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1	ARE EXPOSURE PREDICTIONS, USED FOR THE PRIORITISATION OF
2	PHARMACEUTICALS IN THE ENVIRONMENT, FIT FOR PURPOSE?
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22 Abstract: Prioritisation methodologies are often used for identifying those pharmaceuticals 23 that pose the greatest risk to the natural environment and to focus laboratory testing or environmental monitoring towards pharmaceuticals of greatest concern. Risk-based 24 25 prioritisation approaches, employing models to derive exposure concentrations, are 26 commonly used but the reliability of these models is unclear. The present study evaluated 27 the accuracy of exposure models commonly used for pharmaceutical prioritisation. Targeted 28 monitoring was conducted for 95 pharmaceuticals in the Rivers Foss and Ouse in the City of 29 York, UK. Predicted environmental concentration (PEC) ranges were estimated based on localised prescription, hydrological data, reported metabolism and wastewater treatment 30 31 plant (WwTP) removal rates, and were compared to measured environmental 32 concentrations (MECs). For the River Foss, PECs, obtained using highest metabolism and 33 lowest WwTP removal, were similar to MECs. In contrast, this trend was not observed for 34 the River Ouse, possibly due to pharmaceutical inputs beyond our modelling. 35 Pharmaceuticals were ranked by risk based on either MECs or PECs. With two exceptions 36 (dextromethorphan and diphenhydramine), risk ranking based on both MECs and PECs 37 produced similar results in the River Foss. Overall, these findings indicate that PECs may well 38 be appropriate for prioritisation of pharmaceuticals in the environment when robust and 39 local data on the system of interest are available and reflective of most source inputs to the 40 system.

Keywords: Pharmaceuticals, Prioritisation, Risk ranking, Exposure, Hazard/risk assessment

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INTRODUCTION

47 There is increasing concern over the presence and potential effects of pharmaceuticals in 48 the natural environment. The ubiquitous presence of pharmaceuticals in aquatic systems is 49 well-established [1,2]. Pharmaceuticals are designed to induce a biological response at 50 nanomolar concentrations, raising questions regarding the risk for unintended sub-lethal 51 chronic effects in exposed non-target organisms [3]. Of the approximately 1500 52 pharmaceuticals currently in use in the UK alone, acute ecotoxicity data are available for 53 only a small proportion of these and chronic data are even more scarce [4]. Additionally, 54 little is known about the environmental fate of most pharmaceuticals [5]. Few have undergone extensive fate testing such as quantifying half-lives in environmental matrices, 55 56 partitioning to sludge, soils, or sediment and uptake into terrestrial and aquatic organisms. 57 Therefore substantial knowledge gaps exist that need to be filled before we can fully 58 understand the effects of pharmaceuticals in the natural environment. To fill these gaps 59 experimentally, however, would require substantial effort in terms of time and cost.

60 Prioritisation methodologies provide a useful tool for identifying which of the thousands 61 of pharmaceuticals in use have the greatest potential to cause unintended effects in non-62 target organisms and which therefore should be experimentally tested in terms of their fate 63 and effects [6]. Several prioritisation approaches have been proposed for pharmaceuticals. 64 For example, hazard-based approaches have involved the prediction of persistence, 65 bioaccumulation, and toxicity of a pharmaceutical and these have then been used to 66 develop an overall hazard score. Compounds with the highest scores are considered to have 67 the highest priority [7]. Risk-based approaches have involved the estimation or 68 measurement of pharmaceutical concentrations in environmental media and the

comparison of these concentrations with an effect endpoint, for example predicted noeffect concentrations derived from acute or chronic ecotoxicity data [8–10] or predictions [11], plasma therapeutic concentrations [12], acceptable daily intakes for humans [13] or a combination of these [4]. Risk-based methods have been identified as preferable due to the consideration of effects and environmental occurrence, ruling out the possibility of prioritising compounds that have little chance of accumulating in the environment at ecologically relevant concentrations [6,13].

76 All risk-based approaches require an assessment of the concentration of pharmaceuticals 77 in the environment. Real environmental data are desirable, however, monitoring data are 78 generally lacking for a wide range of pharmaceuticals. Moreover, when monitoring data are 79 available, the relevance of the data is often questionable due to sampling designs that do 80 not consider seasonal biases, hydrologic conditions or spatiotemporal fluctuations [14]. As a 81 result, comparing absolute measured concentrations of pharmaceuticals for prioritisation 82 has been questioned [15]. Furthermore, sufficiently sensitive analytical methods, suitable 83 for complex environmental matrices, or isotopically labelled standards necessary for 84 accurate quantitation are not yet available for the majority of pharmaceuticals in use, 85 making determination of pharmaceuticals in environmental matrices challenging [9,11].

Consequently, many risk-based prioritisation methods have employed exposure prediction models or algorithms to derive predicted environmental concentrations (PECs) in order to prioritise pharmaceuticals that have no monitoring data and/or to provide conservative estimates of environmental concentrations [16]. PECs are typically derived based on data on pharmaceutical usage, degree of metabolism in humans, removal in wastewater treatment plants (WwTP) and environmental dilution. The method most commonly used is based on the approach recommended in the European Medicines Agency
(EMEA) guidelines for assessment of the risk of human pharmaceuticals in the environment
[6,9,11,17–23]. Default parameters (e.g. for dilution of wastewater) proposed by the EMEA
guidance are regularly used in these prioritisation exercises, regardless of their suitability
[6,10,19,20]. The use of site-specific data when performing these calculations for
prioritisation is a rarity [21].

