# SCIENTIFIC REPORTS

Received: 15 March 2017 Accepted: 21 December 2017 Published online: 18 January 2018

## **OPEN** The theoretical molecular weight of NaYF<sub>4</sub>:RE upconversion nanoparticles

Lewis E. Mackenzie 1,2, Jack A. Goode<sup>2</sup>, Alexandre Vakurov<sup>2</sup>, Padmaja P. Nampi<sup>3</sup>, Sikha Saha<sup>4</sup>, Gin Jose<sup>3</sup> & Paul A. Millner<sup>2</sup>

Upconversion nanoparticles (UCNPs) are utilized extensively for biomedical imaging, sensing, and therapeutic applications, yet the molecular weight of UCNPs has not previously been reported. Herein, we present a theory based upon the crystal structure of UCNPs to estimate the molecular weight of UCNPs: enabling insight into UCNP molecular weight for the first time. We estimate the theoretical molecular weight of various UCNPs reported in the literature, predicting that spherical NaYF4 UCNPs ~ 10 nm in diameter will be ~1 MDa (i.e. 10<sup>6</sup> g/mol), whereas UCNPs ~ 45 nm in diameter will be ~100 MDa (i.e. 10<sup>8</sup> g/mol). We also predict that hexagonal crystal phase UCNPs will be of greater molecular weight than cubic crystal phase UCNPs. Additionally we find that a Gaussian UCNP diameter distribution will correspond to a lognormal UCNP molecular weight distribution. Our approach could potentially be generalised to predict the molecular weight of other arbitrary crystalline nanoparticles: as such, we provide stand-alone graphic user interfaces to calculate the molecular weight both UCNPs and arbitrary crystalline nanoparticles. We expect knowledge of UCNP molecular weight to be of wide utility in biomedical applications where reporting UCNP quantity in absolute numbers or molarity will be beneficial for inter-study comparison and repeatability.

Photonic upconversion nanoparticles (UCNPs) have garnered widespread scientific interest due to their unique near infra-red (NIR) excitation and visible luminescence properties; a process known as photonic upconversion. UCNPs are inorganic crystalline nanostructures (typically NaYF<sub>4</sub>) co-doped with rare-earth (RE) ions, (e.g. Yb<sup>3+</sup>,  $Er^{3+}$ ,  $Gd^{3+}$ ); hereby referred to in general terms as NaYF<sub>4</sub>:RE UCNPs. The RE ions act as sensitizers and emitters for photonic upconversion of multiple infra-red photons, resulting in visible luminescence emission. UCNP emission is highly stable<sup>1</sup>, with no photo bleaching, and a relatively long luminescence emission lifetime ranging from hundreds of microseconds to a few milliseconds<sup>2,3</sup>. NIR excitation via upconversion is highly advantageous for biomedical applications, where ultraviolet or visible excitation of fluorophores (e.g. dyes, proteins, or quantum dots) is normally required, with the associated challenges of photo-bleaching and photo-toxicity. Interactions between nearby molecules and the UCNPs crystal structure enables molecular biosensing via luminescence resonance energy transfer (LRET) between UCNPs and molecules in proximity to them<sup>4-10</sup>. As such, UCNPs have found wide utility in biomedical applications, including as imaging contrast labels in cellulo, in vivo, and ex  $vivo^{5,11-19}$ ; as biosensors for detection of antibiotics<sup>20</sup> and toxins in food<sup>21-23</sup>; as biosensors to measure biomarkers in biological fluids (e.g. whole blood, serum, urine)<sup>6-8,24-26</sup>, and as therapeutic agents, against targets such as cancer cells<sup>27,28</sup>. Additionally UCNPs have been applied to nanoscale thermometry<sup>29,30</sup> and photovoltaic applications<sup>31,32</sup>. However, to date, the molecular weight of UCNPs has not been reported: as such, both the molarity of UCNPs in solution, and the absolute number of UCNPs in any given sample has been unknown.

The lack of molecular weight information for UCNPs is a considerable shortcoming in biomedical applications of UCNPs, where precise quantification of UCNP concentration would be highly beneficial for informing of dosage of UCNPs studies, as well as aiding inter-study comparison. Additionally, quantification of UCNP molarity and absolute

<sup>1</sup>Department of Chemistry, Faculty of Sciences, Durham University, Durham, DH1 4ED, United Kingdom. <sup>2</sup>School of Biomedical Sciences, Faculty of Biological Sciences, University of Leeds, Leeds, LS2 9JT, United Kingdom. <sup>3</sup>School of Chemical and Process Engineering, Faculty of Engineering, University of Leeds, Leeds, LS2 9JT, United Kingdom. <sup>4</sup>Leeds Institute of Cardiovascular and Metabolic Medicine (LICAMM), Faculty of Medicine and Health, University of Leeds, Leeds, LS2 9JT, United Kingdom. Correspondence and requests for materials should be addressed to L.E.M. (email: Lewis.E.MacKenzie@Durham.ac.uk)





number of UCNPs would be highly beneficial when constructing biosensors where the ratio of UCNPs compared to other molecules, e.g. antibodies<sup>6–8,25</sup> or oligonucleotides<sup>33</sup>, and as such, is important for informing biosensor design.

The lack of information on UCNP molecular weight is likely due to lack of experimental techniques capable of measuring the molecular weight of large macromolecules such as UCNPs. Using the theory we present in this paper, we predict that the molecular weight of NaYF<sub>4</sub>:RE UCNPs will range from a few mega Daltons (MDa) (i.e.  $10^{6}$  g/mol) for very small UCNPs (~10 nm in diameter), to >100 MDa for UCNPs with a more typical diameter of ~45 nm. This large molecular weight range is well beyond the measurement limits of laboratory techniques such as mass spectrometry and sedimentation velocity analytical ultracentrifugation (svUAC), which are limited to <40 kDa and <5 MDa respectively<sup>34</sup>. Despite this intrinsic limitation, we attempted to employ svAUC to estimate the molecular weight of UCNPs ~30 nm in diameter (which we estimate to correspond to a molecular weight of ~40 MDa), but reliable measurements were not obtained (see the supplementary material and Discussion sections for details).

