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Article:

Mawer, DPC, Eyre, DW, Griffiths, D et al. (7 more authors) (2017) Contribution to Clostridium difficile transmission of symptomatic patients with toxigenic strains who are fecal toxin negative. *Clinical Infectious Diseases*, 64 (9). pp. 1163-1170. ISSN 1058-4838

<https://doi.org/10.1093/cid/cix079>

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1 Contribution to *Clostridium difficile* transmission of symptomatic
2 patients with toxigenic strains who are fecal toxin negative

3

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16

17

18 Key words

19 *Clostridium difficile*; infection; fecal toxin; transmission

20

21 Running title

22 *C. difficile* fecal toxin status & transmission

23

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[Article summary point](#)

Using whole-genome sequencing, in two UK hospitals, patients with diarrhea, toxigenic *Clostridium difficile*, but a negative fecal toxin result, were potential sources for 3% of infections; toxin-positive cases were potential sources for 10%, and another 6% were linked to both groups.

55 Abstract

56

57 Background

58 The role of symptomatic patients who are toxigenic strain-positive (TS+) but fecal toxin-
59 negative (FT-) in transmission of *Clostridium difficile* is currently unknown.

60

61 Methods

62 We investigated the contribution of symptomatic TS+/FT- and TS+/FT+ patients in *C. difficile*
63 transmission in two UK regions. From two-step testing, all glutamate dehydrogenase (GDH)-
64 positive specimens, regardless of fecal toxin result, from Oxford (April2012-April2013) and
65 Leeds (July2012-April2013) microbiology laboratories underwent culture and whole-genome
66 sequencing (WGS), using WGS to identify toxigenic strains. Plausible sources for each
67 TS+/FT+ case, including TS+/FT- and TS+/FT+ patients, were determined using WGS, with
68 and without hospital admission data.

69

70 Results

71 1447/12772(11%) fecal samples were GDH-positive, 866/1447(60%) contained toxigenic *C.*
72 *difficile* and fecal toxin was detected in 511/866(59%), representing 235 Leeds and 191
73 Oxford TS+/FT+ cases. TS+/FT+ cases were three times more likely to be plausibly acquired
74 from a previous TS+/FT+ case than a TS+/FT- patient. 51(19%) of 265 TS+/FT+ cases
75 diagnosed >3 months into the study were genetically-related (≤ 2 single nucleotide
76 polymorphisms) to ≥ 1 previous TS+/FT+ case or TS+/FT- patient: 27(10%) to only TS+/FT+
77 cases, 9(3%) to only TS+/FT- patients, and 15(6%) to both. Only 10/265(4%) were
78 genetically-related to a previous TS+/FT+ or TS+/FT- patient and shared the same ward
79 simultaneously or within 28 days.

80

81 Conclusions

82 Symptomatic TS+/FT- patients were a source of *C. difficile* transmission, although they
83 accounted for less onward transmission than TS+/FT+ cases. Although transmission from
84 symptomatic patients with either fecal toxin status accounted for a low overall proportion
85 of new cases, both groups should be infection control targets.

86 Background

87 *Clostridium difficile* infection (CDI) remains a significant concern for patients and healthcare
88 providers, despite recent falls in incidence in some settings, including the UK.[1] Three UK
89 studies using whole-genome sequencing (WGS) have shown, in endemic settings with
90 routine infection control policies, only a minority of cases are acquired from other, known,
91 cases: 35% of cases in Oxford[2] and Leeds[3], and 37% of ribotype-027 cases in Liverpool[4]
92 were genetically-linked to a previous case. Only a subset of these cases also shared time on
93 the same hospital ward. Studies using other genotyping techniques have found similar
94 results.[5-7] Such findings question the sources of *C. difficile* responsible for most CDIs.

95
96 While hospitalized asymptotically colonized patients are a potential source,[7-9] another
97 group of patients with enhanced potential to transmit *C. difficile* are symptomatic patients
98 who are toxigenic-strain positive (TS+), but fecal toxin negative (FT-). These patients are
99 identified by two-step algorithms for CDI diagnosis.[10] An initial screen (e.g. glutamate
100 dehydrogenase (GDH) enzyme immunoassay (EIA), or toxin gene nucleic acid amplification
101 test [NAAT]) detects the presence of *C. difficile*; the second confirmatory step detects fecal
102 toxin using either EIA or a cell cytotoxin assay (CCT). In the UK TS+/FT- patients are usually
103 regarded as being colonized with *C. difficile* but not infected, based on a large multi-center
104 prospective study showing only patients with detectable fecal toxin had adverse
105 outcomes.[11] However, outside the UK, such patients, typically identified with NAATs, are
106 often,[12] but not universally,[13] regarded as having CDI, and NAAT testing has been
107 recommended in some guidelines.[14] Resolving the disease state of TS+/FT- patients is not
108 a focus of this study; instead we investigated their contribution to onward transmission of *C.*
109 *difficile*.

