Novel molecular subgroups for clinical classification and outcome prediction in childhood medulloblastoma: a cohort study


Summary

Background International consensus recognises four medulloblastoma molecular subgroups: WNT (MB WNT), SHH (MB SHH), group 3 (MB Grp3), and group 4 (MB Grp4), each defined by their characteristic genome-wide transcriptomic and DNA methylation profiles. These subgroups have distinct clinicopathological and molecular features, and underpin current disease subclassification and initial subgroup-directed therapies that are underway in clinical trials. However, substantial biological heterogeneity and differences in survival are apparent within each subgroup, which remain to be resolved. We aimed to investigate whether additional molecular subgroups exist within childhood medulloblastoma and whether these could be used to improve disease subclassification and prognosis predictions.

Methods In this retrospective cohort study, we assessed 428 primary medulloblastoma samples collected from UK Children’s Cancer and Leukaemia Group (CCLG) treatment centres (UK), collaborating European institutions, and the UKCCSG-SIOP-PNET3 European clinical trial. An independent validation cohort (n=276) of archival tumour samples was also analysed. We analysed samples from patients with childhood medulloblastoma who were aged 0–16 years at diagnosis, and had central review of pathology and comprehensive clinical data. We did comprehensive molecular profiling, including DNA methylation microarray analysis, and did unsupervised class discovery of test and validation cohorts to identify consensus primary molecular subgroups and characterise their clinical and biological significance. We modelled survival of patients aged 3–16 years in patients (n=215) who had craniospinal irradiation and had been treated with a curative intent.

Findings Seven robust and reproducible primary molecular subgroups of childhood medulloblastoma were identified. MB SHH remained unchanged and each remaining consensus subgroup was split in two. MB SHH was split into age-dependent subgroups corresponding to infant (<4.3 years; MB SHH-infant; n=65) and childhood patients (≥4.3 years; MB SHH-child; n=38). MB Grp3 and MB Grp4 were each split into high-risk (MB Grp3-HR [n=65] and MB Grp4-HR [n=85]) and low-risk (MB Grp3-LR [n=50] and MB Grp4-LR [n=73]) subgroups. These biological subgroups were validated in the independent cohort. We identified features of the seven subgroups that were predictive of outcome. Cross-validated subgroup-dependent survival models, incorporating these novel subgroups along with secondary clinicopathological and molecular features and established disease risk-factors, outperformed existing disease risk-stratification schemes. These subgroup-dependent models stratified patients into four clinical risk groups for 5-year progression-free survival: favourable risk (54 [25%] of 215 patients; 91% survival [95% CI 82–100%]); standard risk (50 [23%] patients; 81% survival [70–94%]); high-risk (82 [38%] patients; 42% survival [31–56%]); and very high-risk (29 [13%] patients; 28% survival [14–56%]).

Interpretation The discovery of seven novel, clinically significant subgroups improves disease risk-stratification and could inform treatment decisions. These data provide a new foundation for future research and clinical investigations.

Funding Cancer Research UK, The Tom Grahame Trust, Star for Harris, Action Medical Research, SPARKS, The JGW Patterson Foundation, The INSTINCT network (co-funded by The Brain Tumour Charity, Great Ormond Street Children’s Charity, and Children with Cancer UK).

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Introduction

The discovery of molecular disease subgroups represents the most fundamental advance in our understanding of medulloblastoma, the most common malignant brain tumour of childhood. Current international consensus recognises four subgroups of medulloblastoma: WNT (MB WNT), SHH (MB SHH), group 3 (MB Grp3) and group 4 (MB Grp4). Each subgroup is defined empirically by genome-wide transcriptomic and DNA methylation patterns and characterised by distinct clinicopathological and molecular features. By contrast, MB SHH and MB Grp4 have few mutations, but have multiple DNA copy number alterations.
Subgrouping is integral to the 2016 WHO medulloblastoma classification, and is used to direct treatment strategies aimed at improving cure rates (5-year survival across all four subgroups is 65–70%), and reducing long-term intellectual and neuroendocrine impairments associated with existing multimodality therapies. Patients with childhood MB are consistently shown to have a favourable prognosis (>90% 5-year survival14,15) and reduced intensity risk-adapted therapies are being studied in these patients in international clinical trials, whereas SHH pathway inhibitors show promise in MB disease in early-phase trials, although treatment of infants (younger than 3 years at diagnosis) and young children with these inhibitors should be approached with caution, because of the risk of premature fusion of growth-plates.8

Substantial biological heterogeneity is evident within each non-MBWNT subgroup; for instance, TP53 mutations are associated with a poor outcome in MB.11,12 High-risk clinical factors (metastatic disease [M+]; large-cell, anaplastic [LCA] pathology; incomplete surgical resection [R+]; and MYC/MYCN amplification), which are currently used to stratify risk in medulloblastoma in children aged 3 years or older, were derived from cohort-wide investigations before discovery of the consensus subgroups, and thus did not consider their effect.15,16,20,21 Studies that defined the four-subgroup consensus used modestly sized cohorts (typically fewer than 200 patients).2–6 In this Article, we describe comprehensive molecular profiling of clinically annotated discovery and validation cohorts totalling more than 700 tumours. We report the discovery and characterisation of seven stable and reproducible primary subgroups of childhood medulloblastoma (in patients younger than 16 years at diagnosis), which subdivide each of the classic consensus non-MBWNT subgroups (MB, MB, and MB) into two clinically significant groups with distinct clinicomolecular features and survival outcomes.