In this study, we present a theoretical method- based upon the extensively studied and empirically proven theory of crystallography and UCNP structure- to calculate the molecular weight UCNPs, accounting for UCNP composition and morphology. In brief, the crystalline structure of UCNPs is quantified by transmission electron microscopy (TEM), and x-ray diffraction (XRD) experiments. From this information, the total atomic weight within a single NaYF<sub>4</sub>:RE unit cell, and the total number of unit cells within a UCNP can be calculated. Thus, the theoretical molecular weight of UCNPs can be calculated by summing up the total molecular weight contained within all unit cells in a UCNP.

We anticipate that this theoretical framework could be extended to crystalline nanoparticles of arbitrary morphology and composition, provided that the crystalline structure of such nanoparticles are known. As such, we also provide two stand-alone graphical user interfaces (GUIs) for simple calculation of the molecular weight of both NaYF<sub>4</sub>:RE UCNPs and arbitrary crystalline nanoparticles. Knowledge of UCNP molecular weight will likely be highly beneficial for quantification of UCNP concentration in biomedical applications.

### Theory

**Crystalline structure and photonic upconversion properties of UCNPs.** The key to understanding both the optical properties of UCNPs, and their molecular weight, lies in the crystalline structure of UCNPs. UCNPs consist of a crystal lattice made up of repeating crystal unit cells of NaYF<sub>4</sub>, with a fraction of  $Y^{3+}$  ions selectively replaced by RE dopants (see Fig. 1). In UCNPs, photonic upconversion is enabled by the absorption of two or more near-infrared photons, which, via excitation of several long-lived metastable electron states, and subsequent non-radiative multi-phonon and radiative relaxation, produces luminescence emission at visible wavelengths (see Fig. 2). Efficient upconversion requires the crystalline host lattice to be doped with multiple species of lanthanide ions (typically Yb<sup>3+</sup> and Er<sup>3+</sup>), where one lanthanide ion species acts as a photo-sensitizer (typically Yb<sup>3+</sup>) and the other acts as a photonic emitter (typically Er<sup>3+</sup>)<sup>35</sup>. Although many different combinations of lattice and RE dopants have been explored<sup>36</sup>, the combination of Yb<sup>3+</sup> and Er<sup>3+</sup> within a NaYF<sub>4</sub> host lattice has been found to provide high upconversion efficiency, and as such, is commonly used for UCNPs<sup>37,38</sup>. Figure 2





------

Study	NaYF <sub>4</sub> RE dopant composition (%)	UCNP lattice structure	a (Å)	c (Å)	Mean UCNP diameter (nm)	UCNP diameter range (nm)
Sikora <i>et al</i> . <sup>14</sup> .	30% Yb <sup>3+</sup> , 2% Er <sup>3+</sup>	Cubic	5.51	_	~ 30	15-70
Cao et al. <sup>15</sup> .	20% Yb <sup>3+</sup> , 2% Er <sup>3+</sup>	Hexagonal	5.960	3.510	33±1	32-34
Wang et al. <sup>40</sup> .	18% Yb <sup>3+</sup> , 2% Er <sup>3+</sup>	Hexagonal	5.96	3.53	Not reported	Not reported
Wang et al. <sup>40</sup> .	18% Yb <sup>3+</sup> , 2% Er <sup>3+</sup> , 60% Gd <sup>3+</sup>	Hexagonal	6.02	3.60	Not reported	Not reported

Table 1. Crystal lattice parameters of NaYF<sub>4</sub>:RE UCNPs reported in the literature.

shows an exemplar upconversion emission spectrum of NaYF<sub>4</sub>:Yb,Er cubic UCNPs (20% Yb<sup>3+</sup>, 2% Er<sup>3+</sup>) and the corresponding Jablonski diagram for upconversion<sup>39</sup>.

NaYF<sub>4</sub>:RE unit cells are either a cubic or a hexagonal crystal lattice arrangement (see Fig. 1). In the face-centred cubic lattice arrangement (Na<sub>2</sub>Y<sub>2</sub>F<sub>8</sub>), high-symmetry cation sites are formed, and are randomly occupied by either Na<sup>+</sup> or RE<sup>3+</sup> ions (see Fig. 1a), and Y<sup>3+</sup> ions are substituted for other RE<sup>3+</sup> ions, enabling photonic upconversion. In hexagonal unit cells (Na<sub>1.5</sub>Y<sub>1.5</sub>F<sub>6</sub>), there are two relatively low-symmetry cation sites, which contain either Na<sup>+</sup> or RE<sup>3+</sup> ions (see Fig. 1b)<sup>40</sup>. Characterisation of UCNP unit cells is typically conducted by XRD measurements. Several studies have reported the crystal lattice parameters associated with cubic and hexagonal NaYF<sub>4</sub>:RE UCNPs: these are summarised in Table 1. Wang *et al.*<sup>40</sup> report unit cell parameters for cubic ( $\alpha$  phase) and hexagonal ( $\beta$  phase) unit NaYF<sub>4</sub>:RE unit cell configurations (see Fig. 1). The arrangement of ions within unit cells influences the crystal lattice parameters, consequently changing photonic properties, such as upconversion quantum efficiency<sup>40</sup>.

Synthesis of NaYF<sub>4</sub>:RE UCNPs typically creates pseudo-spherical UCNPs with a range of diameters. For example, Sikora *et al.*, (2013) report a Gaussian diameter distribution of UCNPs, ranging between 15–70 nm<sup>14</sup>. and Haro-González *et al.*<sup>41</sup> report a Gaussian diameter distribution of UCNPs UCNPs ranging from ~10–50 nm in diameter (see Table 1).