98 The impact of using PECs for prioritisation has not been explored, although several 99 authors have explored how well PECs compare to measured environmental concentrations 100 (MECs) [1,2,16,24–29]. These comparisons have provided varied results, with some studies 101 showing that PECs adequately represent MECs [24–28], while others suggest the differences 102 are too great to be useful, or that PECs generally under represent MECs [1,16,29]; in 103 addition, these comparative studies concentrate on pharmaceuticals that have been 104 identified as being of concern, or of high usage and generally focus on fewer than 10 105 compounds [28], limiting the relevance of their conclusions across the broader spectrum of 106 physico-chemically diverse pharmaceuticals known to be present in the environment 107 globally.

Usually the determination of PEC relevancy is reliant on determining a PEC/MEC ratio. The acceptability of the PEC depends on how close this ratio is to 1 [29], however the acceptable range varies between studies [28]. This poses a problem when trying to assess the relevance of results across studies because the derivation of these ranges is subjective and dependent on the motive of the study (e.g. prioritisation or risk assessment).

113 In the present study, we evaluate PEC models for use in prioritisation by comparing 114 modelled and monitoring data from a comprehensive set of 95 pharmaceuticals derived 115 from a wide range of therapeutic classes with different modes of action, an extensive range 116 of chemical and physical properties, high and low usage, as well as select pharmaceuticals 117 not thought to be prescribed in the UK. The city of York (population of 227 000) was chosen 118 as the study system due to the availability of local prescription data, a well-defined and 119 accessible hydrological system (i.e. two rivers that pass through the city), and numerous 120 access points to the rivers via bridges, which enables a detailed characterisation of 121 pharmaceutical concentrations throughout the city. The prioritisation approach used to 122 compare PECs and MECs was based on the Fish Plasma Model (FPM) [12]. Studies of this nature that assess a large range of compounds (95), are an important check on ensuring 123 124 that priority compounds identified, using common modelling approaches, are comparable 125 to those using environmental data representative of key seasonal, locational, water 126 treatment and hydrological differences.

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METHODS

128 Study site and sampling

129 We collected and analysed river water samples from eight sites along the Rivers Ouse 130 and Foss in the City of York in the UK where flow conditions were below the long term mean 131 flow and near the Q50 (i.e where flow is equal or exceeded 50% of the time) in February 132 2015 (Figure 1)[30]. Site locations were chosen based on ease of access and their position in 133 relation to WwTP outfalls discharging into these river systems (Supplemental Data, Table 134 S1). Two WwTPs serve the city of York that impact the sampling network. There is a third 135 WwTP; however, it is downstream of the city and sampling points (not included in Figure 1). 136 The first of these two WwTPs (WwTP A) serves a population of 27 900, employs 137 conventional activated sludge (CAS) as secondary treatment and nitrifying filters as a 138 tertiary treatment option, and the second (WwTP B) serves a population of 18 600 and uses trickling filter technology as secondary treatment paired with biological aerated filtration fortertiary treatment.

141 At each site, three 1-L samples were collected at points distributed equidistant across the 142 width of the river channel and homogenised into a single 1 L composite sample. Three 10-143 mL aliquots were taken from the composite sample and filtered through 0.7 µm glass 144 microfiber (GF/F) disposable filters (Whatman Inc.). To ensure that filtration and field 145 handling of samples did not result in cross-contamination, high-performance liquid 146 chromatography (HPLC)-grade water was also filtered and prepared in the field identically to 147 river samples (i.e. a field blank) three times during the sampling. Samples were frozen 148 directly in the field using dry ice and transported to the U.S. Geological Survey (USGS) National Water Quality Laboratory in Denver Colorado, USA. They arrived four days later 149 150 and were immediately thawed and analysed.

151 Analytical Methods

152 Samples were analysed using a direct injection (100 µL) high-performance liquid 153 chromatography/tandem mass spectrometry with an electrospray ionization source (LC-ESI-154 MS/MS) method for the determination of 110 pharmaceuticals, pharmaceutical degradates, 155 and wastewater indicator compounds [31]. Of the 110 compounds, 95 pharmaceuticals 156 were targeted in the present study with method detections limits (MDL) as defined by the 157 US Environmental Protection Agency (USEPA) [32] down to 0.45 ng/L (Table 1). Instrumentation included an Agilent 6410 triple quadrupole MS/MS system coupled with an 158 159 Agilent 1200 Series HPLC. Mobile phases were HPLC-grade water modified with 1M formic 160 acid and 1M ammonium formate (A) and 100% HPLC grade methanol (B). Chromatography 161 gradient and conditions are detailed in Supplemental Data, Table S2. Quantification and

identification was achieved by external calibration with known standards for each of the 162 163 pharmaceuticals and completed using Agilent Mass Hunter software in accordance with the 164 USGS methodology described in Furlong et al. [31]. The MS/MS was operated in multiple 165 reaction monitoring (MRM) mode, where two MRM transitions and correct retention times 166 were required for ion qualification, while quantification was based on the major transition 167 (Supplemental Data, Table S3). Additionally, ion ratios between the major and secondary 168 transitions were required to fall within a compound-specific range determined from the 169 corresponding analytical standard [31]. Concentrations reported in the present study are the median of three aliquots taken from each site. 170

Statistical analysis and quality control. The limit of quantification (LOQ) was established as 2 to 5 times the MDL where the probability of incorrectly reporting the presence of an analyte is less than 1% when concentrations are equal to or greater than the LOQ [33]. Concentrations greater than the LOQ were fully quantitative while concentrations detected between the LOQ and MDL were considered semi-quantitative estimates. To enable the consideration of as many pharmaceuticals as possible, both quantitative and semiquantitative data were used in subsequent data analyses.