Methods

Study design and participants
In this retrospective cohort study, we assessed 428 centrally reviewed, clinically annotated primary medulloblastomas from patients aged 0–16 years at diagnosis, collected from UK Children’s Cancer and Leukaemia Group (CCLG) treatment centres (UK; 366 [86%]), collaborating European institutions in Budapest (Hungary; 20 [5%]) and Warsaw (Poland; 15 [4%]), and samples from the European UKCCSG-SIOP-PNET3 clinical trial (27 [6%]). As is typical for medulloblastoma, we regarded patients younger than 3 years at diagnosis as infants. 108 (26%) of 408 patient samples used were collected in 2010–14, 192 (47%) in 2000–10, 85 (21%) in 1990–2000, and the remaining 23 (6%) were collected before 1990 (18 were from the 1980s, four from the 1970s, and one was from 1968). Year of diagnosis was unavailable for 20 samples.

Tumour samples were provided by the UK CCLG as part of CCLG-approved biological study BS-2007–04; informed, written consent was obtained from parents of all patients because all assessed patients were younger than 16 years. Tumour investigations were done with...
and quality was available. We identified subgroup-specific differentially expressed genes using DESeq2, and these genes were included in ontology enrichment analyses (appendix p 4). We identified GF11 mutations from RNA-seq data (appendix p 4).

MB\textsubscript{MB} mutation data were obtained from a previous study. Although 450K methylation data for MB\textsubscript{MB} subgroup assignment were not available for this sample cohort, the tightly defined age cutoff that we defined between the molecularly determined MB\textsubscript{MB} and MB\textsubscript{MB-Child} subgroups enabled us to infer subgroups for this sequencing cohort (appendix p 4). We tested recurrent MB\textsubscript{MB} mutations (TP53, SUFU, PTCH1, SMO, and TERT) and gene amplifications (MYCN and GLI2) identified by whole genome sequencing, for association with the age-defined MB\textsubscript{MB-Child} or MB\textsubscript{MB-Infant} subgroups using Fisher’s exact test (appendix p 4).

### Statistical analysis

We did survival analyses (overall survival and progression-free survival) on samples from patients aged 3–16 years within our discovery cohort, who received maximal surgical resection and craniospinal irradiation with curative intent. Overall survival was defined as the time from date of surgery to death or date of last follow-up and progression-free survival as the time from date of surgery to first event (progression or relapse) or date of last follow-up. Patients with follow-up time that exceeded 10 years were right-censored at 10 years.

The tightly defined age cutoff between the molecularly determined MB\textsubscript{MB-Infant} and MB\textsubscript{MB-Child} subgroups enabled us to assess an expanded survival cohort of MB\textsubscript{MB-Child} disease (n=55), including additional samples with insufficient DNA for methylation array analysis, classified as MB\textsubscript{MB-Child} on the basis of their age (appendix p 4). In this group, we assessed the prognostic potential of currently used clinical and molecular risk markers (M+ disease, R+ disease, LCA pathology, sex, MYCN amplification, TERT mutation, and TP53 mutation [appendix pp 4–5]). Patients in the MB\textsubscript{MB-Infant} subgroup were typically younger than 3 years at diagnosis and were, therefore, treated on infant protocols. Treatment in this group of patients is heterogeneous, and is focused on omitting or delaying radiotherapy to reduce treatment-associated morbidities as far as possible. As a consequence, we report only overall survival in this group.

We created univariate and cross-validated multivariate Cox models based on subgroups, established risk factors, and cytogenetic changes. Prognostic markers in the multivariate analysis were identified by performing 100 rounds of 10-fold cross-validation, evaluating the performance of markers by measuring area under the curve (AUC) at 5 years for progression-free survival in the left out fold, and calculating the overall mean AUC over all rounds (appendix p 5). We added variables conferring an increase in AUC, as measured by time-dependent receiver operating characteristic curves at 5 years, to the model.

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**Procedures**

We tested medulloblastoma samples with the Illumina HumanMethylation450K DNA methylation array (Illumina, San Diego, CA, USA). The Gene Expression Omnibus accession number for 450K DNA methylation array profiles we used for the determination of human medulloblastoma molecular subgroup status is GSE93646.

To identify methylation-dependent subgroups, we did unsupervised class discovery by NMF-metagen and k-means clustering, testing all combinations of 3–10 metagenes and clusters for reproducibility using bootstrapped resampling methods (250 iterations) as described previously. This analysis identified metagenes (a single score that reflects the methylation status of several CpG loci) representing the main biological effects present in the genome-wide dataset. We assessed cluster stability using the cophenetic index, a shorthand measure of the robustness of sample clustering as determined by consensus non-negative matrix factorisation (appendix p 3). We visualised clusters with t-SNE.

We assessed samples classified with less than 80% confidence (by resampling procedures) as non-classifiable (NC; appendix pp 2–3).

We projected metagenes derived from our discovery cohort onto the validation cohort. Additionally, we combined the discovery and validation cohorts to do equivalent consensus clustering.