**Estimating the number of unit cells in a UCNP.** For the purposes of this study, we assume UCNPs to be spherical, with volume ( $V_{UCNP}$ ) described by:

$$V_{UCNP} = \frac{4}{3}\pi r^3,\tag{1}$$

where *r* is the radius of the UCNP. Note that non-spherical UCNP morphologies can be incorporated by substituting Equation 1 to describe non-spherical volumes. If the UCNP consists of cubic unit cells, then the volume of an individual cubic unit cell ( $uV_{cubic}$ ) is given by:

$$iV_{cubic} = a_c^3. (2)$$

If the UCNP consists of hexagonal unit cells, volume of a hexagonal unit cell  $(uV_{hexagonal})$  is given by:

$$aV_{hexagonal} = \frac{2\sqrt{3}}{4}a_h^2 c_h.$$
(3)

Where  $a_h$  and  $c_h$  are lattice parameters describing hexagonal unit cells. Thus, the number of unit cells in a UCNP (i.e.  $uN_{cubic}$  or  $uN_{hexagonal}$ ) can be estimated by:

$$uN_{cubic} = V_{UCNP}/uV_{cubic},\tag{4}$$

$$uN_{hexagonal} = V_{UCNP}/uV_{hexagonal}.$$
(5)

This calculation assumes the effects of crystal dislocations and rounding error in the total number of unit cells to be negligible, and that lattice parameters are accurate. Further, we assume that UCNPs are composed of 100% cubic or hexagonal unit cells because, to the best of our knowledge, hybrid crystal phase UCNPs have not been reported.

**Estimating the total atomic weight within a single unit cell.** Assuming no RE dopants, the atomic weight of a single cubic NaYF<sub>4</sub> ( $uAW_{cubic}$ ) or hexagonal NaYF<sub>4</sub> unit cell ( $uW_{hex}$ ) is described by:

$$uAW_{cubic} = (2 \times AW_{Na}) + (2 \times AW_{Y}) + (8 \times AW_{F});$$
(6)

$$uAW_{hex} = (1.5 \times AW_{Na}) + (1.5 \times AW_{Y}) + (6 \times AW_{F});$$
<sup>(7)</sup>

where  $AW_{Na}$ ,  $AW_Y$ , and  $AW_F$  are the atomic weight (Da or g/mol) of Sodium, Yttrium, and Fluorine respectively (see Table S1). We assume any mass difference due loss of electrons due to ionisation to be negligible. If RE dopant ions are added during UCNP synthesis, then a fraction of Y<sup>3+</sup> ions are substituted for RE<sup>3+</sup> dopant ions, altering the average atomic weight of unit cells within UCNPs. This RE doping can be accounted for by defining a total additive factor (*af*):

$$af = fRE_{d1} + fRE_{d2} \dots + fRE_{dn},\tag{8}$$

where  $RE_{d1}$ ,  $fRE_{d2}$ , ...,  $fRE_{dn}$  is the fractional percentage of an arbitrary number (*n*) of RE dopants. The total additive factor is a numeric value ranging between 0 and 1, representing the theoretical extremes of 0% and 100% substitution of Y respectively. Thus, total the atomic mass contained within a single cubic or hexagonal unit cell with RE dopants is be calculated by:

$$uAW_{cubic} RE_{Doped} = (2 \times AW_{Na}) + (8 \times AW_F) + (2(1 - af) \times AW_Y) + (2 \times fRE_{d1} \times AW_{RE_{d1}}) + (2 \times fRE_{d2} \times AW_{RE_{d2}}) + ... + (2 \times fRE_{dn} \times AW_{RE_{dn}});$$
(9)

$$A W_{hexagonal} R E_{Doped} = (1.5 \times A W_{Na}) + (6 \times A W_F) + (1.5(1 - af) \times W) + (1.5 \times f R E_{d1} \times A W_{RE_{d1}}) + (1.5 \times f R E_{d2} \times A W_{RE_{d2}}) + ... + (1.5 \times f R E_{dn} \times A W_{RE_{dn}});$$
(10)

where  $uAW_{cubic} RE_{Doped}$  and  $uAW_{hexagonal} RE_{Doped}$  are the average atomic weight of RE doped cubic and hexagonal unit cells, respectively.

**Estimating the theoretical molecular weight of a UCNP.** Once the total number of unit cells within a UCNP (uN) and the total atomic weight (uAW) within each individual unit cell are estimated, the theoretical molecular weight of a cubic lattice UCNP (MWcubic) can be estimated by summing the atomic weight contributions from all unit cells:

$$MW_{cubic} = uAW_{cubic} RE_{Doped} \times uN_{cubic}, \tag{11}$$

$$MW_{hexagonal} = uAW_{hexagonal} RE_{Doped} \times uN_{hexagonal}.$$
(12)

From Equations 4, 5, 11, and 12, it can be seen that the molecular weight of UCNPs scales proportionally to volume, thus spherical UCNPs molecular weight will scale proportionally to the cube of UCNP radius.

#### Methods

ı

**Molecular weight predictions for cubic and hexagonal NaYF<sub>4</sub>:RE UCNPs.** Using the theory presented in Sections 2.4–2.6, the theoretical molecular weight of hexagonal and cubic lattice NaYF<sub>4</sub> UCNPs were calculated, assuming the following typical unit cell lattice parameters: cubic: a = 5.51 Å; hexagonal: a = 5.91 Å, c = 3.53 Å; (see Table 1).

**The effect of RE doping on theoretical molecular weight.** The effect of RE doping was investigated by using the theory presented in Sections 2.4–2.6 to calculate the theoretical molecular weight of NaYF<sub>4</sub>:RE UCNPs incorporating various concentrations of Yb<sup>3+</sup> and Er<sup>3+</sup> dopant ions. We assume that UCNP lattice parameters will remain constant, neglecting the unit cell contraction effect demonstrated by Wang *et al.*<sup>40</sup>, where UCNP unit cell lattice parameters are altered when the concentration of RE dopants is increased<sup>40</sup>.



**Figure 3.** Theoretical molecular weight versus diameter hexagonal and cubic NaYF<sub>4</sub> UCNPs (green and blue respectively). (a) UCNP diameter vs. molecular weight plotted on a standard scale. (b) The same data plotted with a logarithmic scale. Lattice parameters were assumed to be: a = 5.51 Å for cubic UCNPs; a, c = 5.91 Å and 3.53 Å for hexagonal UCNPs.

The theoretical molecular weight of UCNPs reported in the literature. The theoretical molecular weight of various  $NaYF_4$ : RE UCNPs reported in the literature was calculated by incorporating various lattice parameters and mol% of RE dopants from the literature into the theory presented in Sections 2.4–2.6.

