This is an author produced version of Light Signaling-dependent Regulation of Photoinhibition and Photoprotection in Tomato.

White Rose Research Online URL for this paper:
http://eprints.whiterose.ac.uk/125018/

Article:
Wang, F, Wu, N, Zhang, L et al. (9 more authors) (2017) Light Signaling-dependent Regulation of Photoinhibition and Photoprotection in Tomato. Plant Physiology. ISSN 1532-2548

https://doi.org/10.1104/pp.17.01143

© 2017 American Society of Plant Biologists. This is an author produced version of a paper published in Plant Physiology Preview. Uploaded in accordance with the publisher's self-archiving policy.
Running Title: HY5 in photoinhibition and photoprotection

Corresponding author:
Yanhong Zhou
Department of Horticulture, Zijingang Campus, Zhejiang University, 866 Yuhangtang Road, Hangzhou 310058, China.

Telephone: 0086-571-88982276
Fax: 0086-571-86971498
E-mail address: yanhongzhou@zju.edu.cn

Research area: Plant science and Ecology
Title: Light Signaling-dependent Regulation of Photoinhibition and Photoprotection in Tomato

Authors: Feng Wang¹, Nan Wu¹, Luyue Zhang¹, Golam Jalal Ahammed¹, Xiaoxiao Chen¹, Xun Xiang¹, Jie Zhou¹, Xiaojian Xia¹, Kai Shi¹, Jingquan Yu¹,², Christine H. Foyer³, and Yanhong Zhou¹,²,∗

¹Department of Horticulture, Zijingang Campus, Zhejiang University, 866 Yuhangtang Road, Hangzhou, 310058, P.R. China
²Zhejiang Provincial Key Laboratory of Horticultural Plant Integrative Biology, 866 Yuhangtang Road, Hangzhou, 310058, P.R. China
³Centre for Plant Sciences, Faculty of Biology, University of Leeds, Leeds, LS2 9JT, UK

One sentence summary: Far-red light alleviates cold-induced photoinhibition and enhances photoprotection in shade leaves via activation of phyA-dependent HY5-ABI5-RBOH1 signaling pathways.

Footnotes:
List of author contributions
Author contributions
Y.Z. conceived the study and analyzed the data; F.W., N.W., L.Z., and X.C. performed the experiments; G.A., X.X., J.Z., Xi.X., and K.S. discussed the data; Y.Z., J.Y. and C. H. F wrote the article with contributions from the other authors.

Funding information: This work was supported by the National Natural Science Foundation of China (grant nos. 31672198, 31430076), the Fundamental Research Funds for the Central Universities (2016XZZX001-07) and the Fok Ying-Tong Education Foundation (132024).

*Corresponding author; email: yanhongzhou@zju.edu.cn
ACKNOWLEDGMENTS

We are grateful to Prof. Jim Giovannoni of Cornell University and the Tomato Genetics Resource Center at the California University for tomato seeds. We thank Prof. Gang Lu (Zhejiang University) for denoting the CRISP/Cas9 vectors.
Photoreceptor-mediated light signaling plays a critical role in plant growth, development, and stress responses but its contribution to the spatial regulation of photoinhibition and photoprotection within the canopy remains unclear. Here, we show that low red/far-red (L-R/FR) ratio light conditions significantly alleviate PSII and PSI photoinhibition in the shade leaves of tomato plants. This protection is accompanied by a phytochrome A (phyA)-dependent induction of LONG HYPOCOTYL 5 (HY5). HY5 binds to the promoter of ABA INSENSITIVE 5 (ABI5), triggering RESPIRATORY BURST OXIDASE HOMOLOG1 (RBOH1)-dependent H$_2$O$_2$ production in the apoplast. Decreased levels of HY5, ABI5 and RBOH1 transcripts increased cold-induced photoinhibition and abolished L-R/FR-induced alleviation of photoinhibition. L-R/FR illumination induced non-photochemical quenching (NPQ) of chlorophyll a fluorescence and increased the activities of Foyer-Halliwell-Asada cycle enzymes and cyclic electron flux (CEF) around PSI. In contrast, decreased HY5, ABI5 and RBOH1 transcript levels abolished the positive effect of L-R/FR on photoprotection. Loss of PROTON GRADIENT REGULATION5 (PGR5)-dependent CEF led to increased photoinhibition and attenuated L-R/FR-dependent NPQ. These data demonstrate that HY5 is an important hub in the cross-talk between light and cold response pathways, integrating ABA and reactive oxygen species signaling leading to the attenuation of photoinhibition by enhanced induction of photoprotection in shade leaves.
INTRODUCTION

Low temperatures are a major factor limiting the productivity and geographical distribution of plant species. Tropical and subtropical plants are generally sensitive to chilling because of lack of the capacity for cold acclimation (Zhu et al., 2007). Many economically important species such as maize, rice and tomato are unable to survive long term exposures of temperatures below 12 °C. In addition to interspecific differences in chilling sensitivity, the tolerance of a given species to low growth temperatures varies between organs and according to developmental stage. Within the canopy, the upper “sun”, leaves often exhibit a higher sensitivity to chilling than the shade leaves. However, little is known about the mechanisms that contribute to the spatial differences in chilling tolerance.

In many situations, leaves absorb more light than can be effectively utilized in photosynthesis, especially when plants are exposed to stress. The excess light energy has to be dissipated because over-excitation has the potential to damage the photosynthetic machinery, particularly PSII in the process called photoinhibition (Kingston-Smith et al., 1997; Kingston-Smith and Foyer, 2000; Foyer et al., 2017). This process is characterized by the decreases in the maximal photochemical efficiency of PSII (Fv/Fm) and in maximal P700 oxidation (ΔP700_max) in PSI. Meanwhile, plants have evolved a range of photoprotective mechanisms to decrease the probability of damage to the PSII and PSI reaction centers. Photoprotection involves diverse processes such as chloroplast avoidance movement, dissipation of absorbed light energy as thermal energy (NPQ), pseudocyclic electron flow coupled to reactive oxygen species (ROS) scavenging systems (Foyer-Halliwell-Asada cycle), cyclic electron flow (CEF) around PSI and the photorespiratory pathway (Takahashi and Badger, 2011). The dominant component of NPQ is the energy-dependent nonphotochemical quenching (qE), which is induced by an increase in the proton gradient across the thylakoid membrane (ΔpH) under excess light conditions (Munekage et al., 2004). PSII subunit S (PsbS) protein acts as a sensor of lumen pH and may activate qE through conformational changes of LHCII (Li et al., 2002; Ahn et al., 2008). The mechanisms that contribute to NPQ are not completely understood but it is widely accepted that two distinct xanthophyll-dependent quenching mechanisms involving xanthophyll cycle
pigments and lutein 1, respectively, participate in the ΔpH-triggered, PsbS-mediated conformational changes of LHCII (Ruban et al., 2007; Ahn et al., 2008). In the xanthophyll cycle, violaxanthin (V) is converted into zeaxanthin (Z) under high light, via the intermediate antheraxanthin (A), a reaction that is catalyzed by the enzyme violaxanthin deepoxidase (VDE). The presence of Z activates thermal dissipation of the excess energy (Niyogi et al., 1997, 1998). The de-epoxidation state of the xanthophyll cycle pigments is thought to regulate qE-dependent NPQ (Kromdijk et al., 2016). Alterations in VDE activity influence the extent of PSII photoinhibition (Niyogi et al., 1998; Han et al., 2010).

CEF around PSI is considered to involve NAD(P)H dehydrogenase (NDH) complex-dependent and PROTON GRADIENT REGULATION5 (PGR5)/PGRL1 complex-dependent pathways (Shikanai, 2007), the latter being responsible for most of the required additional ΔpH generation across the thylakoid membrane (Munekage et al., 2004). The generation of an increased trans-thylakoid ΔpH gradient by CEF is important for the activation of qE (Munekage et al., 2004). PGR5-PGRL1 dependent CEF pathway is regulated by the chloroplastic redox state and is activated under stress conditions (Okegawa et al., 2008; Strand et al., 2015). Decreases in both CEF and qE resulted in an inhibition of the synthesis of the D1 protein (Takahashi et al., 2009). Moreover, loss of function of proteins involved in CEF around PSI increased the sensitivity of plants to photoinhibition of PSII and also PSI (Munekage et al., 2002). Furthermore, suppression of Foyer-Halliwell-Asada cycle enzymes increased photoinhibition whilst an overexpression of the genes encoding these enzymes tended to decrease photoinhibition (Foyer et al., 1995; Maruta et al., 2010).

The effects of high light intensities or fluctuations in light intensity on the extent of photoinhibition of PSII and PSI have been intensively studied over the past 40 years (Kim and Tokura, 1995). In contrast, relatively little is known about the effects of light quality on photoinhibition or photoprotection. Plants have developed a set of sophisticated photoreceptors, including phytochromes (PHYs), cryptochromes (CRYs), phototropins (PHOTs) and UV-B light photoreceptors (e.g. UVR8) to perceive changes in light quality (Möglich et al., 2010). Of these, blue-light photoreceptors (e.g. PHOT) have been reported to activate chloroplast avoidance movements in sessile plants under excess light conditions (Kasahara et al., 2002). Energy dissipation in green algae is also controlled by the PHOT...
and UVR8 photoreceptors, which are activated by blue and UV light (Petroutsos et al., 2016; Allorent et al., 2016; Allorent and Petroutsos, 2017). PhyA and phyB, which are the photoreceptors for far-red light (FR) and red light (R) respectively, play a central role in regulating the expression of a large number of light-responsive genes that are involved in regulation of a wide range of processes from photomorphogenesis to stress responses (Quail, 2002a, 2002b; Franklin and Quail, 2010; Wang et al., 2016), however, the role of phytochromes in the regulation of photoinhibition has not been well characterized. LONG HYPOCOTYL 5 (HY5), a basic leucine zipper (bZIP) transcription factor, acts downstream of multiple photoreceptors, in the signal transduction pathway that links various signaling pathways including light and phytohormone signaling (Cluis et al., 2004; Jiao et al., 2007; Lau and Deng, 2010). HY5 is also important in the regulation of cold acclimation responses, promoting the expression of a large number of cold-inducible genes (Catala et al., 2011). Interestingly, low temperatures lead to the stabilization of HY5 through exclusion of CONSTITUTIVE PHOTOMORPHOGENIC1 (COP1) from the nucleus. COP1 is an E3 ubiquitin ligase targeting HY5 for proteasome-mediated degradation in response to light (Catala et al., 2011). Whether HY5 is involved in the regulation of photoprotection and photoinhibition is as yet unknown. However, HY5 is known to regulate the expression of several ROS and anthocyanin-related genes (Catala et al., 2011).