We assessed established medulloblastoma clinical, pathological, and molecular features as described previously. Briefly, we defined histopathological variants according to the WHO 2016 guidelines. We assigned metastatic status (M+) based on Chang’s criteria (appendix p 3). Tumours were designated as R+ if their residuum after surgical excision exceeded 1.5 cm\textsuperscript{2}. Pathology was centrally reviewed by three experienced neuropathologists for 380 (89%) of 428 samples, and clinical data were collated from contributing centres and reviewed centrally (appendix p 3). We assessed MYC and MYCN status by fluorescence in situ hybridisation or copy-number estimates from methylation array. We assessed TP53, CTNNB1, and TERT mutation status by Sanger sequencing. We identified subgroup-specific differentially methylated CpG loci or methylated regions (DMRs) using limma or DMRcate (appendix p 3). RNA-seq expression data were generated for discovery cohort samples for which mRNA of sufficient quantity and quality was available. We identified subgroup-specific differentially expressed genes using DESeq2, and these genes were included in ontology enrichment analyses (appendix p 4).

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**See Online for appendix**
Because MBGrp3 and MBGrp4 shared a metagene (V1), which defined a low-risk outcome and implied shared biology, we considered MBGrp3/4 as a single entity, and MBGrp3 and MBGrp4 separately for creation of survival models. In addition to currently understood clinical and molecular risk markers in these groups (M+ disease, R+ disease, LCA pathology, gender, MYC/MYCN amplification, and i17q [isochromosome 17q]), we additionally tested for recurrent cytogenetic changes, MBGrp3 membership, and membership of the high-risk methylomic group composed of members from both MBGrp3 and MBGrp4, defined by metagene V1 (appendix pp 5–6). We categorised independent prognostic markers into risk-stratification schemes (favourable-risk, >90% survival; standard-risk, 75–90% survival; high-risk, 40–75% survival; very high-risk, <40% survival) and survival-dependent ROC analysis of progression-free survival at 5 years, to assess performance by comparison with previously reported classification schemes (appendix pp 5–6).

We constructed Kaplan-Meier curves and compared patient groups with log-rank tests. For Kaplan-Meier comparison of two groups, we calculated hazard ratios (HRs) for the 0–5 year survival interval and 95% CIs from the Wald statistic. We tested the proportionality assumption for Cox modelling using scaled Schoenfeld residuals. Missing data were assumed to be missing completely at random and affected samples were removed from multivariate analyses. We implemented array processing, normalisation, quality-control checks, and copy-number estimation, relative to a panel of 18 normal cerebella with the R packages minfi and conumee (appendix p 2).

The significance threshold was set at p<0.05 for all statistical tests in this study, unless otherwise stated. Significance of association was assessed using Fisher’s exact and chi-squared tests with Yates’ continuity correction. We identified subgroup-specific age-differences between the non-MB WNT or non-MB SHH medulloblastoma subgroups using ANOVA (appendix p 4). Statistical or bioinformatics analyses were done using R (version 3.2.3).

Role of the funding source
The funders of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The corresponding author had full access to all of the data and had the final responsibility to submit for publication.

Results
Clinicopathological and molecular diagnostic characteristics of 428 patients younger than 16 years who had primary childhood medulloblastoma (discovery cohort) are shown in table I. Consensus analysis identified two equally robust cluster solutions (cophenetic index 0.998 [four metagenes] and 0.997 [six metagenes]; appendix p 10). The first cluster solution (four metagenes, four clusters) recapitulated the established four-subgroup consensus, whereas the second (six metagenes, seven clusters) revealed further clusters within the established subgroups (figure 1A, appendix pp 10–11).

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MBWNT tumours formed a single subgroup (n=33) characterised by CTNNB1 mutations, loss of chromosome 6, and an expected favourable prognosis (5-year overall survival: 93% [95% CI 82–100]; figure 1B). Our newly detected metagenes split each remaining consensus subgroup (MBGrp3, MBGrp4, and MBGrp5) in two. MBGrp3 was split into age-dependent subgroups corresponding to infant (<4·3 years; MB SHH-Infant; n=65) and childhood patients (≥4·3 years; MB SHH-Child; n=38) by the respective absence or presence of metagene V4. Both have intermediate prognoses (5-year overall survival MB SHH-Infant: 58% [95% CI 41–82]; MB SHH-Infant: 62% [50–77]; figure 1B). MBGrp3 and MBGrp4 are each split into high-risk (MB Grp3-HR [n=65] and MB Grp4-HR [n=85]) and low-risk (MB Grp3-LR [n=50] and MB Grp4-LR [n=73]) subgroups by common metagene V1 (figure 1A). 5-year overall survival was 37% [95% CI 25–53] in the MB Grp3-LR subgroup, 69% [55–87] in the MB Grp3-HR subgroup, 69% [58–83] in the MB Grp4-LR subgroup, and 80% [70–92] in the MB Grp4-HR subgroup (figure 1B). The subdivision of MB Grp3 and MB Grp4 distinguishes patients with a superior stratification (5-year overall survival AUC 0·610). Moreover, in the patients aged 3–16 years at diagnosis and receiving craniospinal irradiation, the high-risk or low-risk subdivision of MBGrp3/4 stratifies this diagnosis and receiving craniospinal irradiation, the current consensus MBGrp3 and MB Grp4 subgroups (AUC 0·649) [MB Grp3/4 combined subgroup into standard (MB Grp3-LR 81% [95% CI 60–100%]; MB Grp4-LR 81% [71–93%]) and high-risk (MB Grp3-HR 35% [23–55%]; MB Grp4-HR 47% [34–66%]) 5-year progression-free survival outcomes, by contrast with the current consensus MBGrp3 designations, which show intermediate outcomes (figure 1C, 1D).