**UCNP diameter distribution vs. theoretical molecular weight distribution.** UCNP synthesis typically produces a Gaussian distribution of UCNPs diameters. To investigate how such a distribution of UCNP diameters affects the distribution of theoretical UCNP molecular weights, the Gaussian diameter distribution data for a single batch of NaYF<sub>4</sub>:Yb,Er UCNPs was reproduced from data presented in Sikora *et al.*<sup>14</sup>. The theoretical molecular weight for each UCNP diameter in this distribution was calculated by the theory presented in Sections 2.4–2.6. Gaussian fits to the data were calculated by using non-linear least squares fitting in MATLAB (MATLAB 2016a, MathWorks).

**Stand-alone GUIs for calculation of nanoparticle theoretical molecular weight.** Two stand-alone executable graphic user interfaces (GUIs) were created to enable quick and simple calculation of nanoparticle molecular weight. The GUIs were written in MATLAB but do not require an installation MATLAB to run (see supplementary material for more information). Each GUI incorporates different features and assumptions. The first, more simple, GUI was developed to enable other researchers to calculate the theoretical molecular weight of spherical NaYF<sub>4</sub>:RE UCNPs for a user-defined nanoparticle size range. The second, more powerful, GUI was designed to enable users to estimate the theoretical molecular weight of crystalline nanoparticles with arbitrary nanoparticle geometry; arbitrary lattice parameters; and arbitrary elemental composition, across a user-defined range of characteristic nanoparticle sizes. Additional technical information for both GUIs is provided in the supplementary material section. The stand-alone GUIs developed are shown in supplemental Figures S1 and S2. These GUIs are freely available from the University of Leeds Research Data Depository and are attributed with their own citable (https://doi.org/10.5518/173)<sup>42</sup>.

#### Results

**Theoretical molecular weight of cubic and hexagonal NaYF<sub>4</sub>:RE UCNPs.** Hexagonal lattice UCNPs were found to have a greater theoretical molecular weight than cubic lattice UCNPs (see Fig. 3); this is due to the lower volume of hexagonal unit cells, and correspondingly higher density of hexagonal lattice UCNPs. Additionally, because molecular weight scales proportionally with UCNP volume, relatively small changes in UCNP diameter increased molecular weight considerably: e.g. a 20 nm cubic UCNP has a molecular weight of ~10 MDa, whereas a 30 nm UCNP has a molecular weight in excess of 30 MDa (an increase of >20 MDa for a 10 nm change in UCNP diameter).

**The effect of RE doping on UCNP molecular weight.** Increasing  $Yb^{3+}$  or  $Er^{3+}$  dopant % increased the theoretical molecular weight of UCNPs (see Fig. 4) because  $Yb^{3+}$  and  $Er^{3+}$  have a greater atomic mass than  $Y^{3+}$ . However, the difference in theoretical molecular weight between UCNPs doped with  $Yb^{3+}$  and  $Er^{3+}$  was relatively small due to the similar atomic weight of  $Yb^{3+}$  and  $Er^{3+}$  (173.054 and 167.259 g/mol respectively, see Table S1). Hexagonal lattice UCNPs show a slightly higher increase in theoretical molecular weight for a given dopant concentration than cubic lattice UCNPs because hexagonal lattice UCNPs have a greater unit cell density compared to their cubic counterparts.



**Figure 4.** The effect of RE doping on theoretical UCNP molecular weight. (**a**,**b**) theoretical molecular weight vs. RE dopant mol% for cubic lattice UCNPs. (**c**,**d**) theoretical molecular weight vs. RE dopant mol% for cubic lattice UCNPs. Calculations assume that lattice parameters are a = 5.51 Å for cubic lattice UCNPs, a = 5.91 Å; c = 3.53 Å for hexagonal lattice UCNPs, and that lattice parameters are independent of dopant mol%.

**The theoretical molecular weight of NaYF<sub>4</sub>:RE UCNPs reported in the literature.** The theoretical molecular weight of various NaYF<sub>4</sub>:RE UCNPs reported in the literature are shown in Fig. 5.

**UCNP diameter distribution vs. theoretical molecular weight distribution.** The UCNP diameter distribution data from Sikora *et al.*<sup>14</sup> was well-fitted by a Gaussian distribution ( $R^2 = 0.96$ ) (see Fig. 6a). The corresponding theoretical molecular weight distribution (shown in Fig. 6b), demonstrates the exponential relation between UCNP diameter and UCNP molecular weight distribution. Plotted on a logarithmic x-axis scale (Fig. 6c), the resulting molecular weight distribution was well fitted by a Gaussian distribution ( $R^2 = 0.98$ ), indicating that the molecular weight distribution corresponding to a Gaussian diameter distribution is lognormal.

### Discussion

We have provided a theory to estimate the molecular weight of UCNPs. Our theory is required because, to the best of our knowledge, there are no experimental techniques capable of measuring the molecular weight of UCNPs, which we predict will be >5 MDa for UCNPs ~15 nm in diameter, and ~100 MDa for UCNPs ~45 nm in diameter. Mass spectrometry is limited to molecules <40 kDa, and svAUC is limited to measurements of macromolecules <5 MDa<sup>34</sup>.

Despite the aforementioned challenges of experimental verification, we attempted svAUC measurements of UCNPs, because successful svAUC studies of other types of nanoparticles (e.g. SiO<sub>2</sub> nanoparticles) with unknown molecular weight have been reported by others<sup>43,44</sup>. If accurate svAUC measurements of UCNPs could be made, then UCNP molecular weight could potentially be calculated and verified by the theory described by Carney *et al.*<sup>34</sup>, which is based upon accurate quantification of sedimentation and diffusion coefficients from svAUC measurements, and which has been verified for gold nanoparticles ~2 MDa in molecular weight. The full details of the method of our svAUC experiment are provided in the supplementary information. However, our svAUC experiment studying UCNPs was not successful. In brief, our avAUC results showed that the UCNPs (diameter =  $32 \pm 5$  nm, average theoretical molecular weight of ~ 43 MDa) sedimented very rapidly, even at low centrifuge rotor speeds (3,000 rpm), limiting the amount of useable data. At higher rotor speeds UCNPs sedimented to zero sample