Quality control samples were analysed to (1) assess matrix recovery efficiency and identify the presence of matrix interferences that could induce ion suppression or enhancement [34], and (2) identify any blank contamination from sampling and analysis. For recovery assessment, an environmental sample was amended with the pharmaceuticals of interest (matrix spike) to a concentration of 400 ng/L. The aforementioned field blank samples were analysed to identify any potential contributions of pharmaceuticals during sample collection, laboratory processing and analysis. In addition to the field blank and 185 matrix spike samples, analogous laboratory spike and blank samples, using high purity HPLC-

186 grade water, also were analysed with each batch of environmental samples.

187

188 PEC Modelling

189 The calculation of PECs for the 95 pharmaceuticals was based on Equation 1.

190 PEC=
$$\frac{\text{consumption * F}_{\text{excreta}} * (1-WwTP removal)}{\text{inhabitants * WW}_{\text{inhab}} * dilution}$$
 (1)

191 Where the numerator represents the river input rate (ng per day): consumption = 192 amount used per day (ng/day); $F_{excreta}$ is the fraction of pharmaceutical excreted unchanged 193 by patients; and WwTP removal is the fraction of a pharmaceutical removed by water 194 treatment. The denominator is the river flushing rate where: inhabitants = population 195 served by the WwTP; WW_{inhab} = amount of wastewater generated (L/day·person), which has 196 a default value of 200; dilution was based on site-specific conditions in each river.

197 Pharmaceutical usage was generated from localised prescription data released monthly 198 by the National Health Service for January 2015 [35]. Relevant medical practices were 199 selected by postal code (Supplemental Data, Table S4). The F_{excreta} term was obtained from 200 either the peer-reviewed literature or online databases such as Drugbank, MedSafe and 201 RXmed, as well as publicly available pharmaceutical data sheets released by government 202 organisations such as MedSafe New Zealand or the Food and Drug Agency (Supplemental 203 Data, Table S5). When a pharmaceutical was metabolised to conjugated metabolites (e.g. 204 glucuronide or sulfato-conjugates), the portion released as a conjugate was added to the 205 unchanged parent excretion estimate. These metabolites can undergo reactions during 206 water treatment such as cleavage and thus be converted back into their parent compounds,

increasing the parent pharmaceutical load in wastewater effluent [36]. Estimates of
unchanged pharmaceutical excretion varied across sources; this led to a range of possible
unchanged excretion estimates, which were used to calculate a PEC range. For ophthalmic
and topical preparations, metabolism was assumed to be zero and therefore the F_{excreta} was
set to 1 [19].

Wastewater treatment removal was considered in two ways due to the limited availability of removal estimates for all pharmaceuticals in the present study [37]. Firstly, removal values from the literature were collected and, similarly to F_{excreta} estimates, varied substantially (Supplemental Data, Table S5). The range of possible WwTP removal estimates were used to calculate a possible PEC range. Secondly, data gaps were filled using the USEPA's EPISuite software STPWIN program [38], similarly to a recent prioritisation exercise in Asia [20].

219 Evaluation of PECs

Separate PEC ranges were calculated for pharmaceuticals for both the River Foss and River Ouse. The PEC range incorporated a river-specific dilution factor reflecting hydrological conditions on the day of sampling. The lowest $F_{excreta}$ and highest WwTP removal values found in the literature were paired to give a minimum PEC, while the maximum was derived using the highest $F_{excreta}$ and lowest WwTP removal found in the literature. A PEC (worst case) was also calculated which only considered site-specific dilution (ie. $F_{excreta} = 1$, WwTP removal = 0).

227 Prioritisation Approach

The fish plasma model (FPM) approach [12,39], which has been used in previous prioritisation exercises [6], was selected as the method used for prioritisation. Bioconcentration factors (BCFs) for neutral and ionisable compounds were estimated according to the approach of Fu et al. [40] (Supplemental Data, Equations S1-S5) and used to determine fish plasma concentrations (FPCs) based on either PECs or MECs . FPCs were then compared to human plasma therapeutic concentrations (indicated by C_{max}) using Equation 2 to determine the risk quotient (RQ). The K_{ow} and C_{max} for all compounds were collected from the MaPPFAST database complied by Berninger et al. [41].

$$RQ = \frac{PEC^*BCF}{C_{max}}$$
(2)

RQs are ranked from highest to lowest risk, where a larger RQ indicates a greater potential risk. Using this approach, we obtained two ranking lists, one based on FPCs obtained from PECs, the other using FPCs obtained from MECs.