Clinicopathological and biological features were non-randomly distributed in all seven subgroups (figure 1A, appendix pp 12–15). Patients in the MB SHH-Infant subgroup had significantly enriched desmoplastic or nodular pathology compared with all other subgroups (p<0·0001), and TP53 mutation (p=0·0001) and MYCN amplifications (p<0·0001) were significantly more frequent in MB SHH-Infant than in all other subgroups. Patients in the MB Grp3-LR subgroup significantly more frequently had LCA pathology (p<0·0001) and MYC amplification (p<0·0001), than all other subgroups. Although patients in the MB Grp3-HR and MB Grp4-HR subgroups had similar 10-year overall survival (22% [95% CI 10–46] vs 36% [22–59]; figure 1B), patients in the MB Grp3-LR subgroup died later of their disease (ten [36%] of 28 deaths in the MB Grp3-LR subgroup occurred more than 5 years after diagnosis) than did those in the MB Grp4-LR subgroup (33 [92%] of 36 deaths occurred within 5 years of diagnosis; appendix p 26).

Validation by projection of six metagenes onto an independent cohort of 276 patients (table I) confirmed their existence (appendix pp 10–11). Moreover, reapplying consensus clustering to the combined cohort of 704 patients confirmed a seven subgroup model as optimal, giving 100% concordance to the classifications derived separately from our discovery cohort (appendix pp 10–11).

Age distributions differed between the two MB SHH subgroups; age distributions are log-normally distributed and intersect at 4·3 years (figure 2A). The two peak incidences of age at diagnosis in infants and in older children for MB SHH disease,26 when observed as a whole, are resolved by their classification into distinct MB SHH-Infant and MB SHH-Child subgroups (appendix pp 12–13). Each MB SHH subgroup possesses characteristic molecular or clinicopathological features (appendix pp 12–13). LCA pathology (p=0·00050), MYCN amplification (p<0·0001), and mutations of TP53 (p<0·0001) and TERT (p<0·0015) were all significantly enriched in the MB SHH-Infant subgroup compared with the MB SHH-Child subgroup; whereas gender, M+ disease status, and R+ disease status were not significantly different between groups (figure 2B; appendix pp 12–13). TERT promoter mutation and MYCN amplification or LCA pathology were mutually exclusive (figure 2B; appendix pp 12–13). Mutational data from an independent MB SHH cohort26 showed that SUFU mutation was significantly associated with MB SHH-Infant whereas PTCH1 mutations were observed in both MB SHH subgroups (figure 2C). GLI2 amplification, MYCN amplification, and TP53 mutations (both somatic and germline) were significantly associated with the MB SHH-Child subgroup (figure 2C).

Compared with normal cerebella and patients in the MB SHH-Infant subgroup, patients in the MB SHH-Child subgroup had subgroup-specific DNA methylation changes.

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Table 1: Demographics and clinicopathological characteristics of all cohorts

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<th>Discovery cohort (n=428)</th>
<th>Validation cohort (n=276)</th>
<th>MB SHH-Infant survival cohort (n=55)</th>
<th>MB SHH-Child survival cohort (n=175)</th>
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<th>MBWNT subgroup assignment</th>
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Data are n (%) or median (range), unless otherwise specified. MB=medulloblastoma. SHH=sonic hedgehog. TP53=TP53 mutation. MYCN=myc amplification. PTCH1=PTCH1 mutation. CTX=chemotherapy. RTX=radiotherapy. WNT=wnt/wingless. M–=non-metastatic disease. M+=metastatic disease.
(predominantly hypermethylation), at both individual CpG loci and at the gene level (figure 2B; appendix pp 12–13), frequently involving developmental genes (79% of 384 genes with gene ontology term embryonic morphogenesis had aberrant hypermethylation). DNA methylation changes were validated in an independent cohort (appendix pp 12–13). When discovery cohort MB1111 RNA-seq expression data were available (190 [44%] of 428 samples), significant differential expression was observed between the subgroups (1593 genes, fold change >1.5; adjusted p<0.01; appendix pp 12–13). Although there were few recurrent cytogenetic changes, many tumours in patients in the MB1111-Child Subgroup (18 [51%] of 35 tumours) had loss of chromosome 9q, often associated with gain of 9p (appendix pp 12–13).

Figure 1: Novel clinically significant subgroups within the established medulloblastoma subgroups

(A) Non-negative matrix factorisation consensus clustering of methylome data from 428 primary medulloblastomas. Each column represents one patient. Missing data are shown in grey. Residuals from χ² tests indicate where subgroup-enrichment has occurred (darker shades of grey indicate stronger relationships), p values are from χ² tests of enrichment; scale bar for residuals (−2 to 2) is shown. Methylation-derived metagene levels (V1–V6), which define subgroup membership, are also shown (red indicates high metagene levels, blue indicates low levels). (B) Overall survival of patients in the seven identified subgroups. All discovery cohort patients with available overall survival information are shown (n=367). (C) Progression-free survival of patients in the consensus clustering; certain samples could not be confidently classified for the seven subgroup model or the four subgroup model, and were omitted from the figures. DN/MBEN=desmoplastic or nodular medulloblastoma with extensive nodularity. HR=hazard ratio. LCA=large-cell anaplastic. M+=metastatic disease. R+=residual disease.

