameters, vital signs, physical examinations, X-rays, and ECG
187	evaluations. In addition, no notable differences were observed between treatment groups with

respect to prior medications and treatments or study drug compliance were observed. Both the iclaprim and vancomycin treatment groups had a median of 7 treatment days (range: 5 - 14days).

191

192 Efficacy Results

193 Primary Endpoint

In the ITT population, an ECR was reported at ETP for 78.3% (231/295) of patients in
the iclaprim group and for 76.7% (234/305) of patients in the vancomycin group (% difference:
1.58; 95% Confidence Interval (CI): -5.10, 8.26)) (Table 3). A sensitivity analysis adding the 13
excluded patients showed similar results (iclaprim 76.5%, vancomycin 76.2%, % difference
0.25, 95% CI: -6.48, 6.98).

199

200 Secondary Analyses

In the ITT population, the clinical cure rates at TOC were 77.6% (229/295) and 77.7%

202 (237 of 305) for patients treated with iclaprim and vancomycin group, respectively (%

203 difference: -0.08; 95% CI: -6.74, 6.59) (Table 4). Using a modified clinical cure TOC analysis

204 defined by a \ge 90% reduction in lesion size compared with baseline, no increase in lesion size

since ETP and no requirement for additional antibiotics, clinical cure was observed in 71.5% and

206 70.5% of patients receiving iclaprim and vancomycin, respectively (treatment difference: 1.03%,

207 95% CI: -6.23% to 8.29%). The ECR at ETP was comparable for the iclaprim and vancomycin

208 groups among the ITT predefined populations by lesion type, pathogen, diabetes, mild, moderate

and severe renal impairment (Table 4).

For the microbiological outcome at EOT and TOC, 384 (64.0%) patients presented with a

culture-confirmed Gram-positive pathogen at baseline. S. aureus was the most commonly isolated pathogen (N=258) of which 138 (53.4%) were MRSA (Table 2). The MIC_{50}/MIC_{90} values for iclaprim and vancomycin for S. aureus isolates were 0.12 / 0.5 mcg/mL and 1 / 1 mcg/mL, respectively.

215

216 Safety Results

217 Study drug-related TEAEs, treatment emergent SAEs, and deaths among patients in the 218 iclaprim and vancomycin treatment groups are shown in Table 5. The treatment emergent 219 adverse events leading to discontinuation were 5.4% and 5.6% in the iclaprim and vancomycin 220 group, respectively. Similar incidence of nausea (5.7% and 5.6%), infusion site extravasation 221 (4.3% and 4.0%), diarrhea (2.7% and 3.6%), and headache (2.3% and 4.3%), were reported in 222 patients in the iclaprim group compared to vancomycin group, respectively. Although not an a 223 priori hypothesis, there were no study-drug related TEAE related to nephrotoxicity reported for 224 patients treated with iclaprim compared to 2(0.7%) for vancomycin. Per the protocol, 225 nephrotoxicity was predefined as an increase in serum creatinine of 0.5 mg/dL or 50% above 226 baseline for at least two consecutive days. The serum creatinine change from baseline to TOC 227 was 0.7 and 7.7 µmol/L (0.008 and 0.09 mg/dL) in patients in the iclaprim group compared to 228 vancomycin group, respectively. There were no significant differences between treatment groups 229 in mean values or mean changes in other routine serum laboratory parameters, urinalysis results, 230 vital signs or physical examinations during treatment, or at EOT, TOC and follow-up between 231 treatment groups. Eleven (3.7%) patients in the iclaprim group, and nine (3.0%) patients in the 232 vancomycin group had increases in ALT or AST values to >3X upper limit of normal (ULN) 233 during treatment. Three patients (one in the iclaprim and two in the vancomycin group) had a