110
111 We undertook WGS of consecutive *C. difficile* GDH-positive fecal samples, irrespective of the
112 subsequent fecal toxin assay result, in two UK centers, over 9-12 months. WGS, combined
113 with hospital admission and ward movement data, were used to assess the contribution of
114 *C. difficile* TS+/FT- and TS+/FT+ patients to onward transmission.

115

116 Methods

117 Samples and setting

118 Consecutive hospital and community samples submitted for *C. difficile* diagnostic testing
119 were obtained from the microbiology laboratories of 2 UK teaching hospitals following the
120 introduction of two-step testing: Leeds Teaching Hospitals, serving Leeds (population
121 750,000, 07-July-2012 to 06-April-2013), and Oxford University Hospitals, serving
122 Oxfordshire (population 600,000, 01-April-2012 to 31-March-2013). In Leeds and Oxford,
123 repeat samples from the same patient ≤ 14 and ≤ 28 days, respectively, following a toxin-
124 positive sample were not routinely processed. Patient admissions and hospital ward
125 movements were obtained from hospital administration systems. Inclusion of community
126 samples allowed cases diagnosed in the community, but potentially acquired in hospital, to
127 be identified.

128

129 In Leeds, any patient with ≥ 1 episode of unexplained diarrhea was isolated and a fecal
130 sample sent for *C. difficile* testing. TS+/FT+ cases were isolated for the duration of hospital
131 admission. Ward staff could isolate TS+/FT- patients if they were considered a transmission
132 risk. In Oxford, patients with unexplained diarrhea (≥ 3 unformed stools in 24 hours) were
133 isolated and treated empirically with oral vancomycin. TS+/FT+ cases remained isolated
134 until 48 hours following resolution of diarrhea. Treatment and isolation were discontinued
135 in TS+/FT- patients unless clinical suspicion of CDI remained high.

136

137 Diagnostic testing and WGS

138 Leeds samples were tested with GDH EIA, *C. diff* Chek (Techlab, Blacksburg, VA), and when
139 GDH-positive an in-house cell cytotoxicity assay, and Oxford samples with Premier *C. difficile*
140 GDH and GDH-positive samples with Premier Toxins A&B EIA (Meridian Bioscience,
141 Cincinnati, OH). At both centers, GDH-positive samples were cultured as described
142 previously[15] and whole-genome sequenced using Illumina technology. In Leeds, isolates
143 were confirmed as *C. difficile* with MALDI-TOF mass-spectrometry; in Oxford WGS was used.
144 Sequences were mapped to the 630 reference genome[16], and assembled *de novo*[17] (see
145 Supplementary Methods for details). Multi-locus sequence types, STs,[15] were determined
146 *in silico*.

147

148 Toxigenic strains were identified using BLAST searches of *de novo* assemblies (≥ 1000
149 nucleotide identities with toxin A or B genes). Non-toxigenic strains were excluded ($n=249$,
150 most common STs ST15($n=66,27\%$), ST26($n=66,27\%$), ST7($n=51,20\%$), and ST3($n=11,4\%$); the
151 remainder were recognized non-toxigenic STs).

152

153 Definitions

154 Patients with toxigenic *C. difficile* were classified according to fecal toxin result: as TS+/FT+
155 and TS+/FT-. In patients diagnosed with more than one *C. difficile* strain, as defined by WGS
156 (see below), each was considered separately. Some patients had several samples with the
157 same strain, and could be consistently fecal toxin-negative, consistently toxin-positive, or
158 have both fecal toxin-negative and toxin-positive samples. Each TS+/FT+ CDI's origin was
159 determined using standard surveillance definitions.[18] Cases were defined as healthcare-
160 associated if sampled >48 hours after admission or discharged within ≤ 4 weeks, as
161 indeterminate if discharged 4-12 weeks previously, and as community-associated if
162 discharged >12 weeks prior to sampling, or without any hospital admission.