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www.thelancet.com/oncology Published Online May 22, 2017 http://dx.doi.org/10.1016/S1470-2045(17)30243-7
The age distributions of patients in the four MBGrp3 and MBGrp4 subgroups differed significantly \((p<0.0001)\). Patients in the MB Grp3-LR and MB Grp4-LR subgroups were younger at diagnosis than those in the MB Grp3-HR and MB Grp4-HR subgroups (appendix pp 14–15). Infants in the MB Grp3-HR subgroup frequently had amplified \(MYC\).

**Figure 2: MBSHH disease comprises two age-dependent molecular subgroups**

(A) Log-normal age distributions of MB Shh-Infant (red) and MB Shh-Child (dark red). Patient ages at diagnosis are shown as ticks along the x-axis and are coloured by subgroup. (B) Clinicopathological and molecular disease features of MB Shh-Infant and MB Shh-Child subgroups. Residuals from \(\chi^2\) tests indicate where subgroup-enrichment has occurred (darker shades of grey indicate stronger relationships); scale bar for residuals (–4 to 4) is shown. \(p\) values from \(\chi^2\) tests are shown. Differentially methylated probes: Illumina probe identifiers for the top 20 most differentially methylated probes, alongside methylation status of 18 normal cerebella (pink). Each column represents one patient. (C) SHH genome-sequencing data \(^\text{26}\) was classified into methylation subgroups on the basis of age. Each column represents one patient. Amp=amplification. DN/MBEN=desmoplastic or nodular medulloblastoma with extensive nodularity. LCA=large-cell anaplastic. M+=metastatic disease. R+=residual disease.
Figure 3: Characterisation of MB\textsubscript{grp3} and MB\textsubscript{grp4} subgroups

(A) Clinicopathological and molecular disease features. Residuals from \chi\textsuperscript{2} tests indicate where subgroup-enrichment has occurred (darker shades of grey indicate stronger relationships); scale bar for residuals (–6 to 6) is shown. \(p\) values from \chi\textsuperscript{2} tests are shown. (B) Heat map shows the top 20 differentially methylated probes for these subgroups. Methylation data of 18 normal cerebella are shown alongside and magnitude of MB\textsubscript{grp3} and MB\textsubscript{grp4} metagenes is shown below. (C) Identification of MB\textsubscript{grp3} and MB\textsubscript{grp4} medulloblastoma cytogenetic determinants. Markers with \(p<0.05\) and present in at least 10% of one subgroup are ordered by their subgroup association and then by chromosomal order. Residuals from \chi\textsuperscript{2} tests indicate where subgroup enrichment has occurred (darker shades of grey indicate stronger relationships), across all subgroups and within MB\textsubscript{grp3} and MB\textsubscript{grp4} individually. \(p\) values from \chi\textsuperscript{2} tests are shown. \textit{i}17q=ischromosome 17q. LCA=large-cell anaplastic. \(M+=\)metastatic disease. \(R+=\)residual disease.
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[64% of 11 infants]. MB<sub>Grp1</sub> tumours were strongly associated with LCA pathology (20 [35%] of 57) and GFI1 mutations (nine [29%] of 31; figure 3A, appendix pp 14–15). i17q is the sole significantly enriched feature of MB<sub>Grp1</sub> (60 [76%] of 79 [figures 3A, 3C]). Clinico-pathological and molecular disease features of the MB<sub>up</sub> and MB<sub>sub</sub> subgroups are shown in figure 3A.

Several hundred differentially methylated CpG probes or regions defined the four subgroups. MB<sub>Grp1</sub> was characterised by the greatest number of significantly differentially methylated CpGs compared with other subgroups, commonly hypomethylated CpG loci (figure 3B; appendix pp 14–15). Notably, the low-risk subgroups were defined primarily by hypermethylation with respect to normal cerebellum, whereas the high-risk subgroups were defined by hypomethylation (figure 3B; appendix pp 14–15). Cytogenetic changes distinguished each subgroup as unique from the others (figure 3C). These distinguishing cytogenetic features were validated in an independent cohort (appendix pp 14–15).

We did survival analyses in an MB<sub>GrpCD</sub> cohort that included 31 additional SHH cases unsuitable for 450k array analysis and classified as MB<sub>GrpCD</sub> on the basis of age (appendix pp 4–5). In this cohort, one out of three assessable TP53 mutations were germline (appendix pp 16–17). TP53 mutation was significantly associated with MYCN amplification (p=0.022) and LCA pathology (p=0.0033). MYCN amplification was associated with LCA pathology (p<0.0001), and LCA pathology and MYCN amplification were never observed with TERT mutation (p=0.0007 for LCA and p=0.0009 for MYCN amplification). There was no significant association between metastatic (M+) disease and TP53 mutation (p=0.15), or LCA pathology (p=0.67), or an association between subtotally resected (R+) disease and TP53 mutation (p=1), MYCN amplification (p=0.15), or LCA pathology (p=0.013), and between all craniospinally irradiated patients in the MB<sub>Grp4</sub> subgroup aged 3–16 years who had outcome data (n=175), allocation to the MB<sub>Grp1</sub> and MB<sub>Grp4</sub> subgroups was a significant high-risk factor for shorter progression-free survival in univariate analysis (table 3). Additionally, in multivariate analysis, MYC amplification was identified as an independently prognostic high-risk factor, and chromosome 13 loss was associated with an improved outcome (table 3).