240

RESULTS AND DISCUSSION

241 Pharmaceutical Occurrence

242 No pharmaceuticals were detected in the field blanks collected indicating that sample 243 collection, handling, and analysis did not result in measurable contamination of the water 244 samples (i.e. protocols did not generate false positives for the present study). Calculated 245 recoveries from quality control matrix spike samples generally fell within 60-120% and were 246 considered acceptable [42]. Recoveries failing to meet these criteria are identified and 247 subsequently interpreted with caution. Reported values were not corrected for percentage 248 of analyte recovered in environmental matrix spikes [43]. The median matrix recovery was 249 88% while the 25 and 75 percentiles were 81 and 160% respectively; this distribution 250 suggests that some matrix enhancement of compound recoveries is occurring.

251 Of the 95 pharmaceuticals surveyed, 25 compounds were detected and quantified 252 (Figure 2) in the eight water samples collected from the York network. A further 19 253 pharmaceuticals were detected, however only qualitative or semi-quantitative assessment 254 was appropriate due to either quantification limits (11) or unacceptable matrix 255 interferences (7) (Table 1). Of the 25 pharmaceuticals quantified, 10 have not been 256 previously identified in the UK aquatic environment to the authors' knowledge: acyclovir, 257 diphenhydramine, glyburide, hydrocodone, lidocaine, methocarbamol, oseltamivir, 258 sitagliptin, triamterene and loratadine. The remaining 15 pharmaceuticals detected were 259 consistent with the ranges reported previously in the literature (Table 1). Ten pharmaceuticals included in the analysis are not prescribed in the UK and were not detected 260 261 in any samples. Median and maximum detected concentrations, along with detection 262 frequency and matrix recoveries for all target analytes are reported in Table 1.

The concentrations and number of detections between the Rivers Ouse and Foss varied (Fig. 2) with concentrations of six pharmaceuticals in the River Foss being significantly higher than in the Ouse (Student's T-test, p < 0.05). A greater number of and more consistent detections occurred in the River Foss, (Fig. 2) which has both a lower dilution factor and the corresponding WwTP (WwTP B) provides less sophisticated water treatment (trickling filter) compared to the treatment used by WwTP A discharging to the River Ouse (conventional activated sludge).

270 Evaluation of Modelled Concentrations with Monitoring Data

The EMEA PEC model describes an annual average concentration for the region the consumption data cover; in general, usage data from the whole of a country is averaged to give a single PEC [4]. Evaluating this approach with localised, temporally limited samples

would introduce a source of potential error as it has been shown that seasonal usage is 274 275 important for some pharmaceuticals and that demographics in a specific area may differ 276 substantially from the national average [25,26]. To reduce these potential biases, local 277 usage data, corresponding to time of sampling, was used. In addition, site-specific dilution 278 factors were incorporated to avoid the use of EMEA [23] default dilution factors (i.e. 10). 279 The WW_{inhab} term could not be refined to actual discharge because both WwTPs are highly 280 variable and discharge measurements were not available for the sampling dates. This 281 permits a focus on other factors that could be affecting the suitability of PECs such as WwTP 282 removal and metabolism.

283 Overall PEC Performance

284 Many pharmaceuticals targeted were not detected in the monitoring campaign, however 285 based on their PECs, this was not unexpected. To assess the overall performance of the 286 PECs, a semi-quantitative approach was taken. Each of the 77 pharmaceuticals for which a 287 PEC could be calculated were sorted into one of four possible categories (Figure 3). 288 Pharmaceuticals that were expected to be detected in the monitoring campaign (i.e. PEC 289 greater than the corresponding analytical MDL) were sorted into either detected or not 290 detected categories. Similarly, pharmaceuticals not expected to be detected (i.e. PEC less 291 than the respective analytical MDL) were sorted into detected and not detected categories. 292 Overall in the semi-quantitative analysis, the PECs in the two rivers performed well with 79% 293 and 86% of predictions correctly confirmed in the River Foss and Ouse, respectively, by the monitoring data. 294

The large difference in dilution between the two rivers, factors of 17.8 and 540 for the Foss and Ouse respectively, led to larger PECs in the River Foss and therefore a higher 297 number of expected detections. A larger proportion of expected detections were not 298 identified in our monitoring campaign in the Foss in comparison to the Ouse; it could be that 299 pharmaceuticals were missed by our sampling effort, however our results indicate that 300 pharmaceutical concentrations are stable throughout the River Foss over an 8-hour period 301 (Figure 2), which diminishes the likelihood of missing a detection. Conversely, the 302 metabolism or WwTP removal selected from the literature may have produced PECs larger 303 than real-world concentrations. The number of unexpected but detected pharmaceuticals is 304 greater in the River Ouse, despite corrections for upstream contributions detected at site 4, 305 (Figure 2). The River Ouse could be subject to a greater number of sources not reflected in 306 our usage estimate in contrast to the more rural River Foss. Sources of pharmaceuticals 307 beyond the scope of localised prescription data exist within the city include, for example, a 308 substantial tourism industry and two post-secondary institutions. Recent studies have 309 demonstrated the impact of post-secondary institutions [44] and music festivals [45] on 310 MECs, and it is likely that MECs in the Ouse are influenced by demographic factors not 311 inclusive of localised prescription-based usage estimates.