**Figure 5.** Theoretical molecular weight of various UCNPs reported in the literature. (**a**) Sikora *et al.*<sup>14</sup>. (**b**) Cao *et al.*<sup>15</sup>. (**c**,**d**) Wang *et al.*<sup>40</sup>.

concentration, a negative sedimentation coefficient was returned. Additionally, UCNPs were observed to diffuse considerably, further complicating AUC experiments. This unusual behaviour is not typical of the nanoclusters and gold nanoparticles used to demonstrated the molecular weight estimation technique described by Carney *et al.*<sup>34</sup>, and as such UCNP molecular weight could not be estimated by svAUC. Our theory predicts that for UCNPs to be suitable for svAUC analysis, i.e. <5 MDa, requires UCNPs to be <10 nm in diameter. Such small UCNPs have been synthesised by others<sup>45</sup>, but such UCNPs were unfortunately, not available for svAUC testing at this stage. The various challenges associated with svAUC measurement of UCNPs serve to further highlight the need for a method to estimate the molecular weight of UCNPs theoretically.

Although it has not been possible for us at this stage to experimentally validate our estimates of UCNP molecular weight, it may be possible in future to verify some limited predictions of our theory. For example, it may be possible to measure the difference in bulk densities of cubic and hexagonal UCNPs and compare this with predictions from our theory. However, we could not attempt this measurement at this stage because we did not have access to the high temperature crucible equipment required for hexagonal UCNP synthesis<sup>40</sup>.

Despite this lack of current and direct experimental verification, we can be reasonably confident in the accuracy of our theory because it stems directly from the theory of crystallography, which has been a subject of intense study in the past century<sup>46</sup>, combined with empirical measurements of UCNP crystal structure.

Our method to calculate the theoretical molecular weight of NaYF<sub>4</sub>:RE UCNPs relies on two basic assumptions: 1. that UCNPs are crystals of homogenous elemental composition and homogeneous unit cell structure, and 2. that the lattice parameters and diameter data utilized is accurate. These assumptions can be verified by TEM and XRD measurements of UCNP crystal structure. Ensuring accurate lattice parameters is particularly important when estimating the molecular weight of UCNPs with arbitrarily large dopant concentrations. For example, Wang *et al.*<sup>40</sup> experimentally demonstrated that by doping a hexagonal phase NaYF<sub>4</sub>:Yb,Er UCNP (18% Yb, 2% Er) with increasing concentrations of Gd<sup>3+</sup> increases the lattice parameters of the UCNP significantly, resulting in increased unit cell volume. The influence of dopant concentration on crystal lattice parameters can be estimated by Vegard's law<sup>47</sup>, but direct verification of lattice parameters is always preferable to ensure accuracy in molecular weight estimation. If the dependence of lattice parameter on RE dopant percentage is not accounted for, then large errors in the UCNP molecular weight estimation could arise.





UCNP volume/morphology also influences theoretical UCNP molecular weight. We recommend using TEM to directly quantify UCNP morphology with limited assumptions. Other techniques such as such as dynamic light scattering (DLS) and nanoparticle tracking analysis can be used to estimate the equivalent hydrodynamic radius of nanoparticles but these techniques incorporate various assumptions into calculations which can result in systematic innacuracy<sup>44,48</sup>. As such, direct TEM imaging of UCNPs is preferable to ensure theoretical molecular weight is as accurate as possible. In this study we assumed UCNPs are perfectly spherical, but our method could be trivially adapted for arbitrary nanoparticle geometries; e.g. rods<sup>40,49</sup>, triangular<sup>50</sup>, or prism-shaped<sup>51</sup> nanoparticles, and for nanoparticles of varying crystalline composition. The extension of our technique to arbitrary geometries, arbitrary crystal lattice parameters, and arbitrary elemental composition is demonstrated by the development and application of an advanced GUI incorporating all of these variables (see Figure S2). Our theory does not account for any

dislocations in the regular UCNP crystal structure. Instead we assume the influence of any such dislocations to be negligible compared to the molecular weight of whole UCNPs. Our theory also does not account for any surface functionalisation of UCNPs. Thus the molecular weight of UCNPs modified by addition of a silica<sup>8,35,52</sup>, calcium fluoride<sup>53</sup>, or any surfactants or ligands will be greater than that predicted by our theory.

It should be noted that a simple theory for estimation of the molecular weight of a single homogenous gold nanoparticle based upon bulk density of materials was proposed by Lewis *et al.*<sup>54</sup>. However, this simple theory did not account for crystalline unit cell parameters or elemental doping. Further, their theory was not extended to describe the molecular weight distributions of a population of nanoparticles. Our results demonstrate that a Gaussian distribution of UCNP diameters corresponds to a lognormal distribution in molecular weight (as shown in Fig. 6). Mathematically, it is reasonable to expect similar logarithmic relations between UCNP diameter and molecular weight for arbitrary diameter distributions. Such molecular weight distributions may of consequence when studying behaviour of UCNP populations, because minor outliers in UCNP diameter will be extreme outliers in terms of molecular weight.

Estimation of molecular weight of NaYF<sub>4</sub>:RE UCNPs will likely be of utility in various applications, particularly in biomedical imaging, biosensing, and therapeutics. Knowledge of UCNP molecular weight will likely be of great utility in studies where UCNP surfaces are functionalised with additional molecules, e.g. antibodies<sup>6–8,25</sup> or oligonucleotides<sup>33</sup>, because If the molecular weight of UCNPs is known, then the molar concentrations of substances in the functionalisation processes can be determined. When combined with estimation of UCNP surface area, this could inform the UCNP functionalisation for biosensing applications. Knowledge of UCNP molecular weight would also be beneficial in the processing of particles for downstream applications. In particular, steps taken to functionalise the nanoparticles may require separation procedures to remove unreacted moieties or unwanted reactants. If the molecular weight of UCNPs were known, then it may be beneficial for the optimisation of conjugation stoichiometry, which can be concentration dependant; the reaction rates of UCNPs will be heavily influenced by their molecular weight; thus a greater understanding of their molecular weight may increase the knowledge of thermodynamic properties of UCNP systems. This is particularly important when considering the use of bio-receptors with UCNPs where the mass of the particle may affect the binding kinetics of the UCNP-receptor construct.