Spatial variations in the chilling sensitivity of leaves at different positions in a canopy have been reported under field conditions, the upper sun-exposed leaves being more easily injured by a cold episode than the shade leaves. Shading not only decreases the light intensity arriving the leaf surface, but also reduces the R/FR ratios of the light available to the shade leaves (Sasidharan et al., 2009). However, the role of changes in R/FR ratio on cold tolerance within a canopy remains largely unexplored, particularly with regard to effects on photoinhibition and photoprotection. Here we show that spatial differences in cold tolerance and in photoinhibition are linked to light-quality-regulated photoprotection. Data are presented showing that low R/FR ratios induce an accumulation of HY5 transcripts in a phyA-dependent manner. The increased expression of HY5 leads to improved cold tolerance by enhancing ABA signaling through direct binding of HY5 to the ABI5 promoter and induction of RBOH1-dependent apoplastic H$_2$O$_2$ generation. ABI5 participates in the
regulation of photoprotection by inducing a strong antioxidant response, as well as enhancing PGR5-dependent NPQ. Taken together, these data show that phytochrome-mediated HY5-ABA-ROS signaling plays a key role in avoiding cold-induced photoinhibition by inducing photoprotection within the canopy in response to variations in light quality.
RESULTS

Spatial Variation in Photoinhibition is Partially Attributable to the Changes in Light Quality Conditions

Tomato plants were grown to the 11th leaf-stage and then exposed to a cold treatment at 4 °C under white light for 7 d. The degree of photoinhibition of PSII and PSI was then compared in leaves at the 9th and 5th ranks from the base. Leaves at the 9th rank had lower Fv/Fm ratios and lower maximum P700 photooxidation level (ΔP700max), together with higher levels of relative electrolyte leakage (REL) than the 5th leaves (Fig. 1, A and B; Supplemental Fig. S1). Light quality analysis revealed that the R/FR ratio was decreased from 1.3 at the 9th leaf rank to 0.5 at 5th leaf rank.

We then examined the effects of light quality on cold-induced photoinhibition by exposing tomato plants at the 6-leaf stage to a cold treatment at 4 °C under high R/FR ratio, i.e. low FR intensity (L-FR, 133 µmol m⁻² s⁻¹) or low R/FR ratio, i.e. high FR intensity (H-FR, 400 µmol m⁻² s⁻¹) light conditions, respectively, using monochromatic LEDs. In these experiments, the R light intensity was maintained at 200 µmol m⁻² s⁻¹ under both light quality treatment regimes. Chilling-induced decreases in Fv/Fm ratios and in ΔP700max were lower under H-FR than the values determined in plants exposed to the L-FR conditions (Fig. 1, C and D; Supplemental Fig. S2, A and B). NPQ values, PsbS protein accumulation and de-epoxidation state of the xanthophyll cycle, i.e. (A+Z)/(V+A+Z) ratio, were increased after cold stress, especially under H-FR light conditions (Supplemental Fig. S2, C-E). Western Blot analysis revealed that chilling stress induced a decrease in the accumulation of the PsaB and PsaC proteins, especially under L-FR conditions (Fig. 1E). We also tested the acceptor-side limitation in chilled leaves by application of 25 µM methyl viologen (MV). Results showed that chilling-induced decrease of ΔP700max was mostly relieved in the MV-treated leaves, especially under H-FR conditions (Fig. 1F). These results suggest that the main cause of the chilling-induced decrease in ΔP700max is the degradation of PSI submits resulting in an acceptor side limitation in PSI.

RNA-seq analysis was performed on the 4th leaves after exposure of plants to a cold treatment at 4 °C under either L- or H-FR light conditions. This generated a total of 103,463,126 reads, which were aligned to the Solanum lycopersicum reference genome.
Compared with the plants grown under L-FR conditions, a total of 6312 transcripts (3607 increased in abundance and 2705 decreased in abundance) were differentially changed under the H-FR (Supplemental Table S1). An examination of the levels of transcripts encoding photosynthetic proteins, and proteins involved in light and ABA signaling revealed a subset of mRNAs that showed differential changes in response to the light quality, having higher levels in leaves exposed to H-FR light compared to L-FR. These transcripts encoded proteins are involved in PSII (PHOTOSYstem II LIGHT HARVESTING COMPLEX GENE 2.1 and PHOTOSYstem II REACTION CENTER PROTEIN A), PSI (PHOTOSYstem I SUBUNIT I and PHOTOSYstem I, PsA/PsaB PROTEIN), cyclic electron flux (PGR5 and PGR5-LIKE A), plastoquinone cytochrome b$_6$f complexes (CYTOCHROME b561/FERRIC REDUCTASE TRANSMEMBRANE PROTEIN FAMILY and CYTOCHROME B5 ISOFORM B), FAD/NAD(P)-binding oxidoreductase family protein (NAD(P)-BINDING ROSSMANN-FOLD SUPERFAMILY PROTEIN and FAD/NAD(P)-BINDDING OXIDOREDUCTASE FAMILY PROTEIN), oxidoreductase proteins (PEROXIDASE 2 and ASCORBATE PEROXIDASE 3), thermal energy dissipation (CAROTENOID CLEAVAGE DIOXYGENASE 1 and ZEAXANTHIN EPOXIDASE, ZEP; Supplemental Fig. S2F; Supplemental Table S2).

**PhyA Acts as a Positive Regulator for Light Quality-Dependent Regulation of Photoinhibition**

Cold-induced photoinhibition was compared in wild type (WT) tomato leaves and in mutants deficient in phytochrome A (phyA and phyAB1B2) or phytochrome B (phyB1B2) grown under either L-FR or H-FR conditions. The phyA mutants had lower Fv/Fm ratios and $\Delta$P700$_{\text{max}}$ levels than the WT following exposure to a cold treatment at 4 °C for 7 d (Fig. 2A). In contrast, the phyB1B2 plants showed higher Fv/Fm ratios and $\Delta$P700$_{\text{max}}$ values than the WT under these conditions. Moreover, the phyA and phyAB1B2 mutants had lower NPQ values, with less PsbS protein accumulation and lower CEF rates compared to WT plants (Fig. 2, B-D). In contrast, the phyB1B2 plants had higher NPQ values, PsbS protein accumulation and CEF rates than the WT. In addition, H-FR significantly induced increases in Fv/Fm ratios, $\Delta$P700$_{\text{max}}$ values, NPQ values, PsbS protein accumulation and CEF rates in
WT and phyB1B2 mutant, but not in phyA and phyAB1B2 mutants after the cold treatment (Fig. 2, A-D).

The levels of HY5 transcripts were increased in response to a cold treatment in WT tomato leaves grown under H-FR compared to L-FR conditions (Fig. 2E). The H-FR growth regime also resulted in significant chilling-induced increases in the levels of HY5 transcripts in the phyB1B2 mutant leaves but not in those of the phyA or phyAB1B2 mutants. However, an inverse pattern of response to change in FR intensity was observed for COP1 transcripts. Growth under the H-FR light regime decreased the levels of COP1 mRNAs in the WT and in the phyB1B2 mutants under cold stress. In contrast, differences in the FR intensity had little effect on the levels of COP1 transcripts in phyA and phyAB1B2 mutants after the cold treatment.

**FR-Induced HY5 Alleviated Photoinhibition by Induction of Photoprotection**

The levels of HY5 and COP1 transcripts were decreased by 80% and 70%, respectively, in HY5-RNAi and COP1-RNAi plants used for the study (Supplemental Fig. S3A). Cold and FR intensity-induced changes in Fv/Fm ratios, ΔP700_max, survival rates, REL and the levels of oxidized protein, as determined by the presence of protein carbonyl groups, were measured in the HY5-RNAi and COP1-RNAi plants (Fig. 3, A and B; Supplemental Fig. S3, B and C). Cold-induced increases in REL and in the levels of oxidized proteins were higher in the HY5-RNAi plants compared to the WT and COP1-RNAi plants regardless of FR intensity (Supplemental Fig. S3C). In contrast, the Fv/Fm ratios and the ΔP700_max values were much lower in the leaves of the HY5-RNAi plants compared to the WT and COP1-RNAi plants under both FR light conditions (Fig. 3, A and B). The chilling-induced decreases in the Fv/Fm ratios and the ΔP700_max values were significantly less in the COP1-RNAi plants than the WT (Fig. 3, A and B). Significantly, H-FR treatment induced increases in Fv/Fm ratios, ΔP700_max values, survival rates and decreases in REL or the levels of oxidized proteins in the WT and the COP1-RNAi plants, but had little effects on these parameters in the HY5-RNAi plants (Fig. 3, A and B; Supplemental Fig. S3, B and C). Moreover, the HY5-overexpressing plants showed an increased tolerance to the cold treatment compared to the WT (Supplemental Fig. S4). Taken together, these results
indicate that HY5 is required for the light quality-mediated regulation of chilling tolerance in tomato and that COP1 negatively regulates this process.