312 Impact of Metabolism and WwTP Removal Uncertainty on PECs

313 Underestimated PECs: A breakdown of how each pharmaceutical PEC performed in 314 comparison to the MEC is shown for the River Foss (Figure 4) and the River Ouse (Figure 5). 315 While the overall semi-quantitative performance of PECs in the River Ouse was slightly 316 better than the Foss, these results were not repeated when quantitative data were 317 compared. In the Foss and the Ouse, 38% and 78% respectively, of the MEC ranges were 318 entirely greater than the corresponding PEC range. This drops to 12% and 44% respectively 319 when the PEC (worst case) is considered. The PEC (worst case) does not include metabolism 320 or WwTP removal, only dilution, and when this PEC still falls below the MEC it indicates a

321 problem with the consumption estimate. The analytical matrix spike recoveries indicated 322 that matrix enhancement is occurring, which could affect the comparisons with PECs. To 323 investigate, each compound with a MEC range greater than the PEC range was theoretically 324 corrected based on the compound specific matrix recovery. All of the theoretically corrected 325 MEC ranges were still greater than the corresponding PEC ranges in the River Ouse and Foss 326 with one exception, erythromycin, where the MEC range corresponded with the top of the 327 PEC range in the River Foss. Therefore we do not expect our results to be significantly 328 altered by the distribution in matrix recoveries.

329 In the River Foss, three pharmaceuticals (dextromethorphan, diphenhydramine and 330 pseudoephedrine) had greater MECs than PEC (worst case) estimates and are all available 331 over-the-counter (OTC). This consumption pathway was not considered in our consumption 332 estimate as we were unable to access data on sales of OTC medicines. As a result, PECs for these pharmaceuticals should be systematically underestimated [2,24,27]. This was not 333 334 reflected for all OTC pharmaceuticals, similarly to a recent study in Canada [28]. This 335 highlights the need for a new approach to incorporate OTC consumption into WwTP 336 pharmaceutical loadings [4,27]. The results from the River Ouse (Figure 5) are more 337 complicated, a mixture of both OTC and prescription-only pharmaceuticals had MECs which 338 were greater than the PEC (worst case) estimates. This supports our semi-quantitative 339 findings where a problem exists with the consumption estimate and is likely a result of the 340 specific demographics impacting pharmaceutical loads for the River Ouse.

341 *PEC ranges:* The PEC range is large for many of the pharmaceuticals. For instance the 342 paracetamol PEC range covers over 4 orders of magnitude (Figure 4). This large uncertainty 343 is a result of the extensive variability in experimental WwTP removal and F_{excreta} estimates

obtained from the literature. In both rivers, the majority of PEC ranges vary by at least 2 344 345 orders of magnitude, which could be important from both a risk assessment and 346 prioritisation perspective. The large PEC range does mean that, in general, the MEC range 347 did correspond with predictions in the River Foss (Figure 4). The MEC range is typically near 348 the top of the PEC range, where the smallest WwTP removal was paired with the highest 349 unchanged excretion found in the literature. This finding has two implications: firstly, 350 choosing the worst-case fate parameters to estimate PECs is likely the best approach to 351 avoid underestimations of PECs, which is in agreement with PEC approaches in the literature 352 [46]; secondly, anything short of an exhaustive literature review could lead to underestimated PECs in the majority of cases shown in Figures 4 & 5. This is because the 353 354 PEC ranges determined herein are the result of an exhaustive literature review; in a larger 355 scale prioritisation exercise the time resources required to thoroughly check each 356 compound would be impractical and the process itself highly subjective. This could lead 357 authors to different conclusions about the resulting risks and priority compounds as it is a 358 single value computed for the PEC, not a range, which is a substantial flaw not often considered when the fate data used in a PEC are collected in this manner. 359

360 Our results indicate that consideration of metabolism and WwTP removal is essential when calculating PECs because PEC (worst case) is a large overestimate of actual 361 362 concentrations in the majority of cases (Figure 4), also shown by others [6,10,22]. In the 363 River Foss, prescription pharmaceuticals are described well using the PEC approach. This is 364 in sharp contrast in the River Ouse, where multiple consumption sources are likely affecting 365 concentrations of the pharmaceuticals in the environment, making it impossible to evaluate the effect of the fate parameters with the current dataset. Further monitoring that 366 367 incorporates sampling WwTP influents and effluents to compute actual removals will be 368 critical to assessing PECs relative to MECs. In addition, the uncertainty in measured 369 concentrations can be limited by incorporating time-averaged composite samples 370 representative of the average conditions [14]. Further work which includes a seasonal 371 monitoring campaign is suggested to quantify the seasonal variability and magnitude of 372 influence that tourism and post-secondary institutions have on MECs in addition to serving 373 as a check of the findings from the present initial scoping study.