The molecular weight of UCNPs will also be of interest in the study of cytotoxicity, bio-distribution, cellular uptake, metabolism, and excretion of UCNPs in biological systems<sup>12,14</sup>. Currently, it is extremely challenging to compare the results from various imaging and therapeutic studies because UCNP concentration is reported as weight of UCNPs per volume of aqueous media (e.g. mg/mL or similar)<sup>12</sup>. This is a crude measure which does not quantify number of UCNPs in a given sample. For example, nanoparticles can induce membrane damage<sup>55</sup> and initiate apoptosis (programmed cell-death)<sup>56,57</sup>. Reporting the molar concentration of UCNPs would help assessment of UCNP cytotoxic effects. A standardised protocol based on molecular weight of UCNPs would help assessment of accumulation of UCNPs *in vivo* and their clearance time from organs<sup>13</sup> or tumours<sup>53</sup>. Reporting the molar concentration of UCNP composites may also help to develop highly-localised targeted delivery of therapeutic drugs to the required sites in the body, leading to better controlled targeted photodynamic therapy<sup>27</sup>, and potential improvements in targeted drug delivery<sup>16</sup>.

#### Conclusions

We have provided a method to estimate the theoretical molecular weight of UCNPs. This theory is based upon UCNP crystal parameters which can be measured for batches of UCNPs by TEM and XRD techniques. The theory presented here is generalizable to other crystalline nanoparticles where the relevant crystalline lattice parameters are known, i.e. nanoparticle unit cell elemental composition, unit cell size parameters, and nanoparticle morphology. To enhance application of our theory we provide two stand-alone GUIs for calculation of the molecular weight of both UCNPs and arbitrary crystalline nanoparticles respectively. We could not, however, experimentally verify our predictions of UCNP molecular weight with mass spectrometry or svAUC due to the fundamental limitations of this technique. Nevertheless, our theory provides some key predications about the molecular weight of UCNPs. Firstly, that the theoretical molecular weight of UCNPs scales with volume of the nanoparticle. As an example, we predict that a spherical UCNP ~10 nm diameter will have a molecular weight of  $\sim 1$  MDa (10<sup>6</sup> g/mol), whereas a UCNP  $\sim 45$  nm in diameter will be  $\sim 100$  MDa (10<sup>8</sup> g/mol). From this relation, we find that a Gaussian distribution of nanoparticle diameters corresponds to a lognormal distribution of UCNPs molecular weights, and that a small change in UCNP diameter distribution can potentially represent a large change in overall UCNP molecular weight. We also report that Hexagonal crystal lattice phase UCNPs will be of greater molecular weight than cubic lattice phase UCNPs, and that increasing RE dopant % will increase UCNP molecular weight, with the caveat that unit cell lattice parameters may change due to doping.

We expect that the knowledge of UCNP molecular weight will be of utility in a wide variety of biomedical applications, as UCNP concentrations can now be reported in terms of molarity or absolute number of UCNPs instead of the relatively crude measure of sample weight per volume. This will likely aid inter-study comparison of both UCNP dosage and improve methods for creating UCNP biosensors.

#### References

- 1. Zhou, J., Xu, S., Zhang, J. & Qiu, J. Upconversion luminescence behavior of single nanoparticles. Nanoscale 7, (2015).
- Hyppänen, I., Höysniemi, N., Arppe, R., Schaeferling, M. & Soukka, T. Environmental Impact on the Excitation Path of the Red Upconversion Emission of Nanocrystalline NaYF<sub>4</sub>:Yb<sup>3+</sup>, Er<sup>3+</sup>. J. Phys. Chem. C acs.jpcc. 7b01019 https://doi.org/10.1021/acs. jpcc.7b01019 (2017).
- Plohl, O. et al. Optically Detected Degradation of NaYF4: Yb, Tm Based Upconversion Nanoparticles in Phosphate Buffered Saline Solution. Langmuiracs.langmuir. 6b03907 https://doi.org/10.1021/acs.langmuir.6b03907 (2016).

- Chen, F., Bu, W., Cai, W. & Shi, J. Functionalized upconversion nanoparticles: versatile nanoplatforms for translational research. *Curr. Mol. Med.* 13, 1613–32 (2013).
- Wang, F., Banerjee, D., Liu, Y., Chen, X. & Liu, X. Upconversion nanoparticles in biological labeling, imaging, and therapy. *Analyst* 135, 1839–1854 (2010).
- Tang, J., Lei, L., Feng, H., Zhang, H. & Han, Y. Preparation of K+-Doped Core-Shell NaYF4:Yb, Er Upconversion Nanoparticles and its Application for Fluorescence Immunochromatographic Assay of Human Procalcitonin. J. Fluoresc. 26, 2237–2246 (2016).

 Lei, L. et al. A rapid and user-friendly assay to detect the Neutrophil gelatinase-associated lipocalin (NGAL) using up-converting nanoparticles. *Talanta* 162, 339–344 (2017).