### Table 2: Identification of prognostic survival markers in MB<sub>GrpCD</sub> cohort

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<th>n</th>
<th>Univariate (n=55)</th>
<th>Cross-validated multivariate (n=42)</th>
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<td>LCA pathology vs non-LCA pathology</td>
<td>52</td>
<td>2.88 (1.55–7.24)</td>
</tr>
<tr>
<td>TERT wild-type vs TERT mutation</td>
<td>52</td>
<td>2.21 (0.78–6.65)</td>
</tr>
<tr>
<td>R+ vs R– disease</td>
<td>55</td>
<td>3.45 (1.30–9.19)</td>
</tr>
<tr>
<td>Male vs female</td>
<td>55</td>
<td>1.13 (0.45–2.82)</td>
</tr>
</tbody>
</table>

p values are from Cox proportional hazards analyses. The prognostic significance of covariates selected in cross-validated multivariate models are also shown. HR=hazard ratio. M+=metastatic disease. M–=non-metastatic disease. LCA=large-cell anaplastic. R+=residual disease (subtotal surgical resection). R–=no residual disease (gross total resection).

### Table 3: Identification of prognostic survival markers in MB<sub>up</sub> and MB<sub>sub</sub> cohorts

<table>
<thead>
<tr>
<th>n</th>
<th>Univariate (n=175)</th>
<th>Cross-validated multivariate (n=133)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HR (95% CI)</td>
<td>p value</td>
<td>HR (95% CI)</td>
</tr>
<tr>
<td>High-risk methylation group vs low-risk methylation group</td>
<td>175</td>
<td>3.73 (1.94–7.18)</td>
</tr>
<tr>
<td>MYC amplification vs no amplification</td>
<td>173</td>
<td>2.94 (1.06–8.13)</td>
</tr>
<tr>
<td>Loss of chromosome 13 vs no loss</td>
<td>158</td>
<td>0.10 (0.01–0.74)</td>
</tr>
<tr>
<td>MB&lt;sub&gt;up&lt;/sub&gt; vs MB&lt;sub&gt;sub&lt;/sub&gt;</td>
<td>175</td>
<td>2.04 (1.23–3.40)</td>
</tr>
<tr>
<td>M+ vs M– disease</td>
<td>173</td>
<td>1.77 (1.03–3.05)</td>
</tr>
<tr>
<td>i17q vs no i17q</td>
<td>158</td>
<td>1.71 (0.99–2.95)</td>
</tr>
<tr>
<td>Male vs female</td>
<td>175</td>
<td>1.56 (0.86–2.84)</td>
</tr>
<tr>
<td>MYCN amplification vs no amplification</td>
<td>173</td>
<td>0.72 (0.23–2.29)</td>
</tr>
<tr>
<td>LCA pathology vs non-LCA pathology</td>
<td>157</td>
<td>1.08 (0.49–2.39)</td>
</tr>
<tr>
<td>R+ vs R– disease</td>
<td>171</td>
<td>1.22 (0.72–2.09)</td>
</tr>
</tbody>
</table>

Identification of prognostic survival markers in combined childhood non-MB<sub>up</sub> and non-MB<sub>sub</sub> survival cohort (aged 3–16 years, receiving cranio-spinal irradiation, with survival information). p values from Cox proportional hazards analyses are shown. The characteristics of covariates selected in cross-validated multivariate models are also shown. The high-risk methylomic group comprised samples from both MB<sub>Grp3</sub> and MB<sub>Grp4</sub> defined by the shared MB<sub>Grp3</sub>–MB<sub>Grp4</sub> metagene V1. HR=hazard ratio. MB=medulloblastoma. Grp3=group 3. Grp4=group 4. M+=metastatic disease. M–=non-metastatic disease. i17q=isochromosome 17q. LCA=large-cell anaplastic. R+=residual disease (subtotal surgical resection). R–=no residual disease (gross total resection).
A stratification model was developed that divided MBGrp3 into different risk groups for 5-year progression-free survival: favourable risk (chromosome 13 loss and no MYC amplification; 16 [10%] of 153 patients; 92% [95% CI 79–100]); standard risk (MBGrp3-LR or MBGrp4-LR, with no MYC amplification; 50 [33%] patients; 81% [70–94]); high risk (MBGrp3-HR or MBGrp4-HR, with no MYC amplification; 82 [54%] patients; 42% [31–56]); and very high risk (MBGrp4 with MYC amplification; five [3%] patients; 0%; figure 4A; appendix pp 20–21). 156 patients had information for chromosome 13 loss and MYC amplification, of which three were classed as unassignable because they were MBGrp3 with MYC amplification (appendix pp 20–21). This stratification scheme outperformed current risk-stratification models (figure 4B).