163

164 Analysis

165 Single nucleotide polymorphisms (SNPs) between sequences were determined from
166 maximum likelihood phylogenies constructed with phyML[19] after correction for
167 recombination with ClonalFrameML.[20] Sequences related to a previous sequence within
168 ≤ 2 SNPs were considered consistent with plausible direct transmission; ≤ 2 SNPs is expected
169 between transmitted strains obtained ≤ 123 days apart.[2] Results for sequences related to
170 previous sequences within varying thresholds (0-10 SNPs) were generated as a sensitivity
171 analysis. In patients with multiple samples, sequences >10 SNPs different to a previous
172 sequence from the same patient were considered to represent acquisition of a new strain;
173 10 SNPs is considerably more variation than would be expected from within-host diversity
174 and mutation over the one year study period.[2]

175

176 Where the only possible genetically-related sources of a TS+/FT+ case were TS+/FT-
177 patients, the origin was attributed to TS+/FT- patients; similarly, if all possible genetically-

178 related sources were TS+/FT+ cases, the origin was attributed to TS+/FT+ cases. Where a
179 TS+/FT+ case was genetically-linked to either a TS+ patient with both fecal toxin-positive
180 and toxin-negative samples, or several patients including ≥ 1 TS+/FT+ case and ≥ 1 TS+/FT-
181 patient, the origin was denoted as either a TS+/FT+ case or TS+/FT- patient.

182

183 Patients with toxigenic *C. difficile* who shared time on the same ward following the
184 diagnosis of the first patient and before the diagnosis of the second were considered to
185 have had ward contact. Patients admitted to the same ward, but up to 28 days apart, were
186 considered related by possible ward contamination if the first patient was diagnosed before
187 their ward discharge, and the second patient following their admission to the same ward.[5]
188 Patients who shared time in the same hospital, but had no ward or ward contamination
189 contact, were considered to have hospital contact. A sensitivity analysis assumed ward
190 contamination persisted for 365 days.

191

192 Logistic regression was used to test for associations between ST and the proportion of
193 TS+/FT+ cases genetically-related to a previous TS+/FT+ case or TS+/FT- patient, for the 9
194 most common STs (all with ≥ 10 cases).

195

196 Ethics

197 The study was approved by the Berkshire Research Ethics Committee (10/H0505/83) and
198 the Health Research Authority (8-05(e)2010).

199

200 Results

201 8068 hospital and community samples were submitted for *C. difficile* testing in Leeds, and
202 4704 samples in Oxford. 771(10%) and 637(14%) samples were GDH-positive respectively,
203 and, of these, 488(63%) and 372(58%) contained toxigenic *C. difficile* by WGS (Figure 1).
204 Leeds samples were obtained from 367 patients (220 female,60%), median (interquartile
205 range, IQR) 72(52-82) years old, representing 382 genetically distinct
206 infections/colonizations, and Oxfordshire samples from 297 patients (167 female,56%),
207 78(62-86) years old, 302 genetically distinct infections/colonizations.

208

209 In both laboratories, 59% of samples containing toxigenic *C. difficile* had fecal toxin detected
210 despite using different assays, EIA in Oxford (218/372) and CCT in Leeds (289/488). These
211 samples represented 235 distinct TS+/FT+ cases in Leeds, with 3.7 healthcare-
212 associated/indeterminate cases per 10000 bed-days and 7.9 community-associated cases
213 per 100000 person-years, and 191 distinct TS+/FT+ cases in Oxfordshire, 3.2/10000 bed-
214 days and 7.0/100000 person-years, respectively (Figure 1).

215

216 There was considerable genetic diversity amongst the *C. difficile* causing the 426 TS+/FT+
217 cases, with 52 different STs identified. The 10 most frequently isolated STs (common
218 ribotype equivalents) accounted for 285(67%) of cases, and were (in rank order)
219 ST2(014/020), ST8(002), ST6(005), ST11(078), ST10(015), ST5(023/069), ST44(015),
220 ST3(001/072), ST14(014), ST17(018). The epidemic ST1(027/NAP1) strain was only found in
221 three (Leeds) cases.