NPQ, cyclic electron flux (CEF) and Foyer-Halliwell-Asada cycle all play important roles in preventing the photosystems from photodamage or photoinhibition (Foyer et al., 1995; Takahashi et al., 2009; Chen and Gallie, 2012). In comparison to the WT plants, the HY5-RNAi plants showed decreased levels of NPQ, PsbS protein accumulation, antioxidant enzyme activities and CEF rates (Fig. 3, C-F; Supplemental Fig. S5). These parameters were increased in the COP1-RNAi plants relative to the WT. An increase in the FR intensity significantly increased the level of NPQ and the accumulation of PsbS protein, and the abundance of transcripts encoding Foyer-Halliwell-Asada cycle enzymes (Cu/Zn-SUPEROXIDE DISMUTASE, Cu/Zn-SOD; ASCORBATE PEROXIDASE, tAPX; MONODEHYDROASCORBATE REDUCTASE, MDAR; DEHYDROASCORBATE REDUCTASE, DHAR and GLUTATHIONE REDUCTASE 1, GR1), as well as the activities of these enzymes, and the rate of CEF in WT and COP1-RNAi plants (Fig. 3, C-F; Supplemental Fig. S5). The effects of the FR intensity were more pronounced in the COP1-RNAi plants. However, the H-FR treatment had little effect on the level of NPQ, the activities of antioxidant enzymes or the rates of CEF in the HY5-RNAi plants, suggesting that HY5 is essential for the H-FR regulation of photoprotection.

**HY5 is a Transcriptional Activator of ABI5**

While exposure to cold stress had no effect on stomatal movements in HY5-RNAi plants, this treatment caused a decrease in stomatal aperture in COP1-RNAi leaves, especially under H-FR light conditions (Supplemental Fig. S6, A and B). Given that ABA signaling positively regulates stomatal movement, we examined whether HY5 could bind to the promoters of any of the ABA signaling genes. For this analysis, we inspected 2.5 kb sequences upstream of the transcriptional start sites of a set of tomato ABA INSENSITIVE (ABI) genes. Of these, the promoters of three ABA signaling genes (ABI3-1, ABI3-2 and ABI5) contain the G-box sequences: CACGTG (Fig. 4A; Supplemental Fig. S6C). Electrophoretic mobility shift assays (EMSA) were used to analyze whether HY5 binds directly to these promoters in vitro. The probe-protein complex was not detected using...
ABI3-1 and ABI3-2 probes. However, HY5 directly bound to the promoter probe of ABI5 (Fig. 4C; Supplemental Fig. S6D). When the core sequence of G-box element motif in ABI5 probe was mutated in a single base (ABI5-G-mut2) or multiple bases (ABI5-G-mut1, Fig. 4B), the binding to the complexes was decreased, or even totally lost (Fig. 4C). Based on these observations, we conclude that HY5 protein binds specifically to the G-box element sequences of the synthesized probes for the ABI5 promoters in vitro.

To further determine whether the tomato HY5 protein binds directly to the promoter of ABI5 in vivo under cold stress, we performed ChIP-qPCR assays. As shown in Fig. 4D, the ABI5 promoter sequence was substantially enriched in fractions using the anti-HA antibody that immune-precipitates the 3HA-tagged HY5 transgene product in the HY5 overexpressing (OE) lines but not the WT after 6 h of cold stress under H-FR. However, the IgG control antibody failed to pull down ABI5 gene promoter DNA segment (Fig. 4D). We then assessed the levels of ABI5 transcripts in WT, HY5-RNAi and COP1-RNAi plants exposed to H- and L-FR conditions under cold stress conditions (Fig. 4E). No changes in ABI5 transcript levels were detected in the HY5-RNAi plants in relation to the FR intensity. In contrast, an increase in the FR gradually induced increase the abundance of ABI5 transcripts in WT and COP1-RNAi plants, the induction being more significant in the COP1-RNAi plants than the WT. The induction of ABI5 expression was greater in the HY5 overexpressing lines (OE#1 and OE#3, expressing high HY5 protein levels, Supplemental Fig. S4A) than the WT after 6 h of cold stress under the H-FR irradiance regime (Fig. 4F). These results indicate that HY5 binds directly the promoter of ABI5 and activates its expression, subsequently regulates cold tolerance of tomato in response to light quality.

When WT and phytochrome mutant plants were exposed to L-FR and H-FR light conditions at 4 °C, higher levels of ABI5 transcripts were maintained in phyB1B2 mutants compared to the WT, phyA or phyAB1B2 plants under both light quality conditions (Fig. 4G). Moreover, the higher FR intensity increased the levels of ABI5 transcripts in WT and phyB1B2 plants, but not in phyA or phyAB1B2 mutants.

**Role of ABI5 in Light Quality-Regulated Photoinhibition and Photoprotection**
Lines of ABI5-silenced (pTRV-ABI5) plants were generated, using a virus-induced gene silencing (VIGS). These lines showed a reduction in ABI5 transcript levels of 75% (Supplemental Fig. S7, A and B). Tomato pTRV-ABI5 plants showed an increased sensitivity to cold-induced photoinhibition compared to the pTRV plants, as measured by a decrease in the Fv/Fm ratio and in ΔP700_max, as well as an increase in REL (Fig. 5, A and B; Supplemental Fig. S7, C and D). Interestingly, the H-FR-induced cold tolerance and alleviation of photoinhibition observed in the pTRV plants was completely lost in the ABI5-silenced plants, which showed no significant differences in Fv/Fm ratios, ΔP700_max under cold stress at both light quality regimes. These observations clearly indicate that loss of ABI5 function compromised the H-FR-induced alleviation of chilling dependent photoinhibition in tomato. In support of this hypothesis, we observed that the H-FR-induced changes in NPQ, PsbS protein accumulation, and the activities of antioxidant enzymes, as well as the rate of CEF were abolished or attenuated in the pTRV-ABI5 plants (Fig. 5, C-F). These results show that ABI5 functions as a down-stream of HY5 in light-regulated photoprotection.

RBOH1-Dependent ROS Production Prevents Photoinhibition by Activation of Photoprotection

ABA signaling is linked to the upregulation of RBOH-dependent ROS production in response to stress (Murata et al., 2001; Xing et al., 2008; Zhou et al., 2014). The cold treatment used in the present study increased the levels of RBOH1 transcripts and apoplastic H2O2 accumulation (Fig. 6, A and B). Moreover, silencing ABI5 (pTRV-ABI5) abolished the H-FR-dependent induction of RBOH1 expression and apoplastic H2O2 accumulation.

RBOH1-RNAi plants were used to examine whether RBOH1 plays a role in the regulation of cold-induced photoinhibition and photoprotection under L- and H-FR conditions. The RBOH1-RNAi plants showed lower apoplastic H2O2 accumulation and an increased sensitivity to photoinhibition compared to the WT. The response to changes in FR intensities was also compromised in terms of survival rates and changes in the Fv/Fm ratios, ΔP700_max and REL (Fig. 6, C and D; Supplemental Fig. S8). In addition, ABA-induced alleviation of photoinhibition was compromised by treatment with dimethylthiourea.
(DMTU, a ROS scavenger). This effect was also not observed in the RBOH1-RNAi plants (data not shown). Moreover, H-FR-induced changes in NPQ, PsbS protein accumulation and the activities of antioxidant enzymes, as well as the rate of CEF were abolished in the RBOH1-RNAi plants (Fig. 6, E-H). These results suggest that the ABI5-dependent production of H$_2$O$_2$ plays a pivotal role in HY5-regulated photoprotection by functioning as a critical downstream component in light signaling.

**Light-Activated CEF Plays Dual Roles in Preventing Plants from Photoinhibition**

The roles of NPQ and antioxidants in photoprotection are well established (Foyer et al., 1995; Niyogi et al., 1997, 1998; Chen and Gallie, 2012). However, relatively, little is known about the role of PGR5-PGRL1-dependent and NDH-dependent CEF in photoprotection (Shikanai, 2007). Here, we show that chilling stress increased the accumulation of PGR5 transcripts by more than 5-fold but had less effect on the levels of PGRL1A and ORANGE RIPENING (ORR) transcripts (Supplemental Fig. S9). ORR encodes an NDH-M subunit in the tomato Ndh complex (Nashilevitz et al., 2010). Chilling-induced increases in PGR5, PGRL1A and ORR transcripts were greater following exposure of plants to H-FR light conditions. In comparison, PGRL1B transcripts were decreased by the chilling treatment and they were not affected by FR levels. Moreover, PHYA deficiency or silencing of HY5, ABI5 and RBOH1 abolished the H-FR-dependent induction of PGR5, PGRL1A and ORR transcripts. These results suggested the potential involvement of the PGR5-PGRL1 dependent and NDH dependent CEF in the photoprotection in response to the cold stress.

We then generated pgr5 mutants by using a Crisp/cas9 technique and also PGR5-overexpressing (PGR5-OE) tomato plants (Supplemental Fig. S10). The pgr5 plants showed decreased CEF rates while the PGR5-OE plants had increased CEF rates under cold stress (Supplemental Fig. S11C). Moreover, the cold-mediated induction of CEF under H-FR light conditions was lower in the pgr5 plants than the WT. In contrast, this parameter was higher in the PGR5-OE plants exposed to cold stress under H-FR light conditions. Significantly, the cold treatment led to greater decreases in the Fv/Fm ratios and in $\Delta P_{700_{\text{max}}}$, and in increase in REL in the pgr5 plants than the WT (Fig. 7, A and B; Supplemental Fig. S11, A and B), while PGR5-overexpressing significantly increased Fv/Fm ratios and $\Delta P_{700_{\text{max}}}$
values after a cold stress. H-FR induced increases in qE, NPQ, PsbS protein acclimation and (A+Z)/(V+A+Z) ratio in the WT and PGR5-OE plants but not in the pgr5 plants (Fig. 7, C-F). These results suggest that PGR5-dependent CEF is essential for light-regulated photoprotection. To provide further evidence for the roles of HY5, ABI5 and RBOH1 in PGR5-dependent photoprotection, we silenced HY5, ABI5 and RBOH1 in PGR5-OE plants (Supplemental Fig. S12). As observed in WT plants, silencing of these genes in PGR5-OE plants significantly decreased ΔP700 max and compromised H-FR-induced increase in ΔP700 max. These findings show that HY5-ABI5-RBOH1 cascades play a critical role in FR-induced and PGR5-dependent photoprotection in the plants.