- Jo, E. J., Mun, H. & Kim, M. G. Homogeneous Immunosensor Based on Luminescence Resonance Energy Transfer for Glycated Hemoglobin Detection Using Upconversion Nanoparticles. Anal. Chem. 88, 2742–2746 (2016).
- 9. Doughan, S., Uddayasankar, U. & Krull, U. J. A paper-based resonance energy transfer nucleic acid hybridization assay using upconversion nanoparticles as donors and quantum dots as acceptors. *Analytica Chimica Acta* **878**, (Elsevier B.V., 2015).
- Zhang, S. et al. Fluorescence resonance energy transfer between NaYF4:Yb, Tm upconversion nanoparticles and gold nanorods: Near-infrared responsive biosensor for streptavidin. J. Lumin. 147, 278–283 (2014).
- 11. Mader, H. S., Kele, P., Saleh, S. M. & Wolfbeis, O. S. Upconverting luminescent nanoparticles for use in bioconjugation and bioimaging. *Curr. Opin. Chem. Biol.* 14, 582–596 (2010).
- 12. Gnach, A., Lipinski, T., Bednarkiewicz, A., Rybka, J. & Capobianco, Ja Upconverting nanoparticles: assessing the toxicity. *Chem. Soc. Rev. Chem. Soc. Rev.* 44, 1561–1584 (2015).
- Zou, R. et al. Silica shell-assisted synthetic route for mono-disperse persistent nanophosphors with enhanced in vivo recharged nearinfrared persistent luminescence. Nano Res. https://doi.org/10.1007/s12274-016-1396-z (2017).
- 14. Sikora, B. et al. Transport of NaYF4:Er3+, Yb3+ up-converting nanoparticles into HeLa cells. Nanotechnology 24, 235702 (2013).
- Cao, T. et al. Water-soluble NaYF4:Yb/Er upconversion nanophosphors: Synthesis, characteristics and application in bioimaging. Inorg. Chem. Commun. 13, 392–394 (2010).
- Ma, Y. et al. Labeling and long-term tracking of bone marrow mesenchymal stem cells in vitro using NaYF4:Yb3+, Er3+ upconversion nanoparticles. Acta Biomater. 42, 199–208 (2016).
- 17. Kostiv, U. *et al.* RGDS- and TAT-Conjugated Upconversion of NaYF4:Yb3+/Er3+ &SiO2 Nanoparticles: *In Vitro* Human Epithelioid Cervix Carcinoma Cellular Uptake, Imaging, and Targeting. *ACS Appl. Mater. Interfaces* 8, (20422–20431 (2016).
- Shi, Y. et al. Stable Upconversion Nanohybrid Particles for Specific Prostate Cancer Cell Immunodetection. Nat. Publ. Gr. 1–11 https://doi.org/10.1038/srep37533 (2016).
- Rao, L. et al. Erythrocyte Membrane-Coated Upconversion Nanoparticles with Minimal Protein Adsorption for Enhanced Tumor Imaging. Appl. Mater. Interfaces 2159–2168 https://doi.org/10.1021/acsami.6b14450 (2017).
- Hu, G. et al. Upconversion Nanoparticles and Monodispersed Magnetic Polystyrene Microsphere Based Fluorescence Immunoassay for the Detection of Sulfaquinoxaline in Animal-Derived Foods. J. Agric. Food Chem. 64, 3908–3915 (2016).
- Dai, S., Wu, S., Duan, N. & Wang, Z. A luminescence resonance energy transfer based aptasensor for the mycotoxin Ochratoxin A using upconversion nanoparticles and gold nanorods. *Microchim. Acta* 183, 1909–1916 (2016).
- Guo, X., Wu, S., Duan, N. & Wang, Z. Mn 2+-doped NaYF 4: Yb/Er upconversion nanoparticle-based electrochemiluminescent aptasensor for bisphenol A. Anal. Bioanal. Chem. 408, 3823–3831 (2016).
- Chen, Q., Hu, W., Sun, C., Li, H. & Ouyang, Q. Synthesis of improved upconversion nanoparticles as ultrasensitive fluorescence probe for mycotoxins. Anal. Chim. Acta 938, 137–145 (2016).
- 24. Fu, X., Chen, L. & Choo, J. Optical Nanoprobes for Ultrasensitive Immunoassay. Anal. Chem. 1, 124-137 (2016).
- Gao, N., Ling, B., Gao, Z., Wang, L. & Chen, H. Near-infrared-emitting NaYF4:Yb, Tm/Mn upconverting nanoparticle/gold nanorod electrochemiluminescence resonance energy transfer system for sensitive prostate-specific antigen detection. *Anal. Bioanal. Chem.* https://doi.org/10.1007/s00216-017-0212-2 (2017).
- Juntunen, E. et al. Effects of blood sample anticoagulants on lateral flow assays using luminescent photon-upconverting and Eu(III) nanoparticle reporters. Anal. Biochem. 492, 13–20 (2016).
- Liang, L. et al. Facile Assembly of Functional Upconversion Nanoparticles for Targeted Cancer Imaging and Photodynamic Therapy. ACS Appl. Mater. Interfaces 8, acsami.6b00713 (2016).
- Yang, X. et al. Synthesis of a core/satellite-like multifunctional nanocarrier for pH- and NIR-triggered intracellular chemothermal therapy and tumor imaging. RSC Adv. 7, 7742–7752 (2017).
- Geitenbeek, R. G. et al. Na YF<sub>4</sub>:Er<sup>3+</sup>, Yb<sup>+</sup>/SiO<sub>2</sub> Core/Shell Upconverting Nanocrystals for Luminescence Thermometry up to 900 K. J. Phys. Chem. C acs.jpcc.6b10279 https://doi.org/10.1021/acs.jpcc.6b10279 (2017).
- 30. Zheng, K., Zhao, D., Zhang, D., Liu, N. & Qin, W. Temperature-dependent six-photon upconversion fluorescence of Er 3+. J. Fluor. Chem. 132, 5–8 (2011).
- Shao, W. et al. A core-multiple shell nanostructure enabling concurrent upconversion and quantum cutting for photon management. Nanoscale 11, 11081–11095 (2017).
- Li, F.-C. & Kitamoto, Y. Fabrication of UCNPs/TiO2 aerogel photocatalyst to improve photocatalytic performance. 20013, 20013 (2017).
- Park, Y. I et al. Facile Coating Strategy to Functionalize Inorganic Nanoparticles for Biosensing. Bioconjug. Chem. https://doi. org/10.1021/acs.bioconjchem.6b00524 (2016).
- 34. Carney, R. P. et al. Determination of nanoparticle size distribution together with density or molecular weight by 2D analytical ultracentrifugation. *Nat. Commun.* **2**, 335 (2011).
- 35. Arppe, R. *et al.* Quenching of the upconversion luminescence of NaYF<sub>4</sub>:Yb<sup>3+</sup>, Er<sup>3+</sup> and NaYF<sub>4</sub>:Yb<sup>3+</sup>, Tm<sup>3+</sup> nanophosphors by water: the role of the sensitizer Yb<sup>3+</sup> in non-radiative relaxation. *Nanoscale* **7**, 11746–11757 (2015).
- Cong, T. et al. Upconversion luminescence enhancement in NaYF4: Yb3+, Er3+ nanoparticles induced by Cd2+tridoping. Mater. Res. Bull. https://doi.org/10.1016/j.materresbull.2017.02.032 (2017).
- 37. Haase, M. & Schäfer, H. Upconverting nanoparticles. Angew. Chemie Int. 50, 5808-5829 (2011).
- 38. Menyuk, N., Dwight, K. & Pierce, J. W. NaYF4: Yb, Er An efficient upconversion phosphor. Appl. Phys. Lett. 21, 159-161 (1972).
  - Heer, S., Kömpe, K., Güdel, H. U. & Haase, M. Highly efficient multicolour upconversion emission in transparent colloids of lanthanide-doped NaYF4 nanocrystals. Adv. Mater. 16, 2102–2105 (2004).
  - 40. Wang, F. *et al.* Simultaneous phase and size control of upconversion nanocrystals through lanthanide doping. *Nature* **463**, 1061–1065 (2010).
  - Haro-González, P. et al. Optical trapping of NaYF4:Er3+, Yb3+ upconverting fluorescent nanoparticles. Nanoscale 5, 12192-9 (2013).
  - 42. MacKenzie, L. E. Graphic User Interfaces for the calculation of nanoparticle molecular weight., https://doi.org/10.5518/173 (2017).
  - Mittal, V., Völkel, A. & Cölfen, H. Analytical ultracentrifugation of model nanoparticles: Comparison of different analysis methods. Macromol. Biosci. 10, 754–762 (2010).
  - 44. Wohlleben, W. Validity range of centrifuges for the regulation of nanomaterials: From classification to as-tested coronas. J. Nanoparticle Res. 14 (2012).
  - Liu, Q. et al. Sub-10 nm hexagonal lanthanide-doped NaLuF4upconversion nanocrystals for sensitive bioimaging in vivo. J. Am. Chem. Soc. 133, 17122–17125 (2011).
  - 46. Bragg, W. H. & Bragg, W. L. The Reflections of X-rays by Crystals. Proc. R. Soc. A1 88, 428-438 (913).