222

223 Genetic relationships between infections/colonizations

224 Samples were compared with all prior samples from the same center over the study
225 periods, but potential sources were sought only for new TS+/FT+ infections from 3 months
226 into the study at each center (Leeds n=142, Oxfordshire n=123), to ensure sufficient time for
227 their possible sources to have been sampled. Using a threshold of ≤ 2 SNPs to determine
228 genetic relatedness, overall 51/265(19.2%, 95%CI, 14.7-24.5%) TS+/FT+ cases were
229 genetically-related to ≥ 1 sequenced previous TS+/FT+ case or TS+/FT- patient (Table 1).
230 9/265(3.4%, 1.6-6.3%) of TS+/FT+ cases were genetically linked only to TS+/FT- patients and
231 not to previous TS+/FT+ cases. In contrast, 27/265(10.2%, 6.8-14.5%) TS+/FT+ cases were
232 genetically linked to other TS+/FT+ cases, and 15/265(5.7%, 3.2-9.2%) to both TS+/FT+ cases
233 and TS+/FT- patients. There was no evidence of a difference in sources between Leeds and
234 Oxford (Table 1; exact p=0.27).

235

236 Considering the source of *C. difficile* for all patients, TS+/FT- patients as well as TS+/FT+
237 cases, results were similar (Table S1; exact p=0.85 comparing all patients vs. TS+/FT+ cases
238 alone): 75/433(17%) patients could be linked to a previously sequenced TS+/FT+ case or

239 TS+/FT- patient, 16(4%) to only TS+/FT- patients, 36(8%) to only previous TS+/FT+ cases and
240 23(5%) to both.

241

242 There were 13 ST44 infections, none of which were genetically-related to a prior TS+/FT+
243 case, the remaining 8 most common STs were compared with all other STs as the reference
244 group. Within the limits of the relatively small numbers of TS+/FT+ cases within each ST,
245 there was no evidence that CDI caused by any of these STs were more or less likely, to be
246 genetically-related to a previous TS+/FT+ case or TS+/FT- patient ($p \geq 0.18$; Table 2), or that
247 CDI source was associated with patient age, sex or healthcare/community-associated
248 disease (Table 3).

249

250 Over the whole study period at both centers, considering all 684 TS+/FT+ cases and TS+/FT-
251 patients, 535 were not related to any other TS+/FT+ case or TS+/FT- patient within ≤ 2 SNPs.
252 The remaining 149 TS+/FT+ cases and TS+/FT- patients were clustered: sequences included
253 in a cluster were related to ≥ 1 other sequence within ≤ 2 SNPs in the cluster, but not
254 necessarily to all of them. Most clusters contained 2 or 3 patients; 14(9%) patients were in
255 clusters consisting of exclusively TS+/FT- patients, 45(30%) were in exclusively TS+/FT+
256 clusters, and 90(60%) were in clusters with both TS+/FT- patients and TS+/FT+ cases (Figure
257 3).

258

259 [Epidemiological relationships between genetically-related infections/colonizations](#)

260 Only a subset of TS+/FT+ cases and plausible TS+/FT+ or TS+/FT- sources related within ≤ 2
261 SNPs shared a hospital-based epidemiological link. Considering all 265 TS+/FT+ cases from
262 both Leeds and Oxfordshire from 3 months into the study, 27(10%) were genetically-related
263 to only previous TS+/FT+ cases. However, only 6(2%) were genetically-related and shared
264 time on the same ward. A further 4(2%) were genetically-related and were inpatients on the
265 same ward at different times within 28 days. 8(3%) were not admitted to the same ward
266 within 28 days, but were admitted to the same hospital at the same time (Table 1).

267

268 Another 9(3%) TS+/FT+ cases were genetically-related to only previously TS+/FT- patients:
269 5(2%) sharing time on a ward, 1(0.4%) the same ward at different times within 28 days, and

270 1(0.4%) time in the same hospital as above. There was a trend towards potential TS+/FT-
271 sources being more likely to share time on the same ward as the subsequent TS+/FT+ case,
272 compared with potential TS+/FT+ sources (5/9 vs. 6/27, exact p=0.10). An additional 15(6%)
273 TS+/FT+ cases were genetically-related to both a TS+/FT- patient and a TS+/FT+ case, but 14
274 had no hospital-based links with the genetically-related sources, suggesting these patients
275 may share a common indirect source rather than direct hospital-based contact. No
276 additional epidemiological links between genetically-related TS+/FT+ cases and TS+/FT+
277 cases or TS+/FT- patients were identified if ward contamination could persist for up to 365
278 days.