234	diagnosis of acute hepatitis A confirmed by IgM serology. Two of those patients (one in the
235	iclaprim and one in the vancomycin group) had bilirubin increases >2X ULN. These increases
236	resolved to baseline values upon discontinuation of drug in all patients. No subject met Hy's law
237	criteria in this study.
238	One (0.4%) patient in the iclaprim group and 0 patients in the vancomycin group had
239	QTcF intervals >500 msec (i.e., 503 msec) or increased by >60 msec compared with baseline.
240	The QTc prolongation was not reported as an adverse event and resolved to baseline values upon
241	discontinuation of drug.
242	
243	Discussion
244	In this study, iclaprim was non-inferior to vancomycin in the treatment of ABSSSI
245	suspected or confirmed to be caused by Gram-positive organisms, based on the primary endpoint
246	of early clinical response. This Phase 3 study clinical trial also met its secondary endpoints,
247	demonstrating that the clinical cure rates at TOC, both the traditional and the modified composite
248	TOC, were similar for patients treated with iclaprim and vancomycin. Similar treatment
249	outcomes were also noted across a priori identified subgroups. No notable differences in the
250	incidence of TEAEs between the treatment groups were observed.
251	Results in REVIVE-2 were broadly similar to those of REVIVE-1, an identically-
252	designed trial in which iclaprim also achieved non-inferiority to vancomycin. Taken together,
253	these results suggest that iclaprim is efficacious and safe for treatment of serious skin infections
254	suspected to be due to Gram-positive pathogens.
255	In contrast to previous cSSSI studies, a fixed iclaprim dose was used in this study. This
256	fixed dose of iclaprim was selected because, compared to the weight-based dosing regimen used

in the previous Phase 3 studies [4], the fixed dose maximizes by 30% the AUC/MIC and time 257 258 above MIC, the parameters most closely associated with efficacy in animal infection models, 259 while reducing by 10% the steady-state C_{max} (C_{max/ss}), a parameter associated with QTc 260 prolongation in Phase 1 studies. In this study, there was only one patient who received iclaprim 261 (0.3%) with subsequent QTc prolongation. Consequently, the fixed dose of iclaprim may be 262 important especially in patients with borderline QTc prolongation, diabetes, obesity and 263 decreased renal function. No dosage adjustments of iclaprim are needed in these populations. 264 Currently recommended agents for treatment of moderate to severe Gram-positive skin 265 infections include vancomycin, linezolid or daptomycin [12]. Safety issues or resistance to these 266 agents are reported among patients treated for MRSA infections [15-20]. The results of this 267 study, in combination with those of REVIVE-1, suggest that iclaprim may be a useful addition to 268 the treatment armamentarium. Advantages of iclaprim are that it does not appear nephrotoxic, 269 does not require dose adjustments for renal impairment, and does not require therapeutic drug 270 monitoring.

A strength of this Phase 3 study is that greater than 40% of randomized patients had wound infections. This group of infections is typically more difficult to cure compared to abscesses and cellulitis, and their inclusion enhances generalizability of study findings to this important population.

There are limitations to this Phase 3 study. First, 67.5% (405 out 600) of enrolled patients in this study were from the United States, 29.7% (178 of 600) from Europe, and 2.8% (17 of 600) from Latin America. A high proportion of injection drug users (~50%) were included in both treatment groups. Therefore the results may not be generalizable to other practice settings. Second, data on vancomycin trough concentrations were not not analyzed at the

280 central laboratory and local laboratory trough values are not available. However, based on 281 adherence to the prespecified vancomycin dosing nomogram, greater than 95% of patients had 282 the correct dosing interval for this antibiotic, including those patients with renal impairment 283 (creatinine clearance <75 mL/min), for whom the initial dosing interval was based on renal 284 clearance. Third, vancomycin was used instead of a beta-lactam drug for MSSA when obtained 285 from ABSSSI. Compared to vancomycin, beta-lactam drugs are likely a superior agent for 286 MSSA. Fourth, leading edge biopsies and ASO titers were measured to determine GAS etiology 287 of ABSSSI. These diagnostic methodologies are not specific for GAS and may overestimate the 288 true frequency of GAS for ABSSSI.