For comparison, we developed equivalent separate survival stratification schemes for MBGrp3 and MBGrp4 (appendix pp 22–23). Risk factors identified were broadly consistent with the factors identified in the combined scheme, although the combined scheme was a better predictor of progression-free survival than when MBGrp3 and MBGrp4 were considered separately (figure 4B). Taking MBGrp3 patients in isolation, in univariate analysis, a designation of MBGrp3-LR: chromosome 7q status, M+ disease, and male sex were associated with poor progression-free survival, whereas MYCN amplification, R+ disease, and LCA pathology were not (appendix pp 22–23). Chromosome 7q gain and M+ disease were retained as independent prognostic factors in multivariate analysis (appendix pp 22–23). A 5-year progression-free survival model incorporating chromosome 7q gain and M+ disease defined standard-risk (35 [32%] of 110 patients; 87% [95% CI 76–100]) and high-risk groups (75 [68%]; 49% [37–66]), and outperformed other published models by AUC analysis (appendix pp 22–23).

Taking patients with MBGrp3 in isolation, MYC amplification was the only risk factor significantly associated with progression-free survival in multivariate analysis, and outcomes were poor for these very high-risk patients (appendix pp 22–23). Patients in the MBGrp3 with non-MYC amplified tumours were at high risk, with progression-free survival similar to that for the MBGrp4-HR subgroup (51 [91%] of 56 patients; 46% [95% CI 33–64] for MBGrp3 with non-MYC amplified tumours vs 41% [28–60] for MBGrp4-HR). MBGrp4-LR shows a worse outcome than MBGrp4-HR (p=0.040; appendix p 22). LCA pathology (11 [21%] of 53 patients), M+ disease (17 [29%] of 58 patients), and R+ disease (13 [22%] of 59 patients), were frequent in patients in MBGrp3 but none were associated with prognosis, and no stratification scheme based on MBGrp3 alone markedly improved outcome prediction compared with standard stratification schemes (appendix pp 22–23).
The clinicopathological and molecular features of the new seven clinically significant subgroups are summarised in figure 5. The combination of subgroup-specific survival models creates an overarching risk stratification for all childhood medulloblastoma (figure 6A). Patients are stratified into four clinical risk groups for 5-year progression-free survival: favourable risk (comprising MB\textsubscript{WT}, MB\textsubscript{IHC\textsubscript{Child}} with no high-risk features, and non-MYC amplified MB\textsubscript{Gp3\textsubscript{LR}} with chromosome 13 loss; 54 [25%] of 218 patients; 91% [95% CI 82–100]); standard risk (comprising non-MYC features, and non-MYC amplified MB\textsubscript{Gp3\textsubscript{LR}} subgroups; 50 [23%] patients; 90% [95% CI 80–100]); high-risk (comprising non-MYC amplified MB\textsubscript{Gp3\textsubscript{LR}}/MB\textsubscript{Gp4\textsubscript{HR}} subgroups; 82 [38%] patients; 72% [95% CI 60–84]); and metastatic to be classified as favourable. The discovery and validation of seven robust and reproducible primary molecular subgroups of childhood medulloblastoma in this retrospective cohort study represents, to our knowledge, the first clinically significant elaboration of the four-subgroup consensus classification scheme.

**Discussion**

The discovery and validation of seven robust and reproducible primary molecular subgroups of childhood medulloblastoma outperforms current and proposed cytogenetic risk stratifications (figure 6C). Incorporation of M+ disease status into MB\textsubscript{Gp4\textsubscript{LR}} and non-MYC amplified MB\textsubscript{Gp3\textsubscript{LR}} survival modelling does not affect model performance, but potentially allows redistribution of standard-risk patients to create larger favourable (90 [41%] of 218 patients) and high-risk groups (99 [45%] of 218 patients; figure 6A, C; appendix pp 24–25), which could be considered as an alternative stratification scheme. The proposed refinement to the stratification enables additional cases classified as MB\textsubscript{Gp4\textsubscript{LR}} and MB\textsubscript{Gp3\textsubscript{LR}} that do not have copy number information (other than MYC amplification status) and are non-metastatic to be classified as favourable.
established in 2012. While our work supports the stability of the four established groups, it also reveals significant substructures within each group with distinct clinicopathological and molecular features. Importantly, these primary subgroups emerge from unsupervised analysis, and are supported by distinguishing DNA methylation, gene expression, and copy-number profiles, consistent in discovery and validation cohorts. Notably, these subgroups were not identifiable in a previously published dataset, which included fewer samples and, specifically, fewer infant patients. Our seven subgroups reveal a biological overlap between MB Grp3 and MB Grp4. They share a biological signature, defined by a common metagene, indicating a clinicobiological overlap, which might suggest a common origin.