- 47. Goetz, J. et al. Ultrabright Lanthanide Nanoparticles. Chempluschem 81, 497 (2016).
- Domingos, R. F. et al. Characterizing manufactured nanoparticles in the environment: Multimethod determination of particle sizes. Environ. Sci. Technol. 43, 7277–7284 (2009).
  - Na, H., Woo, K., Lim, K. & Jang, H. S. Rational morphology control of β-NaYF4:Yb,Er/Tm upconversion nanophosphors using a ligand, an additive, and lanthanide doping. *Nanoscale* 5, 4242–51 (2013).
  - 50. Jia, H., Xu, W., An, J., Li, D. & Zhao, B. A simple method to synthesize triangular silver nanoparticles by light irradiation. 64, 956–960 (2006).
  - Shan, J., Uddi, M., Wei, R., Yao, N. & Ju, Y. The Hidden Effects of Particle Shape and Criteria for Evaluating the Upconversion Luminescence of the Lanthanides Doped Nanophosphors. J. Phys. Chem. C 114, 2452–2461 (2010).
  - 52. Lü, Q., Guo, F., Sun, L., Li, A. & Zhao, L. Silica-/titania-coated Y2O3: Tm3+, Yb 3+ nanoparticles with improvement in upconversion luminescence induced by different thickness shells. J. Appl. Phys. 103 (2008).
  - Li, H., Hao, S., Yang, C. & Chen, G. Synthesis of Multicolor Core/Shell NaLuF4:Yb3+/Ln3+ @CaF2 Upconversion Nanocrystals. Nanomaterials 7, 34 (2017).
  - Lewis, D. J., Day, T. M., MacPherson, J. V. & Pikramenou, Z. Luminescent nanobeads: attachment of surface reactive Eu(III) complexes to gold nanoparticles. *Chem. Commun.* 1433–1435 https://doi.org/10.1039/B518091K (2006).
  - 55. Nel, A. E. et al. Understanding biophysicochemical interactions at the nano-bio interface. Nat. Mater. 8, 543-557 (2009).
  - Bexiga, M. G. *et al.* Cationic nanoparticles induce caspase 3-, 7- and 9-mediated cytotoxicity in a human astrocytoma cell line. Nanotoxicology 5, 557–567 (2011).
  - Hou, Z. et al. UV-Emitting Upconversion-Based TiO 2 Photosensitizing Nanoplatform: Near-Infrared Light Mediated in Vivo Photodynamic Therapy via Mitochondria-Involved Apoptosis Pathway. ACS Nano 9, 2584–2599 (2015).
  - Krämer, K. W. et al. Hexagonal Sodium Yttrium Fluoride Based Green and Blue Emitting Upconversion Phosphors. Chem. Mater. 16, 1244–1251 (2004).
  - 59. Momma, K. & Izumi, F. VESTA 3 for three-dimensional visualization of crystal, volumetric and morphology data. J. Appl. Crystallogr. 44, 1272-1276 (2011).

### Acknowledgements

The authors would like to extend a special thanks to Amy Barker (School of Molecular and Cellular Biology, University of Leeds) for her technical assistance and expertise in conducting and analysing AUC experiments. We are also grateful to Professor Peter Stockley (School of Molecular and Cellular Biology, University of Leeds) for granting access to the AUC facilities. L.E. MacKenzie was supported by a grant from the Biotechnology and Biological Sciences Research Council Tools and Development Resources Fund (BBSRC TDRF) (BB/N021398/1). J.A. Good is supported by a grant from the Medical Research Council (MRC) (MR/N029976/1). A. Vakurov was supported by a grant from the Natural Environment Research Council (NERC). (NE/N007581/1). Padmaja P. Nampi is supported by a European Commission Marie Skłodowska-Curie Individual Fellowship for Experienced Researchers (H2020-MSCA-IF-2015).

### **Author Contributions**

L.E.M. and J.A.G. conceived the research concept and wrote the manuscript. L.E.M. performed all calculations, provided Figs 2, 3, 4, 5, 6, and created the stand-alone GUIs. J.A.G. provided Fig. 1. A.V., P.P.N., S.S., G.J. and P.M, contributed to and reviewed the manuscript.

### **Additional Information**

Supplementary information accompanies this paper at https://doi.org/10.1038/s41598-018-19415-w.

Competing Interests: The authors declare that they have no competing interests.

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2018