- 374
- 375

376 Implications for prioritisation

377 Risk ranking order is important as it dictates which pharmaceuticals are of highest risk 378 and thus, most likely to receive further costly investigations into effects and occurrence [4]. 379 Therefore we evaluated the similarities and differences between risk rankings obtained 380 based on MECs and rankings based on PECs for the River Foss (Figure 6A) and River Ouse 381 (Figure 6B). In the River Foss, while there was some variability in the ranking position of 382 individual compounds, generally, the rankings based on MECs and PECs followed a similar 383 trend. Compounds identified as highest risk based on MECs also were identified as highest 384 risk based on PECs and those ranked as lower risk based on MECs also ranked as lower risk 385 using PECs (Figure 6A). The exceptions were dextromethorphan and diphenhydramine 386 where the rank position was much higher based on MECs than based on PECs. This degree 387 of similarity was not observed in the River Ouse (Figure 6B). Eight of the MEC ranks are higher risk than their PEC rank counterparts, which visually, is a more variable but gentler 388 389 rise (Figure 6B). This indicates that the degree in which PECs were underestimated in the 390 River Ouse affects prioritisation ranking order trends.

CONCLUSIONS

We have presented real-world monitoring data for a comprehensive set of 95 392 393 pharmaceuticals in two rivers that run through the city of York, UK. During a snapshot 394 sampling where flow conditions were below the long-term mean and near the Q50 in 395 February 2015, 25 pharmaceuticals were quantified (i.e. detected), 10 of which had not 396 been previously measured in the UK aquatic environment. Site-specific PEC ranges varied up 397 to four orders of magnitude due to the variability in metabolism and WwTP removal values 398 found in the literature. The largest unchanged excretion paired with the lowest WwTP 399 removal approach provided the greatest comparability to measured concentrations. Some 400 of the observed differences between MECs and PECs might be explained by complex social 401 demographics, such as tourism or post-secondary institutions, which are suspected of 402 influencing wastewater loading estimates. When PECs and MECs were used to prioritise the 403 detected pharmaceuticals based on risk, generally the two approaches provided similar 404 ranking outcomes for well-defined systems such as the River Foss, but were less comparable 405 in the more complicated system, the River Ouse. The findings for the Foss, in particular, provide some confidence in the use of PECs in prioritisation exercises for pharmaceuticals. 406

407

SUPPLEMENTAL DATA

- 408 Table S1 National grid references of sampling site locations
- 409 Tables S2-S3 Analytical operating conditions.
- 410 Tables S4-S5 PEC parameters.

411 Equations S1-S5 Bioconcentration factor equations.

412

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420

421 *Data availability*—Data, associated metadata, and calculation tools are available by 422 contacting the corresponding author (alistair.boxall@york.ac.uk).

423

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Figure 1. Locations of the 8 sampling sites around the city of York, UK. A and B represent the WwTPsthat service the city. Grab samples were collected in February 2015.

Table 1. Occurrence data for the 8 water samples collected during February 2015 from the sampling network with matrix recovery and method detection
 limits for each of the 95 pharmaceuticals, pharmaceutical degradates and wastewater indicators targeted.

Pharmaceutical	Source or use	MDL (ng/L)	Detection Frequency %	Max (ng/L)	Median (ng/L)	Matrix recovery % (median)	Detected in the UK (ng/L)
10-Hydroxy- amitriptyline	Degradate of amitriptyline	1.7	0	ND	ND	110	
Abacavir	Antiviral	4.1	0	ND	ND	73	
Acyclovir ^a	Antiviral	4.4	13	7.9	7.9	60	
Albuterol ^a	β2-adrenergic receptor	1.2	0	ND	ND	180	$38 - 470^{2 e}$
Alprazolam	Benzodiazepine	4.3	0	ND	ND	75	
Amitriptyline	Antidepressant	19	25	<mdl< td=""><td><mdl< td=""><td>250</td><td>1.0 – 72^{f,g}</td></mdl<></td></mdl<>	<mdl< td=""><td>250</td><td>1.0 – 72^{f,g}</td></mdl<>	250	1.0 – 72 ^{f,g}
Amphetamine	Psychostimulant	4.1	0	ND	ND	76	1.1 -4 ^f
Antipyrine ^b	Analgesic	58	20	<mdl< td=""><td><mdl< td=""><td>87</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>87</td><td></td></mdl<>	87	
Atenolol	Beta blocker	2.7	13	25	25	97	<1 – 530 ^e
Benztropine ^{b,c}	Anticholinergic	7.9	0	ND	ND	300	
Bupropion	Antidepressant	3.6	0	ND	ND	86	
Carbamazepine	Anticonvulsant	0.84	38	27	22	80	<0.5 - 52 ^{e,h}
Carisoprodol	Muscle relaxant	2.5	0	ND	ND	81	
Chlorpheniramine ^{a,c}	Antihistamine	0.94	13	2.4	2.4	220	
Cimetidine ^c	H2-receptor antagonist	5.6	38	<mdl< td=""><td><mdl< td=""><td>100</td><td><0.5 – 202^e</td></mdl<></td></mdl<>	<mdl< td=""><td>100</td><td><0.5 – 202^e</td></mdl<>	100	<0.5 – 202 ^e
Citalopram ^c	Antidepressant	1.3	50	37	14	170	53 ⁱ
Clonidine	Antihypertensive	30	0	ND	ND	87	
Dehydronifedipine	Nifedepine metabolite	4.9	0	ND	ND	78	
Desmethyl-diltiazem ^c	Degradate of diltiazem	2.5	25	48	44	210	
Desvenlafaxine	Antidepressant, venlafaxine metabolite	3.8	88	85	16	87	7.3 – 290 ^{i,j}