279

280 To test the robustness of our observations to the SNP threshold used to define plausible
281 direct transmission, the number of TS+/FT+ cases genetically-related to a previous TS+/FT+
282 case or TS+/FT- patient within varying SNP thresholds from 0 to 10, and any associated
283 hospital-based epidemiological links, were determined (Figure 3). As expected, as the
284 number of SNPs used to define plausible direct transmission increased, the percentage of
285 TS+/FT+ cases genetically-related to a previous TS+/FT+ or TS+/FT- patient increased.
286 However, the number of TS+/FT+ cases genetically-related and with plausible
287 epidemiological contact, i.e. sharing hospital wards, remained relatively constant from 2
288 SNPs onwards, supporting the 2 SNP threshold used for the main analysis.

289

290 Discussion

291 We used WGS and ward admission data to investigate the proportion of CDI cases
292 potentially acquired from symptomatic patients with toxigenic *C. difficile*, but with no
293 detectable fecal toxin. TS+/FT+ CDI cases were three times more likely to be genetically-
294 related to a previous TS+/FT+ case (27/265) than a TS+/FT- patient (9/265). Considering the
295 subset of potential sources that also shared time on the same ward, or were admitted to
296 the same ward within 28 days, i.e. the most probable of the genetically-plausible
297 transmission events, CDI cases were 1.7 times more likely to be related to a previous
298 TS+/FT+ case compared with a TS+/FT- patient (10/265 vs. 6/265). However, As there were
299 also 1.7 times as many TS+/FT+ as TS+/FT- patients in the study, making linkage to a
300 previous TS+/FT+ case 1.7 times more likely than to a TS+/FT- patient based on the relative

301 ~~frequencies of TS+/FT+ and TS+/FT- patients alone. Therefore, the rate of transmission, on~~
302 ~~a per patient basis, from each TS+/FT+ or TS+/FT- patient is likely to be very similar. the rate~~
303 ~~of transmission from each TS+/FT+ or TS+/FT- patient is likely to be very similar.~~ By contrast,
304 asymptotically colonized patients are likely less infectious. Using national databases and
305 a transmission model, individual hospitalized CDI cases have been estimated to transmit *C.*
306 *difficile* at a rate 15 (95%CI 7.2-32) times that of hospitalized asymptotically colonized
307 patients.[21] However, as asymptomatic carriage is more common than CDI (e.g. 8-fold in
308 hospitals[9]), colonized patients, as a group, could still account for a substantial amount of
309 transmission. In a Canadian study, isolation of all asymptotically colonized patients
310 reduced CDI incidence by 62% compared with historic controls.[9]

311
312 The overall number of our TS+/FT+ CDI cases potentially attributable to the combination of
313 TS+/FT+ cases and TS+/FT- patients with diarrhea was low: 19% of TS+/FT+ CDI cases were
314 genetically-related to a previous TS+/FT+ or TS+/FT- patient, only 6% also shared a hospital
315 ward at the same time or within 28 days, and only 10% had any form of hospital contact.
316 This supports previous WGS-based studies, at both our hospitals[2,3] and others[4], that
317 found that only a minority of CDIs are acquired from other cases in endemic settings. The
318 proportion in the present study is lower than the 35-37% identified previously. The most
319 likely explanation is the very small number of infections with the epidemic ST1(027/NAP1)
320 strain, reflecting falling UK incidence[22,23], and the burden of transmissions attributable to
321 ST1 in previous studies.[3]

322
323 Our study has several limitations. Only patients with diarrhea were sampled, and at the
324 discretion of individual practitioners. However, the ratio of toxin-positive stools sequenced
325 to samples tested was 3.6%(289/8068) in Leeds, and 4.6%(218/4704) in Oxford, suggesting
326 rates of testing were high, including compared with the UK average from 2008 of 6.45%,
327 when testing was principally based on toxin detection.[24] Of those tested, some patients
328 with *C. difficile* will have been missed by the GDH assay (sensitivity 92.3-97.1%[11,25]). In
329 addition, 2.6% of isolates failed WGS and were excluded. We therefore may have missed
330 some links between TS+/FT- or TS+/FT+ patients and TS+/FT+ CDI cases, modestly
331 underestimating the frequency with which this occurs.[5] However, if cases were missed at

332 random, we believe the relative amount of transmission attributable to TS+/FT+ cases and
333 TS+/FT- patients has been robustly estimated. We did not gather data on factors that might
334 influence a TS+/FT- patient's potential to transmit *C. difficile*, including duration and severity
335 of diarrhea, antibiotic exposure, or the timing and duration of isolation. In addition,
336 systematic serial sampling was not undertaken to allow an assessment of the duration of
337 detectable *C. difficile*. Our study was performed in a setting where the majority of CDI arises
338 from a diverse range of endemic strains; findings may vary in higher incidence settings,
339 including where the epidemic ST1(027/NAP1) strain dominates.