In conclusion, in this Phase 3 study, iclaprim was non-inferior to vancomycin with respect to the early clinical response at an early time point in the treatment of ABSSSI caused or suspected to be caused by Gram-positive organisms. These results suggest iclaprim may serve as an alternative option for treatment of ABSSSI caused by Gram-positive pathogens, including drug-resistant bacteria. In hospitalized ABSSSI patients with co-morbidities such as renal impairment and/or diabetes, iclaprim may provide advantages over vancomycin due to the fixed dose regimen and absence of nephrotoxicity.

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407 Table 1: Key inclusion and exclusion criteria

Inclusion Criteria	Exclusion Criteria
Provision of informed consent	ABSSSI of the following categories: severely impaired arterial blood supply such that amputation of the infected anatomical site was likely, more than one abscess, infected diabetic foot ulcers, infected decubitus ulcers, infected human or animal bites, necrotizing fasciitis or gangrene, uncomplicated skin or skin structure infection, self-limiting infections
Evidence of systemic involvement as defined by having at least 1 of the following conditions within 24 hours of randomization considered to be pathogen-related:	Skin and/or skin structure infection that could be treated by surgery alone
Fever (>38°C/100.4°F orally, rectally, or tympanically)	Infections associated with a prosthetic device, and suspected or confirmed osteomyelitis or septic arthritis or endocarditis
Enlarged and/or tender proximal lymphadenopathy and/or lymphangitis	Known or suspected concurrent infection or conditions requiring systemic anti-microbial treatment, prophylaxis, or suppression therapy
Elevated total peripheral white blood cells (WBCs) >10,000/mm ³	Known or suspected human immunodeficiency virus (HIV)-infected patients with a cluster of differentiation (CD4) count <200 cells/mm ³ recorded in the last 30 - 60 days; absolute neutrophil count (ANC) <500 cells/mm ³ ; organ transplant within the preceding 6 months; requirement for corticosteroids >20 mg/day prednisolone or equivalent, or received corticosteroids >20 mg per day prednisolone or equivalent in the past 3 days
>10% immature neutrophils (bands) regardless of total peripheral WBC count	Cardiovascular conditions and treatments: patients known to have congenital or sporadic syndromes of QTcF prolongation; type I A or III anti-arrhythmic drugs; nonsustained ventricular tachycardia (NSVT) defined as >10 consecutive ventricular beats at a rate of >120 beats per minute (bpm) with a duration of <30 seconds, bradycardia (<40 bpm), and QT/QTcF interval outside the normal range defined as: QTcF >500 msec

Elevated C-reactive protein	Received more than one dose of a short-acting (i.e., q12h dosing or
	less) systemic antibiotic active against Gram-positive pathogens
	within the last 7 days, unless there was documented evidence of
	treatment failure or demonstrated resistance of Gram-positive
	pathogens to the prior antibiotic therapy.

Characteristics	Iclaprim (n=295)	Vancomycin (n=305)
Age (yr), mean (SD) median	50.0 (15.65)	50.8 (15.03)
Gender, no. (%)		
Female	103 (34.9)	108 (35.4)
Male	192 (65.1)	197 (64.6)
Race, no. (%)		
White	267 (90.5)	276 (90.5)
Black	12 (4.1)	11 (3.6)
American Indian or Alaska Native	2 (0.7)	3 (1.0)
Native Hawaiian or other Pacific Islander	1 (0.3)	3 (1.0)
Multi-racial	0	2 (0.7)
Other	13 (4.4)	10 (3.3)
Weight (kg), mean (SD)	84.2 (20.78)	85.5 (22.17)
Geographic region, no. (%)		
United States	200 (67.8)	205 (67.2)
Europe	84 (28.5)	94 (30.8)
Latin America	11 (3.7)	6 (2.0)
Severe Infections ^a	185 (62.7)	198 (64.9)
Lesion Type		
Major Cutaneous Abscess, no. (%)	53 (18.0)	45 (14.8)