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Pharmaceutical	Source or use	MDL (ng/L)	Detection Frequency %	Max (ng/L)	Median (ng/L)	Matrix recovery % (median)	Detected in the UK (ng/L)
Dextromethorphan ^{a,c}	Cough suppressant	1.6	25	6.7	6.0	140	
Diazepam	Benzodiazepine	0.45	63	1.3	1.0	81	0.6– 1.1 ^{f,g}
Diltiazem ^c	Calcium channel blocker	5.1	63	44	9.1	180	<1 – 49 ^e
Diphenhydramine ^a	Antihistamine	2.9	25	6.0	5.6	100	
Erythromycin ^c	Macrolide antibiotic	27	25	180	170	250	<0.5 – 1000 ^{k,l}
Ezetimibe ^c	Cholesterol-reducing agent	13	25	<mdl< td=""><td><mdl< td=""><td>160</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>160</td><td></td></mdl<>	160	
Fadrozole ^b	Aromatase inhibitor	1.5	0	ND	ND	92	
Fenofibrate	H2-receptor antagonist	1.3	0	ND	ND	100	
Fexofenadine	Antihistamine	4.0	100	130	18	90	64 ^j
Fluconazole ^a	Antifungal	36	0	ND	ND	76	
Fluoxetine ^c	Antidepressant	5.4	0	ND	ND	360	$6.2 - 34^{f,m}$
Fluticasone ^c	Synthetic corticosteroid	0.92	63	<mdl< td=""><td><mdl< td=""><td>86</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>86</td><td></td></mdl<>	86	
Glipizide	Antidiabetic	17	0	ND	ND	82	
Glyburide	Antidiabetic	0.79	88	3.1	<mdl< td=""><td>81</td><td></td></mdl<>	81	
Hydrocodone	Opioid, codeine metabolite	2.1	25	39	34	110	
Hydrocortisone	Natural glucocorticoid hormone	29	0	ND	ND	77	
Hydroxyzine	Glucocorticoid hormone	1.5	0	ND	ND	110	
Iminostilbene	Carbamazepine degradate	73	0	ND	ND	98	
Ketoconazole ^c	Antifungal	56	0	ND	ND	430	
Lamivudine ^c	Antiretroviral	3.2	0	ND	ND	160	
Lidocaine ^a	Topical anesthetic	3.1	75	9.6	8.9	84	
Loperamide ^c	Antidiarrheal	5.7	0	ND	ND	420	
Loratadine ^a	Antihistamine	1.4	88	8.5	1.5	120	
Pharmaceutical	Source or use	MDL (ng/L)	Detection	Max	Median	Matrix	Detected in

			Frequency %	(ng/L)	(ng/L)	recovery % (median)	the UK (ng/L)
Lorazepam	Benzodiazepine (anxiolytic)	58	0	ND	ND	84	
Meprobamate	Anxiolytic	17	0	ND	ND	74	
Metaxalone ^b	Muscle relaxant	7.8	0	ND	ND	80	
Metformin	Antidiabetic	6.6	100	1300	630	120	2300 ⁱ
Methadone ^c	Synthetic opioid	3.8	0	ND	ND	200	10 – 18 ^g
Methocarbamol	Muscle relaxant	4.4	25	10	8.7	81	
Methotrexate	Chemotherapy agent	11	0	ND	ND	76	<6.3 ⁿ
Metoprolol ^c	Beta-blocker	14	0	ND	ND	86	<0.5 – 12 ^e
Morphine	Analgesic (opioid)	2.8	30	21	19	84	$0.6 - 36^{f,g}$
Nadolol	Beta-blocker	16	0	ND	ND	85	
Nevirapine ^c	Antiretroviral	3.0	25	<mdl< td=""><td><mdl< td=""><td>81</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>81</td><td></td></mdl<>	81	
Nizatidine ^c	Acid inhibitor (ulcers)	9.5	0	ND	ND	240	
Noreistherone	Oral contraceptive component	2.2	13	<mdl< td=""><td><mdl< td=""><td>85</td><td><10 – 17^s</td></mdl<></td></mdl<>	<mdl< td=""><td>85</td><td><10 – 17^s</td></mdl<>	85	<10 – 17 ^s
Nordiazepam	Benzodiazepine, diazepam metabolite	21	0	ND	ND	82	0.1 – 6.8 ^f
Norverapamil ^c	Verapamil metabolite	1.7	0	ND	ND	400	
Omeprazole ^c	Proton pump inhibitor	2.8	0	ND	ND	260	
Oseltamivir	Antiviral	2.9	38	3.6	<mdl< td=""><td>85</td><td></td></mdl<>	85	
Oxazepam	Benzodiazepine (anxiolytic)	28	0	ND	ND	81	$0.9 - 21^{f}$
Oxycodone	Opioid analgesic	5.0	0	ND	ND	90	$0.4 - 7.1^{f,g}$
Paracetamol ^a	Analgesic	3.6	63	1000	260	88	52 – 2400 ^{d,e}
Paroxetine ^c	Antidepressant	4.1	0	ND	ND	300	
Penciclovir ^c	Antiviral	8.1	0	ND	ND	160	
Pharmaceutical	Source or use	MDL (ng/L)	Detection Frequency %	Max (ng/L)	Median (ng/L)	Matrix recovery % (median)	Detected in the UK (ng/L)