DISCUSSION

The management of light energy usage in photosynthesis is a key concept of photosynthetic regulation (Foyer et al., 2017). A wide range of mechanisms have evolved to protect the photosystems from the potentially damaging effects of the high irradiances that occur in the natural environments. While photoinhibition may not be such a common phenomenon in nature as was once thought because recovery without damage is facilitated by the protective component of NPQ (Foyer et al., 2017), understanding the regulation of NPQ at different leaf ranks within the plant canopy is crucial to plant productivity. The recent demonstration that acceleration of the NPQ relaxation can lead to significant increases in crop yield (Kromdijk et al., 2016), highlights the importance of understanding how photosynthetic efficiency is regulated. The data presented here provides new information concerning the regulation of leaves to shading. We show that spatial variations in susceptibility to cold-induced photoinhibition are attributable to differences in the R/FR ratios experienced by the leaves. In particular, phyA-mediated induction of HY5 under different R/FR regimes plays a critical role in photoprotection. Through binding to the promoter of ABI5, HY5 triggers enhanced protoprotection through induction of an apoplastic H2O2 burst that influences antioxidant status, CEF and NPQ. This enhanced photoprotection allows shade leaves to avoid photoinhibition.

Chlorophyll-containing cells absorb blue light and R light, whereas FR photons are either transmitted or reflected. This leads to a decrease in the R/FR ratios experienced deep
leaves within the vegetative canopy compared to leaves exposed to full sunlight (Sasidharan et al., 2009). The data presented here show that the upper leaves experiencing a high R/FR ratios (i.e. low FR intensity, L-FR) growth environment have a higher degree of photoinhibition compared to shade leaves that experience a low R/FR ratios (i.e. high FR intensity, H-FR) growth environment (Fig. 1, A and B; Supplemental Fig. S1). To exclude the potential effects of other parameters such as leaf developmental stage and light intensity on photoinhibition and cold tolerance, plants were exposed to R at same light intensity and only FR intensities were changed. Photoinhibition and electrolyte leakage were significantly decreased, while NPQ values, PsbS protein accumulation and (A+Z)/(V+A+Z) ratio were increased under the low-R/FR ratios light growth regime (Fig. 1, C-F; Supplemental Fig. S2, A-E). Therefore, spatial differences in sensitivity to photoinhibition and in cold tolerance are largely attributable to differences in R/FR ratios within the growth environment. Moreover, the low R/FR ratio experience by the leaves plays a positive role in tolerance to excess light such that shade leaves are less sensitive to photoinhibition. By using western blotting against PSI subunits PsaB and PsaC, and suppling of MV, an artificial electron acceptor from PSI, we found FR plays a critical role in photoprotection by alleviating the degradation of PSI submits and the release of acceptor side limitation of PSI (Fig. 1, E and F).

It is widely accepted that the primary functions of phytochromes is to detect environmental fluctuations in the relative proportions of R and FR radiation (Chen et al., 2004). Data presented here show that the FR receptor phyA and R receptor phyB are respectively, positive and negative regulators of photoinhibition (Fig. 2, A-D). These findings are in agreement with earlier studies demonstrating that phyA and phyB1B2 deficient tomato plants had increased and decreased sensitivities to chilling (Wang et al., 2016). It is of interest to note that the higher tolerance to cold observed in shade leaves contrasts markedly with the reported decreased resistance to herbivory and pathogens. Numerous studies have reported that the low R/FR ratios experienced by shade leaves increase plant population densities, and increase herbivory and disease (Xie et al., 2011; Ballaré, 2014). In contrast to the increases in cold tolerance observed here in phyB tomato plants, similar mutants in Arabidopsis were reported to be more sensitive to Pseudomonas
syringae pv. tomato DC3000 (de Wit et al., 2013). Recent studies have demonstrated that
high R/FR ratios have important effects on plant defenses through effects on JA signaling
and other defense pathways (Cerrudo et al., 2012; Nagata et al., 2015). Therefore, plants
appear to have evolved different mechanisms for coping with biotic stress and abiotic
stresses through the integration of light signaling pathways with those involving the
perception of other environmental stimuli.

Multiple photoreceptors promote the accumulation of HY5 in response to changing light
conditions. As a member of the bZIP transcription factor family, HY5 plays a critical role in
different plant processes such as hormone-, nutrient-, abiotic stress- and redox-signaling
pathways (Gangappa and Botto, 2016). This places HY5 at the center of the transcriptional
network hub that regulates plant responses to environmental change. One mechanism by
which this is achieved is through regulation of the nuclear abundance of COP1, an E3
ubiquitin ligase that targets HY5 for proteasome-mediated degradation in darkness
(Osterlund et al., 2000; Yi and Deng, 2005). Exposure to cold stress induced HY5
expression in WT and phyB1B2. However, mutation in phyA abolished cold-induced
transcript of HY5 under both L-FR and H-FR light conditions (Fig. 2E). In addition, changes
in the COP1 transcript levels were in contrast with those in HY5 transcript levels in WT
leaves and in phytochrome-deficient mutants. These results suggest that the induction of
HY5 and COP1 in response to the cold stress is phyA and phyB-dependent respectively. To
date, our knowledge of the role of HY5 in plant cold responses was limited to the regulation
of anthocyanin accumulation in Arabidopsis (Catala et al., 2011). The data presented here
show that HY5 and COP1 are positive and negative regulators for the plant cold response
leading to the regulation of photoinhibition (Fig. 3; Supplemental Figs. S3-S5).

ABA signaling is known to play an important role in responses to various
environmental stresses (Zhao et al., 2013). Analysis using EMSA and ChIP-qPCR assay
revealed that HY5 binds to the G-box element of the ABI5 promoter in vitro and in vivo with
a high specificity (Fig. 4, A-D; Supplemental Fig. S6, C and D). In the absence of HY5, the
ability of H-FR-induced signals to increase ABI5 transcript levels was impaired (Fig. 4E).
The induction of ABI5 expression was also found to be phyA-dependent and significantly
increased in HY5-overexpressing plants compared to WT plants after exposure to cold stress
ABI5 had been shown to be involved in the regulation of seed germination and responses to drought (Chen et al., 2008). ABI5 involvement in drought stress responses has been assessed through adjustments in ROS scavenging and in osmotic potential in cotton (Mittal et al., 2014). ABA signaling, like brassinosteroid signaling, is known to have a role in the induction of apoplastic H$_2$O$_2$ accumulation in plants (Zhou et al., 2014). The data presented here show that silencing ABI5 compromised the H-FR-induced alleviation of photoinhibition (Fig. 5; Supplemental Fig. S7), as well as the induction of RBOH1 expression and H$_2$O$_2$ accumulation in the leaf apoplast (Fig. 6, A and B). Taken together, these findings strongly suggest that ABI5 not only functions as a downstream component of the light-regulated cold tolerance pathway in a HY5-dependent manner, but that it is also linked to ROS signaling. Furthermore, RBOH1-RNAi plants failed to respond to changes in FR intensities in terms of effects on Fv/Fm and $\Delta$P700$_{\text{max}}$ (Fig. 6, C-H; Supplemental Fig. S8). These findings suggest that RBOH1-dependent H$_2$O$_2$ production plays an essential role in the adjustment of the photosynthetic processes to changes in light quality. Taken together, our results demonstrate that light quality signaling participates in the regulation of the responses of photosynthesis to chilling by regulation of HY5-ABI5-RBOH1 signaling pathways.

Plants absorb sunlight to power the photochemical reactions of photosynthesis with the generation of ROS, a process that is increased under stress (Foyer et al., 2012). While ROS are highly reactive and have been proposed to accelerate photoinhibition through direct oxidative damage to PSII/PSI (Nishiyama et al., 2006), they are also vital signals relaying information concerning changes in the redox status of the chloroplast to the nucleus stress (Foyer et al., 2012). Plants have developed diverse photoprotection mechanisms to limit light-induced damage to the photosynthetic apparatus (Takahashi and Badger, 2011). Thermal energy dissipation, cyclic electron flow and the direct transfer of energy and electrons to oxygen in pseudocyclic electron flow fulfill crucial roles in photosynthetic regulation and photoprotection (Foyer et al., 2012). The data presented here show that the increased sensitivity to cold-induced photoinhibition observed in the HY5-RNAi, pTRV-ABI5 and RBOH1-RNAi plants was linked to decreased capacity of photoprotection (Figs. 3, 5 and 6; Supplemental Figs. S3-5, 7 and 8). These findings suggest that the HY5-ABI5-
RBOH1 signaling pathway plays a critical role in the induction of the photoprotection mechanisms that serve to avoid cold-induced photoinhibition.