408 Table 2: Baseline and demographic characteristics among the ITT population by treatment

Cellulitis / Erysipelas, no. (%)	115 (39.0)	125 (41.0)
Wound Infection, no. (%)	127 (43.1)	135 (44.3)
Mean lesion Size, cm ² (SD)	372.3	357.00
	(305.752)	(271.077)
Comorbidities, no. (%)		
Diabetes	36 (12.2)	36 (11.8)
Renal Impairment, CrCL		
(ml/min)		
60-89	35 (12.0)	53 (17.9)
30-59	17(5.8)	13 (4.4)
<=29	2 (0.7)	2(0.7)
Illicit drug use	144 (48.8)	160 (52.5)
Fever (oral temperature	80 (27.1)	80 (26.2)
>38°C/100.4°F), no. (%)		
Leukocytes (per mm ³), mean (SD)	9.5 (3.4)	9.4 (3.8)
median (min, max)	9.2 (1.7,	8.4 (2.9,
	22.2)	23.1)
Baseline microbiology, no. (%)		
Exclusively Gram-positive pathogens	170 (89.5)	167 (86.1)
Mixed Gram-positive and	20 (10.5)	27 (13.9)
Gram-negative	_= (1000)	(2007)
Concomitant aztreonam use, no. (%)	13 (4.4)	20 (6.6)
Concomitant metronidazole use, no. (%)	9 (3.1)	11 (3.6)

^aSevere infections defined as an infection at baseline with one or more of the following criteria: 409

410 fulfilled the published definition for systemic inflammatory response syndrome (SIRS) by

411 having ≥ 2 of the following findings: body temperature $\geq 38^{\circ}$ C or $\leq 36^{\circ}$ C, heart rate ≥ 90 bpm,

respiration rate >20 breaths/minute, and WBC >12000/mm³ or <4000/mm³ or >10% bands; 412

413	evaluated as having severe tenderness or severe erythema at the infection site; and/or Positive
414	blood cultures at baseline.
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429	Table 3: Microbiological character	istics at study entry for the	TTT population by treatment
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Pathogen	Iclaprim (n=295)	Vancomyc4a0 (n=305)
Positive ABSSSI Culture	199 (72.9)	431 214 (73.5)
Staphylococcus aureus		432
MRSA	69 (23.4)	69 (22.6) ₄₃₃
MSSA	60 (20.3)	60 (19.7) 434
Beta-hemolytic Streptococci	76 (26.1)	93 (28.6)
Positive Blood Culture at Baseline, no. (%) ^a	7/274 (2.6)	435 13/283 (4.6)
Infection Site Pathogen, no. (%)		436
Multiple	21 (11.0)	26 (13.4) ⁴³⁷
Single	149 (78.4)	141 (72.7)

439 ^a In the iclaprim group, there were S. aureus (N=2), S. epidermidis (N=2), and 1 each of S.

443 aureus and S. salivarius, and 1 patient with both S. epidermidis and M. luteus.

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⁴⁴⁰ agalactiae, S. dysagalactiae, and Micrococcus luteus. In the vancomycin group, there were S.

⁴⁴¹ aureus (N=3), and 1 each of S. epidermidis, S. hominis, S. massiliensis, S. anginosus, S.

⁴⁴² salivarius, Bacillus spp (non-anthracis), Atopobium parvulum, as well as 1 patient with both S.