These primary subgroups may be further subdivided by the presence or absence of secondary molecular characteristics, many of which, in turn, have subgroup-specific clinical and prognostic significance (e.g., MYC amplification and non-MBCSHH or non-MBWNT, etc.).
amplification in MB$_{\text{Grp4}}$ or $TP53$ mutation, MYCN amplification, LCA pathology, M+ disease, and R+ disease in MB$_{\text{Grp1-LR}}$. Some of these secondary features have been described and assigned clinical significance in previous studies; in this Article, their association with specific novel subgroups (eg, chromosome 11 loss and chromosome 17 gain in MB$_{\text{Grp4}}$) has revealed the underlying biological basis of these subgroup-specific biomarkers. Moreover, re-evaluation of currently used high-risk factors derived from cohort-wide studies that did not consider subgroup shows that their importance is either low (eg, LCA pathology, M+ disease, or R+ disease in MB$_{\text{Grp1}}$; MYCN in MB$_{\text{Grp4}}$) or high (MYCN amplification, LCA pathology, $TP53$ mutation, and M+ disease in MB$_{\text{Grp4-LR}}$ MYC in MB$_{\text{Grp4}}$; M+ in MB$_{\text{Grp4}}$) when considered in the context of our new subgroups. Finally, the biological definition of MB$_{\text{Grp4-LR}}$ patients older than 3 years is at odds with current clinical definitions of infant disease (<3 years) and this should prompt consideration in the future as to whether infant treatment protocols are appropriate for MB$_{\text{Grp4-LR}}$ patients older than 3 years. Survival modelling in children younger than 3 years is qualitatively different from analysis in those over 3 years of age, because of the heterogeneity of treatment of infant disease. As such, we regarded further risk modelling in this patient group to be outside the scope of this study, to be addressed in future investigations. To our knowledge, no previous study has directly assessed survival of the molecularly-defined MB$_{\text{Grp4-LR}}$ subgroup. The overall survival at 5 years that we observed in MB$_{\text{Grp4-LR}}$ disease (62%, 95% CI 50–77) is lower than previously reported in an international meta-analysis of the MB$_{\text{Grp4}}$ subgroup in age-defined infants (<4 years at diagnosis; 77%), but these patients were not molecularly defined and, as such, are not directly comparable.

Our survival analysis focused on the 3–16-year-old clinical group who received conventional therapies: surgical resection followed by adjuvant radiotherapy with or without chemotherapy at diagnosis with curative intent. Combined risk-modelling across all patients in the non-MB$_{\text{ext}}$ or non-MB$_{\text{Grp4}}$ subgroups identified MYC amplification, high-risk methylation subgroup membership, and loss of chromosome 13 as independent risk factors. Survival models incorporating these factors outperformed the clinical risk-stratification used in current clinical trials (HIT-SIOP-PNET5-MB$^5$) and subgroup-dependent cyogenetic stratification schemes.

We have defined a risk-stratification of childhood medulloblastoma that allows patients to be assigned into four overarching risk groups. Favourable-risk patients, including both MB$_{\text{ext}}$ and novel non-MB$_{\text{ext}}$ groups, should be urgently considered for therapy-reducing strategies. Very high-risk patients, typically refractory to conventional therapies (eg, amplified MYCN, mutated $TP53$, LCA pathology, and M+ disease in MB$_{\text{Grp4}}$ and amplified MYC in MB$_{\text{Grp4}}$) should be prioritised for alternative upfront treatment strategies. The priority for high-risk patients, comprising the novel MB$_{\text{Grp4-LR}}$ and patients with non-amplified MYC in the MB$_{\text{Grp1-LR}}$ subgroup, and a standard-risk group, comprising all other patients, should be optimisation of current therapies and the application of novel, biologically targeted agents.

We note the limitations of developing survival models in retrospective patient cohorts, who received heterogeneous treatments. Notwithstanding that models were developed using patients aged 3–16 years, who all received maximal surgical resection and craniospinal irradiation with curative intent, caution should be applied to their clinical implementation. We also note the statistical limitations of stratifications identifying small numbers of patients (eg, very high-risk, 13% of patients). Moreover, some of the identified biomarkers (notably loss of chromosome 13) have not previously been reported as prognostic. We therefore emphasise that validation in additional cohorts, and ideally in prospective, uniformly treated patients in clinical trials, is essential. A small number of samples (<5 samples) from this study were used to assist with the creation of the four-subgroup classification consensus. Similarly, our own publication that described four methylation-dependent subgroups of medulloblastoma contained 87 samples that overlapped with this study, although the previously published study contained fewer samples (discovery cohort size of 100 and validation cohort size of 130 patients) and DNA methylation profiling was at much lower resolution (1505 vs >400 000 CpG loci).

The existence of novel primary medulloblastoma subgroups, coupled with the characterisation of secondary prognostic features within each group, represents a significant advance in our understanding of medulloblastoma biology and its application in clinical management and future trials design. We provide clear evidence of the shared biology between MB$_{\text{Grp3}}$ and MB$_{\text{Grp4}}$ which affects clinical behaviour and has significant implications for understanding disease biology, developmental origins, and experimental modelling. These investigations constitute a blueprint for a new consensus in medulloblastoma molecular subclassification with important implications for future molecular diagnostics and clinical management.

**Contributors**

ECS, DW, SB, and SCC designed the study and wrote the manuscript. JCL, SC, AJ, and RMH did laboratory experimentation and analysis. ECS, DW, TS, and SN did bioinformatics analysis. SN, DH, GR, and AI prepared figures. TSJ, AJ, and SBW provided central pathology review. BP, AM, AJ, SBW, TSJ, and SB gathered samples and patient data and provided clinical interpretation. All authors contributed to and approved the final manuscript.

**Declaration of interests**

We declare no competing interests.
Acknowledgments
This study was funded by Cancer Research UK (C8464/A13457), The Tom Grahame Trust, Star for Harris, Action Medical Research, SPARKS, The JGW Patterson Foundation, and The INSTINCT network (co-funded by The Brain Tumour Charity, Great Ormond Street Children’s Charity, and Children with Cancer UK). TSJ is supported by the National Institute for Health Research and a Great Ormond Street Hospital UCL Biomedical Research Centre award.

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