The data presented here show that exposure to H-FR intensities induce NPQ, PsbS protein accumulation and CEF, as well as increasing the activities of five enzymes involved in antioxidant reactions in plants experiencing cold stress. Moreover, loss of HY5, ABI5 or RBOH1 functions compromised the H-FR-induced NPQ, CEF and the increases in antioxidant enzyme activities at low temperatures (Figs. 3, C-E, 5, C-F and 6, D-H; Supplemental Fig. S5). We conclude that the HY5-ABI5-RBOH1 pathway is required for the FR induction of photoprotection in response to cold stress. It is worth noting that phyA, HY5-RNAi, pTRV-ABI5 and RBOH1-RNAi plants all showed reduced accumulation of the NPQ effector protein PsbS relative to WT and they showed little response to increases in FR light intensities (Figs. 2D, 3E, 5E and 6G). Reduced accumulation of PsbS and the insufficient trans-thylakoid $\Delta p\text{H}$, caused by their severely damaged CEF may contribute to the impaired NPQ in these plants (Figs. 2B-D, 3C-E, 5C-E and 6E-G). In addition, a FR-induced increase in $(A+Z)/(V+A+Z)$ ratios was not observed in these plants (data not shown). It is plausible that the FR-activated and phyA-mediated HY5-ABI5-RBOH1-dependent signaling pathway is linked to a NPQ-specific effect on photoprotection. While transcript of PGR5 was under the regulation by HY5, ABI5 and RBOH1 in response to the change in FR intensity, silencing of HY5, ABI5 and RBOH1 in PGR5-OE plants also compromised H-FR-induced increase in $\Delta P700$max. (Supplemental Fig. S12). In this case, $\Delta P700$max was influenced by both the inherent PGR5, which could be modified by light conditions, and 35S promoter driven PGR5, which is insensitive to the changes in light conditions, respectively. This is why H-FR altered the $\Delta P700$max in WT and PGR5-OE plants to a similar degree. However, we could not exclude the possibility for the involvement of other regulatory mechanisms. HY5 is also required for the suppression of excessive ROS accumulation during acclimation to low temperatures (Catala et al., 2011). Similarly, ABA signaling also plays a role in the expression and/or activities of antioxidant enzymes, a role that is dependent to a large extent on the induction of apoplastic $H_2O_2$ production (Zhang et al., 2007). We have previously reported that apoplastic $H_2O_2$ production plays a critical role in cold acclimation by protection of PSII.
Therefore, FR-induces photoprotection and suppresses excessive ROS accumulation in a HY5-, ABI5- and RBOH1-dependnet manner. The role of SOD, APX, MDAR, DHAR and GR as well as NPQ in photoprotection has been well established in plants including tomato (Foyer et al., 1995; Chen and Gallie, 2012; Duan et al., 2012). The results presented here show that PGR5-dependent CEF is important in photoprotection in tomato leaves experiencing cold stress (Fig. 7; Supplemental Figs. S9-S12). Similar to the apoplastic H$_2$O$_2$-dependent induction of the antioxidant response, the induction of CEF was also shown to be dependent on apoplastic H$_2$O$_2$ production (Fig. 6E). These observations are in agreement with earlier findings showing that H$_2$O$_2$ participates in the induction of CEF (Strand et al., 2015; Guo et al., 2016). In agreement with the role of CEF in the activation of ATP production and qE (Munekage et al., 2004; Guo et al., 2016; Yamori et al., 2016), we show that loss of PGR5 functions in the pgr5 mutant impaired H-FR-induced qE, NPQ, PsbS protein accumulation and increases in (A+Z)/(V+A+Z) ratios (Fig. 7, C-F). These results not only demonstrate the involvement of apoplastic H$_2$O$_2$ in the induction of ROS scavenging, CEF and NPQ, but also emphasize the roles of CEF in photoprotection.

**MATERIALS AND METHODS**

**Plant Material and Growth Conditions**

Wild-type tomato (Solanum lycopersicum) cv ‘Ailsa Craig’ and cv ‘Moneymaker’, and the phyA, phyB1B2, and phyAB1B2 mutants in the cv Moneymaker background were obtained from the Tomato Genetics Resource Center (http://tgrc.ucdavis.edu). HY5-RNAi, COP1-RNAi and RBOH1-RNAi plants were generated as described previously (Liu et al., 2004; Guo et al., 2016). These transgenic plants were identified by resistance to Basta and then by quantitative real-time (qRT)-PCR analysis for the transgene. For the generation of HY5 overexpressing transgenic plants, a 474 bp full-length HY5 cDNA fragment was obtained by RT-PCR using the primer pair HY5-OE-F with an Ascl site and HY5-OE-R with a SaII site (Supplemental Table S3). The PCR product was cloned into pFGC1008-HA vector behind the CaMV 35S promoter to generate the HY5-OE-HA clone. The tobacco rattle virus (TRV)-based vectors (pTRV1/2) were used for the virus-induced gene silencing (VIGS) of tomato HY5, ABI5 and RBOH1 genes with the specific PCR-amplified primers...
listed in Supplemental Table S3 (Liu et al., 2002). VIGS was performed as described previously (Xia et al., 2014).

PGR5 CRISPR/Cas9 vector was constructed as described by Pan et al. (2016). The target sequence (TTGGAAAGGCAGTGAGATCA) was designed using a web tool of CRISPR-P (Lei et al., 2014). The synthesized sequences were annealed and inserted into BbsI site of AtU6-sgRNA-AtUBQ-Cas9 vector, and the AtU6-sgRNA-AtUBQ-Cas9 cassette was inserted into the HindIII and KpnI sites of pCAMBIA1301 binary vector. To obtain the tomato PGR5 overexpressing construct, the 357 bp full-length coding DNA sequence (CDS) was amplified with the primers PGR5-OE-F and PGR5-OE-R (Supplemental Table S3) using tomato cDNA as the template. The PCR product was digested with Ascl and KpnI and inserted behind the CaMV 35S promoter in the plant transformation vector pFGC1008-HA. The resulting plasmids (HY5-OE-HA, PGR5 CRISPR/Cas9 vector and PGR5-OE-HA) were transformed into Agrobacterium tumefaciens strain EHA105, and then introduced into tomato seeds of Ailsa Craig via a method as previously described (Fillatti et al., 1987). Two independent homozygous lines of the F2 generation were used for the study. Two independent pgr5 lines, pgr5#4 and pgr5#5 which mutated at the first base of the protospacer adjacent motif (PAM) and stopped translation immediately (Supplemental Fig. S10, A-C).

Seedlings were grown in pots with a mixture of three parts peat to one part vermiculite, receiving Hoagland nutrient solution. The growth conditions were as follows: 12 h photoperiod, temperature of 25/20 °C (day/night), and photosynthetic photo flux density (PPFD) of 600 µmol m⁻² s⁻¹.

Cold, Light and Chemical Treatments

Plants at the 11-leaf-stage were used for the determination of spatial variation in photoinhibition. Experiments were carried out in growth rooms with a 12 h photoperiod, and a PPFD of 200 µmol m⁻² s⁻¹ by providing white light from directly above the plants. Light quality analysis revealed that R/FR ratios decreased from 1.3 at the 9th leaf rank to 0.5 at 5th leaf rank. Growth room temperatures were controlled at either 25 °C (optimal growth temperatures) or 4 °C (cold stress). Other light quality treatments were carried out in
controlled environment growth chambers (Conviron E15; Conviron, Manitoba, Canada) on
plants at the 6-leaf stage. Plants were grown under a 12/12 h light/dark cycle, with 85%
humidity. For these light quality treatments, plants were exposed to cold stress at 4 °C under
either high R/FR ratio (1.5), i.e. low FR intensity (L-FR, 133 µmol m⁻² s⁻¹) or low R/FR
ratio (0.5), i.e. high FR intensity (H-FR, 400 µmol m⁻² s⁻¹) light conditions. R light, supplied
by LED (λ_max = 660 nm, Philips, Netherlands), was maintained at 200 µmol m⁻² s⁻¹. FR was
supplied by a FR LED (λ_max = 735 nm, Philips, Netherlands). R/FR ratios were calculated via
the quantum flux densities measured between 655 and 665 nm divided by the quantum flux
densities measured between 730 and 740 nm.

To determine the cause of light-induced changes in photooxidizable P700 in plants
exposed to low growth temperatures, fully expanded leaves were excised from the plants at
6-leaf stage and put onto petri dishes containing either water or 25 µM methyl viologen
(MV). Leaves were allowed to float on either 100 mL of 25 µM MV or water for 3 h in
darkness at 25 °C. The petri dishes were then transferred to the 4 °C chambers and exposed
to different light quality (L-FR or H-FR) conditions (R/FR ratio, 1.5 or 0.5) for 6 h. The
maximum level of P700 photooxidation (ΔP700_max) was then determined in the MV-treated
leaves and water-treated controls using the Dual-PAM-100 system (Heinz Walz, Effeltrich,
Germany).

Cold Tolerance Assays

Cellular membrane permeability, measured as relative electrolyte leakage (REL), was
determined after 7 d exposure to the cold stress, as described previously (Cao et al., 2007).
Levels of oxidized leaf proteins were assayed by immunoblot detection as described
previously (Wang et al., 2016). Plant death was recorded after 6 days recovery from the cold
treatment, i.e. after return to optimal temperatures (25 °C) with a 12/12 h light/dark cycle
(PPFD of 600 µmol m⁻² s⁻¹) and 85% humidity.

Chlorophyll Fluorescence Measurements

Plants were dark-adapted for 30 min prior to measurement. The maximum quantum
yield of PSII (Fv/Fm) and NPQ were determined with the Imaging-PAM (IMAG-MAXI;
Heinz Walz, Effeltrich, Germany) as previously described (Jin et al., 2014). qE was simultaneously measured with the Dual-PAM-100 system (Heinz Walz, Effeltrich, Germany). Fluorescence quenching was induced by 10 min of actinic illumination with white light. The maximal fluorescence in the dark-adapted state (Fm) and in the light-adapted state (Fm') and after 10 min of dark relaxation following actinic illumination (Fm'') were determined using a saturating pulse of light applied at 2 min intervals. Energy-dependent quenching (qE) was calculated according to the equations $qE = \frac{Fm}{Fm'} - \frac{Fm}{Fm''}$ (Liu and Last, 2015).

P700 were measured simultaneously with the Dual-PAM-100 system (Heinz Walz, Effeltrich, Germany) after leaves had dark-adapted for 30 min (to obtain open reaction centers). The maximum P700 photooxidation level ($\Delta P700_{\text{max}}$) was determined using a saturation pulse (100 ms; 10,000 μmol m$^{-2}$ s$^{-1}$) under an FR background (720 nm; about 0.3 μmol m$^{-2}$ s$^{-1}$) according to the method of Klughammer and Schreiber (2008). The decrease in $\Delta P700_{\text{max}}$ is an indicator of PSI photoinhibition.