447 Table 4: Clinical responses for primary endpoint and secondary analyses in the ITT population

448 by treatment

Clinical Responses	Iclaprim (n=295)	Vancomycin (n=305)	Treatment Difference (%; 95% Confidence Interval)
Primary Endpoint			
Early Clinical Response (ECR) at Early Time Point (ETP) in ITT, no. (%) - Total	231 (78.3)	234 (76.7%)	1.58 (-5.10, 8.26)
Early Clinical Response (ECR) at Early Time Point (ETP) in ITT, no. (%) - US	173/200 (86.5)	164/205 (80.0)	6.50 (-3.35, 16.14)
Early Clinical Response (ECR) at Early Time Point (ETP) in ITT, no. (%) – EU and LA	58/95 (61.1)	70/100 (70.0)	-8.9 (-24.02, 5.11)
Secondary Analyses			
ECR at ETP among major cutaneous abscess, no. (%)	45 (84.9)	40 (88.9)	-3.98 (-17.29, 9.33)
ECR at ETP among cellulitis / erysipelas, no. (%)	81 (70.4)	91 (72.8)	-2.37 (-13.79, 9.05)
ECR at ETP among wound infections, no. (%)	105 (82.7)	103 (76.3)	6.38 (-3.35, 16.12)
ECR at ETP among MRSA infected, no./total no. (%)	61/69 (88.4)	53/69 (76.8)	11.59 (-5.80, 28.48)

ECR at ETP among MSSA infected, no./total no. (%)	50/60 (83.3)	51/60 (85.0)	-1.67 (-20.15, 16.90)
ECR at ETP among S. pyogenes infected, no./total no. (%)	64/76 (84.2)	74/93 (79.6)	4.6 (-4.29, 11.07)
ECR at ETP among diabetics, no./total no. (%)	26/36 (72.2)	29/36 (80.6)	-8.33 (-31.95, 15.99)
ECR at ETP among mild renal impairment (creatinine clearance 60-89 ml/min), no./total no. (%)	27/35 (77.1)	39/53 (73.6)	3.56 (-17.38, 24.72)
ECR at ETP among moderate and severe renal impairment (creatinine clearance <60 ml/min), no./total no. (%)	13/19 (68.4)	11/15 (73.3)	-12.2 (-45.42, 24.19)
Clinical cure at TOC, no. (%)	229 (77.6%)	237 (77.7%)	-0.08 (-6.74, 6.59)
Modified clinical cure ^a at TOC, no. (%)	211 (71.5%)	215 (70.5%)	1.03% (-6.23 to 8.29)

449 ^aModified clinical cure defined as a \geq 90% reduction in lesion size compared to baseline, no

450 increase in lesion size since ETP, and no requirement for additional antibiotics (except

451 aztreonam or metronidazole) or unplanned significant surgical procedures after ETP.

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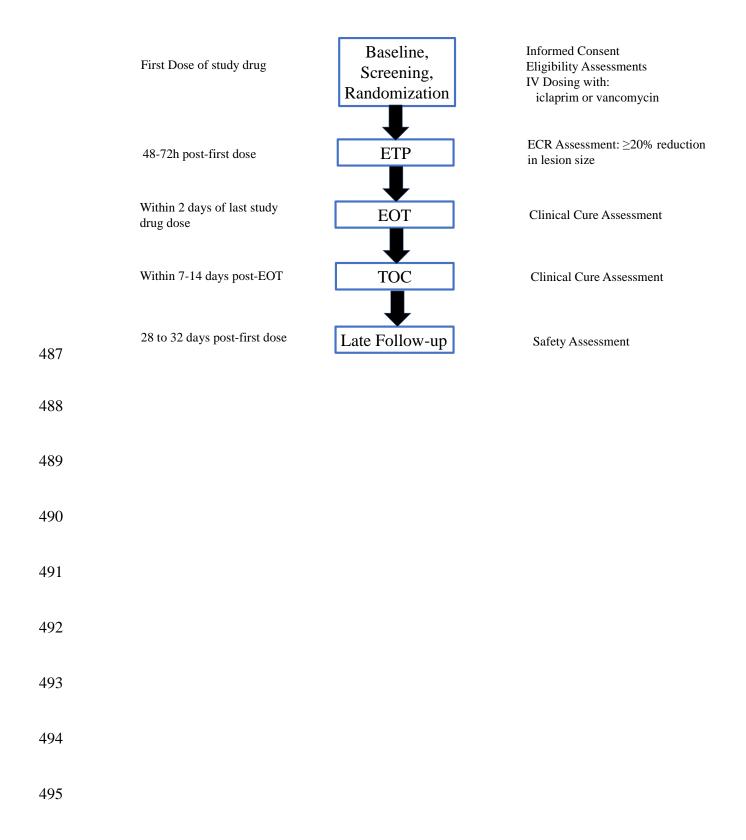
455 Table 5: Safety Parameters by Treatment