340

341 Despite these limitations, we demonstrate that patients with toxigenic *C. difficile* without
342 detected fecal toxin account for a quarter or more of potential within hospital transmission
343 events from symptomatic patients. More intensive infection control interventions around
344 such cases, including routine isolation, should be considered to mitigate transmission risk.
345 Compared with asymptomatically colonized patients, TS+/FT- patients represent a good
346 initial target for expanding infection control efforts, as they are less numerous, and, as
347 discussed above, appear more infectious[21] on a per patient basis. However, ultimately if
348 the findings of [9] can be replicated, isolation of asymptomatically colonized patients, who
349 are each less infectious, but more numerous, may result in greater reductions in
350 transmission. Substantially greater resource requirements limit the later approach. Some
351 GDH-positive fecal toxin-negative patients may carry non-toxigenic *C. difficile* and not pose
352 an infection control risk. Patients with toxigenic *C. difficile* could be identified by screening
353 with a toxin gene NAAT, or using a three-step strategy (GDH-positive, fecal toxin-negative
354 samples tested with a toxin gene NAAT).

355

356 The results of this and previous studies in both Oxford and Leeds suggest CDI cases, and also
357 symptomatic patients with toxigenic *C. difficile* with a negative fecal toxin result, are not
358 sources for the majority of CDI. Major unanswered questions remain, including what
359 proportion of CDI cases can be explained by healthcare-associated and community contact
360 with asymptomatically colonized people, and the extent to which other possible sources
361 including food[26,27] and the environment[28] contribute to CDI. In addition to reducing
362 the risk of CDI through antimicrobial stewardship,[23] understanding the relative

363 importance of each of these reservoirs across a range of settings is required to develop
364 rational control policies and reduce the incidence of CDI. Meanwhile, efforts to reduce
365 hospital transmission from symptomatic patients with toxigenic *C. difficile* with a negative
366 fecal toxin result should be implemented.

367

368 [Funding](#)

369 This study was supported by the UK Clinical Research Collaboration (Wellcome Trust [grant
370 087646/Z/08/Z]; Medical Research Council [grant G0800778]; and the National Institute for
371 Health Research); and the NIHR Oxford Biomedical Research Centre. DWC and TEAP are
372 NIHR senior investigators. DWE is a NIHR clinical lecturer.

373

374 [Acknowledgements](#)

375 The authors thank Claire Berry and Faye Pinker for their help collecting hospital admissions
376 data in Leeds.

377

378 [Potential conflicts of interest](#)

379 M.H.W has received consulting fees from Actelion, Astellas, MedImmune, Merck, Pfizer,
380 Sanofi-Pasteur, Seres, Summit, and Synthetic Biologics; lecture fees from Alere, Astellas,
381 Merck & Pfizer; and grant support from Actelion, Astellas, bioMerieux, Da Volterra, Merck
382 and Summit.

383

384 D.P.C.M, D.W.E, D.G, W.N.F, J.S.H.M, T.P.Q, T.E.A.P, D.W.C and A.S.W all have no conflicts of
385 interest to declare.

386

387 [Data deposition](#)