Post-illumination chlorophyll fluorescence (CEF around PSI) was monitored by the transient increase of dark-level chlorophyll fluorescence after actinic light (AL) illumination (250 μmol m$^{-2}$ s$^{-1}$ for 3 min) had been turned off by using a Dual-PAM-100 instrument (Heinz Walz, Effeltrich, Germany; Nashilevitz et al., 2010).

Activity of Antioxidant Enzymes and Pigment Analysis

Frozen leaf segments (0.3 g) were ground with 2 mL ice-cold buffer containing 50 mM PBS (pH 7.8), 0.2 mM EDTA, 2 mM AsA, and 2% (w/v) polyvinylpolypyrrolidone. The homogenates were centrifuged at 4 °C for 20 min at 12,000 g, and the resulting supernatants were used for the determination of enzymatic activity. The protein concentration was determined with bovine serum albumin as standard (Bradford, 1976). The activity of superoxide dismutase (SOD), ascorbate peroxidase (APX), monodehydroascorbate reductase (MDAR), dehydroascorbate reductase (DHAR) and glutathione reductase (GR) was measured following the protocol used as previously described (Xia et al., 2009). Total pigments were extracted as previously described (Xu et al., 2006). Xanthophyll cycle pigments (V, violaxanthin; A, antheraxanthin; Z, zeaxanthin) were analysed using a
C30 column (YMC Inc., Wilmington, NC) equipped for HPLC (Waters, Watford, Hertfordshire, United Kingdom) as described previously (Xu et al., 2006), with the following modification to the elution program. Mobile phases A (90% methanol), and B (tert-butyl methyl ether) were applied as follows: 92% A, 8% B, a linear gradient to 75% A and 25% B by 30 min, and gradient changed to 30% A, 70% B by 35 min, held until 50 min, then changed to 92% A and 8% B by 50.01 min, and then held to the end of analysis (60 min). The de-epoxidation state of the xanthophyll cycle pigments is defined as the \((A+Z)/(V+A+Z)\) ratio, where A, Z, and V are the concentrations of antheraxanthin, zeaxanthin and violaxanthin, respectively.

**Determination of Stomatal Aperture and Visualization of Cellular H\(_2\)O\(_2\) Accumulation**

Tomato stomatal apertures were measured as described previously (Xia et al., 2014) by peeling off the abaxial epidermises with forceps and floating it on a buffer containing 30 mM KCl, 10 mM 2-(N-morpholino)-ethanesulfonic acid (MES, pH 6.15). All images were captured using a light microscope equipped with a digital camera (Leica Microsystems, Wetzlar, Germany).

The localisation of H\(_2\)O\(_2\) accumulation in leaves was visualised at the subcellular level using cytochemical CeCl\(_3\) staining and transmission electron microscopy (H7650, Hitachi, Tokyo, Japan) as described previously (Xia et al, 2009).

**Thylakoid Isolation and Immunoblot Analysis**

Total protein was extracted from tomato leaves following exposure to a cold stress at 4 °C under either H-FR or L-FR light conditions for 1 d as described by Wang et al. (2016). After quantification of total protein concentrations, samples of 50 µg protein were separated by SDS-PAGE electrophoresis, and immuno-labelled with primary antibodies raised against PsbS (AS09533; Agrisera, Sweden). Following incubation with secondary anti-rabbit antibodies (Invitrogen, Sweden), enhanced chemical luminescence (ECL) was performed to detect labelled proteins.

Fractions of intact chloroplasts were prepared from (10 g) leaves harvested from tomato plants that had been grown at either 25 °C or 4 °C for 3 d under either H-FR or L-FR
conditions as described by Hertle et al. (2013). Thylakoid fractions were prepared from isolated chloroplasts by osmotic rupture. After centrifugation (4 °C, 14,000 g, 3 min), the pellet containing the thylakoid membranes was resuspended in a buffer containing 10 mM Tris/HCl (pH 6.8), 10 mM MgCl$_2$ and 20 mM KCl. The chlorophyll (Chl) concentration of the membranes was quantified spectrophotometrically as described by Porra et al. (1989). The thylakoid membranes (15 µg Chl at 1 mg Chl/mL) were solubilized using 2% (wt/vol) n-dodecyl-β-D-maltoside (β-DM; Anatrace), as described by Kromdijk et al. (2016). Following incubation at 30 min at 4 °C with gentle agitation, insoluble fractions were removed by centrifugation (15,000 g) for 10 min at 4°C. The solubilized membrane proteins were subjected to SDS-PAGE (15% polyacrylamide) electrophoresis. Proteins were then transferred on to nitrocellulose membranes (BioRad, Hercules, CA, USA), which were then incubated with antibodies against PsaB (AS10695; Agrisera, Sweden) or PsaC (AS10939; Agrisera, Sweden). Secondary antibodies use in these studies were anti-rabbit (Invitrogen, Sweden). Signal detection was using enhanced chemical luminescence (ECL).

RNA Extraction and qRT-PCR Analysis

Total RNA was extracted from tomato leaves using RNAprep Pure Plant Kit (Tiangen Biotech Co., Ltd. Beijing, China) according to the manufacturer’s instruction. Residual DNA was removed with RNase Mini Kit (Qiagen). The extracted RNA was reverse transcribed using a ReverTra Ace qPCR RT Kit (Toyobo, Osaka, Japan), following the manufacturer’s recommendations. qRT-PCR experiments were performed using a Power SYBR Green PCR Master Mix kit (Takara, Chiga, Japan). qRT-PCR was performed with 3 min at 95 °C, followed by 40 cycles of 30 s at 95 °C, 30 s at 58 °C and 1 min at 72 °C. The tomato ACTIN2 gene was used as an internal control. Primers sequence can be found in Supplemental Table S4. The relative gene expression was calculated following previously described formulae (Livak et al., 2001).

RNA-seq Analysis

For tomato RNA-seq analysis, leaves tissues from 6-leaf stage tomato seedlings were collected from L-FR and H-FR treatments after 6 h under 4 °C to conduct the RNA-seq analysis. Total RNA was isolated using TRIzol reagent (Biotopped) and RNA integrity was
evaluated using a Bioanalyzer 2100 (Agilent). The RNA samples were then subjected to RNA sequencing by LC Sciences. Genes with P value < 0.05 and fold change ≥ 2 were regarded as differentially expressed genes.

**Recombinant Protein and Electrophoretic Mobility Shift Assay**

The full-length coding region of HY5 was first PCR amplified using the primers in Supplemental Table S3, then, the product was digested with BamHI and SacI and ligated into the same sites of pET-32a vector. The recombinant vector was transformed into E. coli strain BL21 (DE3). The recombinant histidine-tagged HIS-HY5 proteins were induced by isopropyl β-D-1-thiogalactopyranoside and purified following the instructions of the Novagen pET purification system.

For binding assay, probes were biotin end-labeled following the instructions of the Biotin 3’ End DNA Labeling Kit (Pierce, 89818) and annealed to double-stranded probe DNA by incubating sequentially at 95 °C for 5 min, then the temperature decreased from 95 °C to 55 °C by 40 cycles (-1 °C/cycle, 1 cycle/min), 55 °C for 30 min, from 55 °C to 25 °C by 30 cycles (-1 °C/cycle, 1 cycle/min), finally, 4 °C for 5 min. EMSA of the HY5-DNA complexes was performed using biotin-labeled probes according to the instructions of the Light Shift Chemiluminescent EMSA kit (Thermo Fisher Scientific, 20148). Briefly, 0.5 μg of HY5 fusion proteins were incubated together with biotin-labeled probes in 20 μL reaction mixtures containing 10 mM Tris-HCl, 1 mM DTT, 150 mM KCl, 100 mM ZnCl₂, 50 ng μL⁻¹ poly (dl-dC), 2.5% glycerol, 0.05% Nonidet P-40, and 0.5 μg mL⁻¹ BSA for 20 min at room temperature and separated on 6% native polyacrylamide gels in Tris-glycine buffer at 100 V. After electrophoresis, the gel was dried and autoradiographed as described previously (Xu et al., 2014).

**Chromatin Immunoprecipitation (ChIP) Assay**

ChIP assays were performed following the instructions of the EpiQuik™ Plant ChIP Kit (Epigentek, P-2014) as described previously (Li et al., 2011). Approximately 1 g of leaf tissue was harvested from 35S-HY5-HA and wild-type plants after cold stress under H-FR condition. Chromatin was immunoprecipitated with an anti-HA antibody (Pierce, 26183),
and the goat anti-mouse IgG (Millipore, AP124P) was used as the negative control. Both immunoprecipitated DNA and input DNA were analyzed by qRT-PCR (Light Cycler; Roche). Primers for ChIP-qPCR of the ABI5 promoters were listed in Supplemental Table S5. Each ChIP value was normalized to its respective input DNA value. All ChIP-qPCR experiments were independently performed in triplicate.

**Statistical Analysis**

The experimental design was a completely randomized block design with four replicates. Each replicate contained 6-12 plants. Analysis of variance (ANOVA) was used to test for significance. When interaction terms were significant (P< 0.05), differences between means were analyzed using Turkey comparisons. Significant differences between treatment means are indicated by different letters.

Sequence data from this article can be found in the GenBank/EMBL data libraries under the accession numbers listed in Supplemental Table S4.

**Figure Legends**

**Figure 1.** Spatial variation in photoinhibition is partially attributable to the changes in light quality conditions. A and B, Maximum photochemical efficiency of PSII (Fv/Fm, A), maximum P700 photooxidation level (ΔP700\textsubscript{max}, B) in leaves at the 9\textsuperscript{th} (Up) and 5\textsuperscript{th} (Down) ranks from the base in plants at 11-leaf stage under white light conditions after exposure to 4 °C for 7 d. C and D, Fv/Fm (C) and ΔP700\textsubscript{max} (D) at 4\textsuperscript{th} leaves of the tomato plants at 6-leaf stage grown in temperature-controlled chambers at 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. E, Immunoblot detection of thylakoid proteins (PsaB and PsaC) separated by SDS-PAGE. Detached leaves were exposed to 25 °C or 4 °C for 3 d under L-FR or H-FR. F, Effect of methyl viologen (MV) on the ΔP700\textsubscript{max} under cold stress in different light quality. After treated with 25 μM MV for 3 h in darkness at 25 °C, leaves were transferred to 4 °C for 6 h under different light
quality conditions. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates except for Fv/Fm which was the mean for 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.