Category	Iclaprim (N=299)	Vancomycin (N=302)
Any drug-related TEAE ^a , no. (%)	140 (46.8%)	133 (44.0%)
Study drug related TEAE, no. (%)	42 (14.0%)	39 (12.9%)
TEAE leading to discontinuation of study drug, no. (%)	16 (5.4%)	17 (5.6%)
TEAE related SAEs ^a , no. (%)	14 (4.7%)	12 (4.0%)
Mean QTcF prolongation, msec (SD)	9.9 (14.6)	3.8 (16.3)
Mean serum creatinine change from baseline to TOC, umol/L (SD)	0.7 (18.0)	7.7 (39.8)
Mean serum creatinine change from baseline to TOC, mg/dL (SD)	0.008 (0.20)	0.09 (0.45)
Nephrotoxicity	0	2 (0.7)
Deaths, no. (%)	0	1 (0.3)
TEAE by system organ class, no. (%)		
Nausea	17 (5.7)	17 (5.6)
Infusion site extravasation	13 (4.3)	12 (4.0)
Hypokalemia	6 (2.0)	11 (3.6)
Diarrhea	8 (2.7)	11 (3.6)
Vomiting	7 (2.3)	7 (2.3)
Pyrexia	7 (2.3)	5 (1.7)
Hypertension	7 (2.3)	5 (1.7)

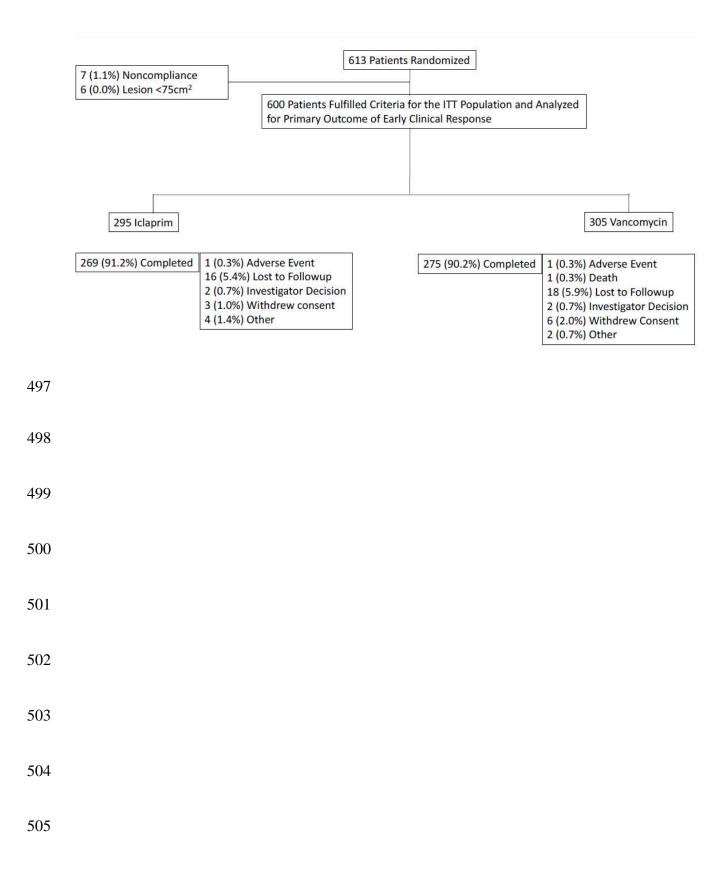
Headache	7 (2.3)	13 (4.3)
Anemia	6 (2.0)	6 (2.0)
Increased AST ^b	6 (2.0)	5 (1.7)
Increased ALT ^b	5 (1.7)	7 (2.3)
Pruritis	2 (0.7)	7 (2.3)