Thiabendazole ^b	Fungicide	0.82	0	ND	ND	83	
Pharmaceutical	Source or use	MDL (ng/L)	Detection Frequency %	Max (ng/L)	Median (ng/L)	Matrix recovery % (median)	Detected in the UK (ng/L)
Theophylline	Diuretic	8.3	0	ND	ND	75	
Temazepam	Benzodiazepine (hypnotic)	9.2	25	<mdl< td=""><td><mdl< td=""><td>81</td><td>1.4 – 78</td></mdl<></td></mdl<>	<mdl< td=""><td>81</td><td>1.4 – 78</td></mdl<>	81	1.4 – 78
Tamoxifen ^c	Cancer treatment	11	0	ND	ND	3300	<10 – 210 ^{k,o}
Sulfamethoxazole	Sulfonamide antibiotic	13	38	<mdl< td=""><td><mdl< td=""><td>80</td><td>1.8 – 8^{e,j}</td></mdl<></td></mdl<>	<mdl< td=""><td>80</td><td>1.8 – 8^{e,j}</td></mdl<>	80	1.8 – 8 ^{e,j}
Sulfamethizole ^b	Sulfonamide antibiotic	21	0	ND	ND	82	
Sulfadimethoxine ^b	Sulfonamide antibiotic	33	0	ND	ND	83	
Sitagliptin	Antihyperglycemic	20	25	36	20	81	
Sertraline ^c	Antidepressant	3.3	0	ND	ND	300	
Ranitidine ^a	Acid inhibitor (ulcers)	38	100	180	72	100	<3 - 73 ^{e,h,q}
Raloxifene	modulator	4.9	0	ND	ND	420	
Quinine ^{a,c}	Antimalarial, flavouring agent	16	50	41	23	140	
Pseudoephedrine ^a	Decongestant	5.5	13	8.5	8.0	81	12 – 17 ^g
Propranolol	Beta blocker	13	50	27	18	110	3.9- 220 ^{k,p}
Propoxyphene	Opioid analgesic	3.4	0	ND	ND	140	9 -680 ^{ĸ,o}
Promethazine ^{a,c}	Antihistamine	10	50	<mdl< td=""><td><mdl< td=""><td>190</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>190</td><td></td></mdl<>	190	
Prednisone	Synthetic corticosteroid	84	0	ND	ND	120	
Prednisolone	Synthetic corticosteroid, prednisone metabolite	75	0	ND	ND	91	
Piperonyl butoxide ^b	Pesticide, lice treatment	1.5	13	2.8	2.8	87	
Phenytoin	Antiepileptic	94	0	ND	ND	78	
Phendimetrazine ^b	Appetite suppressant	16	0	ND	ND	86	
Phenazopyridine ^b	Urinary tract analgesic	2.7	0	ND	ND	84	
Pentoxyfylline ^c	Cardiovascular drug	4.7	10	<mdl< td=""><td><mdl< td=""><td>86</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>86</td><td></td></mdl<>	86	

Tiotropium ^c	Bronchodilator	8.6	0	ND	ND	220		
Tramadol	Opioid analgesic	3.0	50	77	49	90	3.0 – 7700 ^{e,f}	
Triamterene	Diuretic	2.6	25	4.2	<mdl< td=""><td>80</td><td></td></mdl<>	80		
Trimethoprim	Antibiotic	3.8	75	31	22	86	<1.5 – 180 ^{e,r}	
Venlafaxine	Antidepressant	0.90	38	15	12	95	1.1 – 85	
Verapamil ^c	Calcium channel blocker	3.1	0	ND	ND	550		
Warfarin	Anticoagulant	3.0	25	<mdl< td=""><td><mdl< td=""><td>84</td><td></td></mdl<></td></mdl<>	<mdl< td=""><td>84</td><td></td></mdl<>	84		
Wardnin Anticoagulant 3.0 25 <mdl< th=""> <mdl< th=""> 84 % = percentage; ng/L = nanograms per litre; MDL = Method detection limit; ND = Not detected *</mdl<></mdl<>								



Figure 2. A heat map of the mean pharmaceutical concentration at each of the 8 sampling sites along the Rivers Ouse and Foss. Numbers refer to the specific sampling sites listed in Figure 1. Significant differences in concentrations between the River Ouse and Foss were found for the 6 pharmaceuticals that were detected frequently enough to compute a student's t-test, * indicates a p ≤ 0.05 .



Figure 3. A semi-quantitative analysis of PEC performance in the rivers based on the monitoring campaign results. A compound is expected to be detected when the PEC is greater than the respective analytical method detection limit.



Figure 4. PEC range and MEC range for compounds quantified in the River Foss. The worst case PEC is also plotted (open circles) where $F_{excreta} = 1$ and WwTP removal = 0. The MEC range is based on the results from sampling sites 1-3 (Figure 1).



Figure 5. PEC range and MEC range for compounds quantified in the River Ouse. The worst case PEC is also plotted (open circles) where $F_{excreta} = 1$ and WwTP removal = 0.The MEC range is based on the results from sites 5-7 (Figure 1) and corrected for the upstream contributions.



the River Ouse. Ranks are presented by decreasing risk, where rank 1 corresponds to highest risk. River Foss. (B) The range of possible ranks resulting from risk quotients calculated using MECs or PECs in