**Figure 2.** Roles for tomato phytochromes in the light quality regulation of photoinhibition and the expression of light signaling genes (HY5 and COP1). A, Fv/Fm and ΔP700max of the tomato phytochrome mutant plants after exposure to a cold at 4 °C under L-FR or H-FR light conditions for 7 d. B, Post-illumination chlorophyll fluorescence (CEF around PS I) in tomato plants after exposure to a cold at 4 °C for 3 d under L-FR and H-FR conditions. C and D, Changes of NPQ (C) and PsbS protein (D) in wild type (WT) and phytochrome mutant plants exposed for either 3 d or 1 d to cold stress (at 4 °C) under L-FR and H-FR light conditions. E, Levels of HY5 and COP1 transcripts at 6 h after tomato phytochrome mutants were exposed to 4 °C under L-FR or H-FR light conditions. For the L-FR and H-FR, R/FR ratio treatments of 1.5 or 0.5 respectively, plants were kept under R (200 µmol m⁻² s⁻¹) light conditions, supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.

**Figure 3.** HY5 alleviated photoinhibition by induction of photoprotection. A and B, Fv/Fm (A) and ΔP700max (B) of the wild type (WT), HY5-RNAi and COP1-RNAi tomato plants after exposure to a cold stress at 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, Post-illumination chlorophyll fluorescence (CEF around PSI, C) and NPQ (D) in WT, HY5-RNAi and COP1-RNAi tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. E, Immunoblot analysis of PsbS in WT, HY5-RNAi and COP1-RNAi tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal
total proteins amounts based on Coomassie blue. F, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the WT, HY5-RNAi and COP1-RNAi tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m$^{-2}$ s$^{-1}$) supplemented with different intensities of FR (133 and 400 µmol m$^{-2}$ s$^{-1}$). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.

**Figure 4.** HY5 induced transcript level of ABI5 by binding to promoter of ABI5. A and B, G-box elements in the promoter of tomato ABI5 gene (A) and oligonucleotide used in the electrophoretic mobility shift assays (EMSA, B). Numbering is from predicted transcriptional start sites. The ABI5 probe contains one G-box (ABI5-G-wt), whereas in the ABI5-G-mut1 and ABI5-G-mut2 probes the G-box core sequence was mutated. The WT and mutated G-box sequences are underlined. The mutated bases were indicated in red. C, HY5 directly binds to the G-box of ABI5 promoter in vitro. Recombinant HY5 was purified from E. coli cells and used for DNA binding assays with probes of ABI5-G-wt, ABI5-G-mut1 and ABI5-G-mut2. The protein purified from empty vector was used as the negative control. D, Direct binding of HY5 to the ABI5 promoter was analyzed using ChIP-qPCR in 35S-HY5-3HA-overexpressing (HY5-OE#1) tomato plants. HY5-OE#1 plants at 6-leaf stage were exposed to 4 °C under H-FR condition and input chromatin was isolated from leaf samples at 6 h. The epitope-tagged HY5-chromatin complex was immunoprecipitated with an anti-HA antibody. A control reaction was processed side-by-side using mouse IgG. Input- and ChIP-DNA samples were quantified by qRT-PCR using primers specific for the promoter of the ABI5 gene. The ChIP results are presented as percentage of the input DNA. OE, overexpressing; #1, line of HY5-OE plants. E and F, Transcript level of ABI5 gene at 6 h after HY5-RNAi and COP1-RNAi tomato plants exposed to 25 °C or 4 °C under different R/FR light regimes (E), and two independent HY5 overexpressing transgenic tomato lines (HY5-OE#1, OE#3) exposed to 4 °C under H-FR conditions (F). G, Transcript level of ABI5 gene at 6 h after WT and phytochromes mutants of tomato exposed to 4 °C under different
R/FR light regimes. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m\(^{-2}\) s\(^{-1}\)) supplemented with different intensities of FR (133 and 400 µmol m\(^{-2}\) s\(^{-1}\)). Four independent experiments were performed yielding similar results. Different letters indicate significant differences (P < 0.05) according to the Tukey’s test.

**Figure 5.** Role of ABI5 in light quality-regulated photoinhibition and photoprotection. A and B, Fv/Fm (A) and \(\Delta P700_{max}\) (B) of the non-silenced (pTRV) and silenced (pTRV-ABI5) tomato plants grown in temperature-controlled chambers at 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, Post-illumination chlorophyll fluorescence (CEF around PSI, C) and NPQ (D) in the pTRV and pTRV-ABI5 tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions.

E, Immunoblot analysis of PsbS in pTRV and pTRV-ABI5 tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal total proteins amounts based on Coomassie blue. F, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the pTRV and pTRV-ABI5 tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m\(^{-2}\) s\(^{-1}\)) supplemented with different intensities of FR (133 and 400 µmol m\(^{-2}\) s\(^{-1}\)). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P < 0.05) according to the Tukey’s test.

**Figure 6.** RBOH1-dependent ROS production prevents plants from photoinhibition by activating photoprotection. A and B, Transcript level of RBOH1 gene at 6 h (A) and cytochemical localization of H\(_2\)O\(_2\) accumulation in leaf mesophyll cells at 1 d as visualized by CeCl\(_3\) staining and TEM (B) after pTRV and pTRV-ABI5 tomato plants exposed to 4 °C under different R/FR light regimes. The arrows indicate CeCl\(_3\) precipitates. Scale bars = 0.5 µm. C and D, Fv/Fm (C) and \(\Delta P700_{max}\) (D) of the wild type (WT) and RBOH1-
RNAi tomato plants were exposed to 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. E and F, Post-illumination chlorophyll fluorescence (CEF around PSI, E) and NPQ (F) in the WT and RBOH1-RNAi tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. G, Immunoblot analysis of PsbS in WT and RBOH1-RNAi tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal total proteins amounts based on Coomassie blue. H, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the WT and RBOH1-RNAi tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.

Figure 7. PROTON GRADIENT REGULATION5 (PGR5)-dependent CEF plays dual roles in preventing plants from photoinhibition. A and B, Fv/Fm (A) and ΔP700 max (B) of the wild type (WT), pgr5 mutant (pgr5#5) and PGR5-overexpressing (OE-PGR5#3) transgenic plants grown at 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, qE (C) and NPQ (D) in the WT, pgr5#5 mutant and OE-PGR5#3 tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. E and F, PsbS protein (E) and de-epoxidation state of the xanthophyll cycle (F) in the WT, pgr5#5 mutant and OE-PGR5#3 tomato plants after exposure to 4 °C for 1 d and 3 d, respectively, under L-FR and H-FR conditions. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates, except for Fv/Fm ratios, which are the means for 15 leaves from independent plants.
independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.

**Supplemental Data**

The following supplemental materials are available.

**Supplemental Figure S1.** Effect of spatial variation on the cold tolerance of tomato.

**Supplemental Figure S2.** Effect of FR intensity on the cold tolerance and photoprotective response of tomato.

**Supplemental Figure S3.** Silencing efficiency and cold tolerance of HY5-RNAi and COP1-RNAi tomato plants.

**Supplemental Figure S4.** Cold tolerance of HY5-overexpressing transgenic tomato lines.

**Supplemental Figure S5.** Transcripts of genes involved in Foyer-Halliwell-Asada cycle in WT, HY5-RNAi and COP1-RNAi tomato plants.

**Supplemental Figure S6.** HY5 regulated ABA-mediated stomatal movement and directly binds to the G-boxes of the ABI promoters in vitro.

**Supplemental Figure S7.** Silencing efficiency and cold tolerance of ABI5 silenced plants.

**Supplemental Figure S8.** Cold tolerance of WT and RBOH1-RNAi transgenic tomato plants.

**Supplemental Figure S9.** Relative expression of CEF related genes in response to cold stress and far red light.

**Supplemental Figure S10.** Transgenic tomato of pgr5 mutant and PGR5-overexpressing plants.

**Supplemental Figure S11.** Cold tolerance and cyclic electron flux (CEF) around PSI in pgr5 mutant and OE-PGR5 plants.

**Supplemental Figure S12.** Changes of ΔP700max in WT and OE-PGR5 tomato plants as altered by the silencing of HY5, ABI5 or RBOH1.

**Supplemental Table S1.** Differentially expressed genes of tomato plants after exposure to a cold at 4 °C under H-FR and L-FR light conditions.
**Supplemental Table S2.** Differentially expressed genes in the photosystems and photoprotection of tomato plants after exposure to a cold at 4 °C under H-FR and L-FR light conditions.