- 457 Note: The order of the TEAE by system organ class was listed in the order of most frequent (top)
- 458 to least frequent (bottom) for iclaprim.
- 459 ^aAbbreviations: TEAE, treatment emergent adverse events; SAE, severe adverse event; ALT,
- 460 Alanine aminotransferase; AST, aspartate aminotransferase
- 461 ^bInvestigator reported

470	Figure Legend
471	Figure 1: Schedule of visits. Abbreviations: IV, intravenous; ECR, early clinical response; ETP,
472	early time point; EOT, end of therapy; TOC, test of cure
473	Figure 2: Disposition of patients
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496 Figure 2: Disposition of Patients



Conflict of Interest

508	T.H. has received consultancy fees from Basilea Pharmaceutica, Genentech, Medicines
509	Company, and Motif Biosciences, and grant support from Basilea and Achaogen. TMF has
510	served as a consultant for Motif BioSciences, Allergan, Medicines Company, Merck, Nabriva,
511	Paratek, and Cempra. AT has served as a consultant for Motif BioSciences. AFS has served as a
512	consultant to, received research support from, or been a speaker for: Abbott, Actavis, Alios,
513	Astellas, AstraZeneca, Bayer, BMS, Cardeas, Medicines Company, Merck, Pfizer, Roche,
514	Tetraphase, Theravance, and Wockhardt Pharma. MHW has received consulting fees from
515	Abbott Laboratories, Actelion, Astellas, Astra-Zeneca, Bayer, Biomèrieux, Cerexa, Cubist,
516	Durata, The European Tissue Symposium, The Medicines Company, MedImmune, Merck, Motif
517	Biosciences, Nabriva, Optimer, Paratek, Pfizer, Qiagen, Roche, Sanofi-Pasteur, Seres, Summit,
518	and Synthetic Biologics; lecture fees from Abbott, Alere, Astellas, Astra-Zeneca, Merck, Pfizer
519	& Roche; grant support from Abbott, Actelion, Astellas, Biomèrieux, Cubist, Da Volterra,
520	MicroPharm, Morphochem AG, Sanofi-Pasteur, Seres, Summit and The European Tissue
521	Symposium, Merck. PH is a former employee of Arpida. MD has received speaker's and/or
522	consultancy fees from AstraZeneca, Bayer, Janssen-Cilag, Motif BioSciences, Novartis, Pfizer,
523	and Merck, Sharp & Dohme Corp., a subsidiary of Merck & Co., Inc. R.C. has received
524	consultancy fees from Cempra Pharmaceuticals, PRA International, Furiex Pharmaceuticals,
525	Inimex Pharmaceuticals, Dr. Reddy's Laboratories, Cubist Pharmaceuticals, Cerexa/Forest
526	Laboratories, AstraZeneca, GlaxoSmithKline, Pfizer, Merck, Trius Therapeutics, ContraFect,
527	Theravance, and Astellas Pharma and served on an advisory board for Pfizer, Polymedix, Trius
528	Therapeutics, Rib-x Pharmaceuticals, Seachaid Pharmaceuticals, BioCryst Pharmaceuticals,

- 529 Durata Therapeutics, Achaogen, Gilead Sciences, ContraFect, Cempra, and Nabriva
- 530 Therapeutics. P.M. is an employee of Covance. R.C. received research grants from Theravance,
- 531 Innocoll, and The Medicines Company. RS and DBH are employees of Motif BioSciences.