388 Sequences generated during this study can be found on the NCBI short read archive under
389 BioProject PRJNA327723.

390

391

392

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Possible transmission source	Genetically linked ≤ 2 SNPs (% n)	Epidemiological links between genetically linked cases, No. (% n, % genetically linked)		
		Shared time on same ward	Shared same ward within 28 days	Shared time in same hospital only
Leeds (CDI cases, n = 142)				
Prior TS+/FT+ cases	17 (12)	2 (1, 12)	2 (1, 12)	7 (5, 41)
TS+/FT- patients	5 (4)	2 (1, 40)	1 (1, 20)	1 (1, 20)
Both	11 (8)	0 (0, 0)	0 (0, 0)	1 (1, 9)
Total	33 (23)	4 (3, 12)	3 (2, 9)	9 (6, 27)
Oxford (CDI cases, n = 123)				
Prior TS+/FT+ cases	10 (8)	4 (3, 40)	2 (2, 20)	1 (1, 10)
TS+/FT- patients	4 (3)	3 (2, 75)	0 (0, 0)	0 (0, 0)
Both	4 (3)	0 (0, 0)	0 (0, 0)	0 (0, 0)
Total	18 (15)	7 (6, 39)	2 (2, 11)	1 (1, 6)
Combined (CDI cases, n = 265)				
Prior TS+/FT+ cases	27 (10)	6 (2, 22)	4 (2, 15)	8 (3, 30)
TS+/FT- patients	9 (3)	5 (2, 56)	1 (1, 11)	1 (1, 11)
Both	15 (6)	0 (0, 0)	0 (0, 0)	1 (1, 7)
Total	51 (19)	11 (4, 22)	5 (2, 10)	10 (4, 20)

479

480 **Table 1. Proportion of toxigenic strain-positive, fecal toxin-positive (TS+/FT+) CDI cases**481 **genetically (≤ 2 SNPs) and epidemiologically related to prior TS+/FT+ cases and TS+/FT-**482 **patients.**

483

484
485

Genetically-related to prior TS+/FT+ case or TS+/FT- patient					
ST	n	Total (% n)	Odds ratio (95% CI)	P value	
All other STs	114	24 (21)	1	-	
2	33	9 (27)	1.41 (0.58, 3.42)	0.45	
5	17	4 (24)	1.15 (0.34, 3.86)	0.82	
6	23	2 (9)	0.36 (0.08, 1.63)	0.18	
8	21	3 (14)	0.63 (0.17, 2.30)	0.48	
10	14	1 (7)	0.29 (0.04, 2.32)	0.24	
11	20	5 (25)	1.25 (0.41, 3.78)	0.69	
14	10	3 (30)	1.61 (0.39, 6.69)	0.51	
44	13	0 (0)	-	-	

486
487

488 **Table 2. Association between ST and proportion of CDI cases genetically-related to prior**
489 **TS+/FT+ cases and TS+/FT- patients.** Each ST in the table was compared to all other STs (the
490 reference group) by logistic regression.

491
492

	No genetically linked source	TS+/FT- source	TS+/FT+ source	Both	p value	p value, any source vs no genetically- linked source
Classification (row %)					0.99	0.83
Community-associated	53 (83%)	1 (2%)	7 (11%)	3 (5%)		
Indeterminate	22 (85%)	1 (4%)	2 (8%)	1 (3%)		
Healthcare-associated	139 (79%)	7 (4%)	18 (10%)	11 (6%)		
Age					0.76	0.59
Median	75	82	79	78		
IQR	54 - 83	69 - 86	24 - 85	58 - 84		
Sex (row %)					0.5	0.35
Female	115 (79%)	4 (3%)	17 (12%)	10 (7%)		
Male	98 (84%)	5 (4%)	9 (8%)	5 (4%)		

494

495 **Table 3. Patient demographics according to CDI source (n=265).** Age and sex were not
496 recorded for 2 patients. Exact p values are shown for classification and sex; p values for age
497 were calculated with the Kruskal-Wallis rank test.

498

499

500 [Figure legends](#)

501

502 **Figure 1. Samples and patient demographics for Leeds (panel A) and Oxfordshire (panel B).**

503 Each percentage uses the row above as denominator. Distinct infection is one >10 SNPs
504 distinct to any previous infection in the same patient. HA, healthcare-associated. CA,
505 community-associated. MALDI-TOF MS, matrix assisted laser desorption time of flight mass
506 spectrometry. Age and sex were not recorded for 3 Oxfordshire patients.

507

508 **Figure 2. Numbers of patients in clusters related within ≤ 2 SNPs.** Clusters consisting
509 exclusively of toxigenic strain-positive, fecal toxin-negative (TS+/FT-) patients are shown in
510 blue, clusters consisting exclusively of TS+/FT+ cases in red, and clusters with both TS+/FT-
511 patients and TS+/FT+ cases in orange.

512

513 **Figure 3. Proportion of Leeds and Oxfordshire CDI cases genetically-related to a previous**
514 **toxigenic strain-positive, fecal toxin-positive (TS+/FT+) case or TS+/FT- patient within**
515 **varying SNP thresholds.** Bars are shaded according to the fecal toxin status of the
516 genetically-related potential sources of infection.

517