**Supplemental Table S3.** PCR primer sequences used for vector construction.

**Supplemental Table S4.** List of primer sequences used for qRT-PCR analysis.

**Supplemental Table S5.** Primers used for ChIP-qPCR assays.

**LITERATURE CITED**


resistance to Botrytis cinerea and jasmonate responses via a COI1-JAZ10-dependent, salicylic acid-independent mechanism. Plant Physiol 158: 2042-2052


Kingston-Smith AH, Foyer CH (2000) Bundle sheath proteins are more sensitive to oxidative damage than those of the mesophyll in maize leaves exposed to paraquat or low temperatures. J Expt Bot 51: 123-130


Li ZF, Zhang LX, Yu YW, Quan RD, Zhang ZJ, Zhang HW, Huang RF (2011) The ethylene response factor AtERF11 that is transcriptionally modulated by the bZIP transcription factor HY5 is a crucial repressor for ethylene biosynthesis in Arabidopsis. Plant J 68: 88-99


Murata Y, Pei ZM, Mori IC, Schroeder JI (2001) Abscisic acid activation of plasma membrane Ca\textsuperscript{2+} channels in guard cells requires cytosolic NADPH and is differentially disrupted upstream and downstream of reactive oxygen species production in abi1-1 and abi2-1 protein phosphatase 2C mutants. Plant Cell 13: 2513-2523


Figure 1. Spatial variation in photoinhibition is partially attributable to the changes in light quality conditions. A and B, Maximum photochemical efficiency of PSII (Fv/Fm, A), maximum P700 photooxidation level (ΔP700max, B) in leaves at the 9th (Up) and 5th (Down) ranks from the base in plants at 11-leaf stage under white light conditions after exposure to 4 °C for 7 d. C and D, Fv/Fm (C) and ΔP700max (D) at 4th leaves of the tomato plants at 6-leaf stage grown in temperature-controlled chambers at 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. E, Immunoblot detection of thylakoid proteins (PsaB and PsaC) separated by SDS-PAGE. Detached leaves were exposed to 25 °C or 4 °C for 3 d under L-FR or H-FR. F, Effect of methyl viologen (MV) on the ΔP700max under cold stress in different light quality. After treated with 25 µM MV for 3 h in darkness at 25 °C, leaves were transferred to 4 °C for 6 h under different light quality conditions. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates except for Fv/Fm which was the mean for 15 leaves from independent plants. Different letters indicate significant differences (P<0.05) according to the Tukey’s test.
Figure 2. Roles for tomato phytochromes in the light quality regulation of photoinhibition and the expression of light signaling genes (HY5 and COP1). A, Fv/Fm and \( \Delta P_{700}^{\text{max}} \) of the tomato phytochrome mutant plants after exposure to a cold at 4 °C under L-FR or H-FR light conditions for 7 d. B, Post-illumination chlorophyll fluorescence (CEF around PS I) in tomato plants after exposure to a cold at 4 °C for 3 d under L-FR and H-FR conditions. C and D, Changes of NPQ (C) and PsbS protein (D) in wild type (WT) and phytochrome mutant plants exposed for either 3 d or 1 d to cold stress (4 °C) under L-FR and H-FR light conditions. E, Levels of HY5 and COP1 transcripts at 6 h after tomato phytochrome mutants were exposed to 4 °C under L-FR or H-FR light conditions. For the L-FR and H-FR, R/FR ratio treatments of 1.5 or 0.5 respectively, plants were kept under R (200 µmol m\(^{-2}\) s\(^{-1}\)) light conditions, supplemented with different intensities of FR (133 and 400 µmol m\(^{-2}\) s\(^{-1}\)). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.
Figure 3. HY5 alleviated photoinhibition by induction of photoprotection. A and B, Fv/Fm (A) and ΔP700_max (B) of the wild type (WT), HY5-RNAi and COP1-RNAi tomato plants after exposure to a cold stress at 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, Post-illumination chlorophyll fluorescence (CEF around PSI, C) and NPQ (D) in WT, HY5-RNAi and COP1-RNAi tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. E, Immunoblot analysis of PsbS in WT, HY5-RNAi and COP1-RNAi tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal total proteins amounts based on Coomassie blue. F, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the WT, HY5-RNAi and COP1-RNAi tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.
Figure 4. HY5 induced transcript level of ABI5 by binding to promoter of ABI5. A and B, G-box elements in the promoter of tomato ABI5 gene (A) and oligonucleotide used in the electrophoretic mobility shift assays (EMSA, B). Numbering is from predicted transcriptional start sites. The ABI5 probe contains one G-box (ABI5-G-wt), whereas in the ABI5-G-mut1 and ABI5-G-mut2 probes the G-box core sequence was mutated. The WT and mutated G-box sequences are underlined. The mutated bases were indicated in red. C, HY5 directly binds to the G-box of ABI5 promoter in vitro. Recombinant HY5 was purified from E. coli cells and used for DNA binding assays with probes of ABI5-G-wt, ABI5-G-mut1 and ABI5-G-mut2. The protein purified from empty vector was used as the negative control. D, Direct binding of HY5 to the ABI5 promoter was analyzed using ChIP-qPCR in 35S-HY5-3HA-overexpressing (HY5-OE#1) tomato plants. HY5-OE#1 plants at 6-leaf stage were exposed to 4 °C under H-FR condition and input chromatin was isolated from leaf samples at 6 h. The epitope-tagged HY5-chromatin complex was immunoprecipitated with an anti-HA antibody. A control reaction was processed side-by-side using mouse IgG. Input- and ChIP-DNA samples were quantified by qRT-PCR using primers specific for the promoter of the ABI5 gene. The ChIP results are presented as percentage of the input DNA. OE, overexpressing; #1, line of HY5-OE plants. E and F, Transcript level of ABI5 gene at 6 h after HY5-RNAi and COP1-RNAi tomato plants exposed to 25 °C or 4 °C under different R/FR light regimes (E), and two independent HY5 overexpressing transgenic tomato lines (HY5-OE#1, OE#3) exposed to 4 °C under H-FR conditions (F). G, Transcript level of ABI5 gene at 6 h after WT and phytochromes mutants of tomato exposed to 4 °C under different R/FR light regimes. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Four independent experiments were performed yielding similar results. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.
Figure 5. Role of ABI5 in light quality-regulated photoinhibition and photoprotection. A and B, Fv/Fm (A) and ΔP700_max (B) of the non-silenced (pTRV) and silenced (pTRV-ABI5) tomato plants grown in temperature-controlled chambers at 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, Post-illumination chlorophyll fluorescence (CEF around PSI, C) and NPQ (D) in the pTRV and pTRV-ABI5 tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. E, Immunoblot analysis of PsbS in pTRV and pTRV-ABI5 tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal total proteins amounts based on Coomassie blue. F, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the pTRV and pTRV-ABI5 tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m⁻² s⁻¹) supplemented with different intensities of FR (133 and 400 µmol m⁻² s⁻¹). Data are presented as the mean of 4 biological replicates (±SD) except for Fv/Fm which was the mean for 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.
Figure 6. RBOH1-dependent ROS production prevents plants from photoinhibition by activating photoprotection. A and B, Transcript level of RBOH1 gene at 6 h (A) and cytochemical localization of H$_2$O$_2$ accumulation in leaf mesophyll cells at 1 d as visualized by CeCl$_3$ staining and TEM (B) after pTRV and pTRV-ABI5 tomato plants exposed to 4 °C under different R/FR light regimes. The arrows indicate CeCl$_3$ precipitates. Scale bars = 0.5 µm. C and D, Fv/Fm (C) and ΔP700$^{\max}$ (D) of the wild type (WT) and RBOH1-RNAi tomato plants were exposed to 25 °C or 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. E and F, Post-illumination chlorophyll fluorescence (CEF around PSI, E) and NPQ (F) in the WT and RBOH1-RNAi tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. G, Immunoblot analysis of PsbS in WT and RBOH1-RNAi tomato plants after exposure to 4 °C for 1 d under L-FR and H-FR conditions. Samples were loaded at equal total proteins amounts based on Coomassie blue. H, Activity of antioxidant enzymes (SOD, APX, MDAR, DHAR and GR) involved in Foyer-Halliwell-Asada cycle after the WT and RBOH1-RNAi tomato plants exposure to 25 °C or 4 °C under L-FR or H-FR light conditions for 3 d. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m$^{-2}$ s$^{-1}$) supplemented with different intensities of FR (133 and 400 µmol m$^{-2}$ s$^{-1}$). Data are the means (±SD) of 4 biological replicates except for Fv/Fm ratios, which are the means of 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.
Figure 7. PROTON GRADIENT REGULATION5 (PGR5)-dependent CEF plays dual roles in preventing plants from photoinhibition. A and B, Fv/Fm (A) and ΔP700\textsubscript{max} (B) of the wild type (WT), pgr5 mutant (pgr5#5) and PGR5-overexpressing (OE-PGR5#3) transgenic plants grown at 4 °C under L-FR or H-FR light conditions for 7 d. The false color code depicted at the bottom of the image ranges from 0 (black) to 1.0 (purple) represents the level of damage in leaves. C and D, qE (C) and NPQ (D) in the WT, pgr5#5 mutant and OE-PGR5#3 tomato plants after exposure to 4 °C for 3 d under L-FR and H-FR conditions. E and F, PsbS protein (E) and de-epoxidation state of the xanthophyll cycle (F) in the WT, pgr5#5 mutant and OE-PGR5#3 tomato plants after exposure to 4 °C for 1 d and 3 d, respectively, under L-FR and H-FR conditions. For the L-FR and H-FR, R/FR ratio at 1.5 and 0.5, respectively, plants were kept at R conditions (200 µmol m\textsuperscript{-2} s\textsuperscript{-1}) supplemented with different intensities of FR (133 and 400 µmol m\textsuperscript{-2} s\textsuperscript{-1}). Data are the means (±SD) of 4 biological replicates, except for Fv/Fm ratios, which are the means for 15 leaves from independent plants. Different letters indicate significant differences (P< 0.05) according to the Tukey’s test.


Duan M, Ma NN, Li D, Deng YS, Kong FY, Lv W, Meng QW (2012) Antisense-mediated suppression of tomato thylakoidal ascorbate


Kingston-Smith AH, Foyer CH (2000) Bundle sheath proteins are more sensitive to oxidative damage than those of the mesophyll in maize leaves exposed to paraquat or low temperatures. J Expt Bot 51: 123-130


Murata Y, Pei ZM, Mori IC, Schroeder JJ (2001) Abscisic acid activation of plasma membrane Ca2+ channels in guard cells requires cytosolic NADPH and is differentially disrupted upstream and downstream of reactive oxygen species production in abi1-1 and abi2-1 protein phosphatase 2C mutants. Plant Cell 13: 2533-2543


